Original Article Association of interleukin-6 polymorphisms with susceptibility to hepatocellular carcinoma

Xiaohuan Zheng¹, Cuiping Han¹, Rong Shan^{1,2}, Haitao Zhang¹, Zhaomin Zheng¹, Yuanshui Liu¹, Aiguang Wang¹

¹Department of Oncology, Qianfoshan Hospital Affiliated to Shandong University, Jinan, Shandong, China; ²Department of Function, Jinan Infectious Disease Hospital, Jinan, Shandong, China

Received January 1, 2015; Accepted April 14, 2015; Epub April 15, 2015; Published April 30, 2015

Abstract: Target: Our study was to investigate the effects of interleukin-6 (*IL*-6) polymorphisms (rs2069837 and rs17147230) on the risk for hepatocellular carcinoma (HCC). Methods: A total of 226 HCC cases and 220 healthy controls were admitted into the study and genomic DNA was extracted from the peripheral blood. The genotyping was conducted by the method of polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP). Odds ratio (OR) and 95% confidence interval (Cl) were used to evaluate the relationship of *IL*-6 rs2069837 and rs17147230 polymorphisms with HCC susceptibility. Results: The frequency of GG genotype of rs2069837 was higher in HCC patients, compared with controls (P < 0.05). Moreover, the results indicated that GG genotype was related with increased risk for HCC (OR = 2.303, 95% Cl = 1.056-5.025). Similarly, the risk for G allele carriers was higher than that of A allele (OR = 1.392, 95% Cl = 1.046-1.852). For rs17147230, TT genotype showed strong effect on HCC susceptibility (OR = 2.089, 95% Cl = 1.135-3.845) and T allele appeared to be a risk factor for HCC (OR = 1.326, 95% Cl = 1.010-1.740). Further analysis showed that G-T haplotype was associated with increased risk for HCC (OR = 3.125, 95% Cl = 1.845-5.294, P = 0.000). Conclusion: *IL*-6 rs2069837 as well as rs17147230 were associated with HCC susceptibility. In addition, G-T haplotype also served as a genetic-susceptibility factor for HCC.

Keywords: Interleukin-6, hepatocellular carcinoma, polymorphisms

Introduction

Primary liver cancer (PLC) commonly originates from hepatic parenchymal cells or intrahepatic biliary epithelial cells, which includes hepatocellular carcinoma (HCC), intrahepatic cholangiocarcinoma (ICC) and the mixed type of both. In most cases, PLC refers to HCC, because the majority of liver cancers occur in hepatocytes [1-3].

Prevalent in global scale, the incidence of HCC ranks the sixth among all the malignant tumors with about 620,000 new cases every year. In addition, ranking only second to lung cancer and gastric cancer, HCC becomes the third cause of death with the death toll of 600,000 annually. More seriously, up to 55% of new HCC cases (more than 300,000 cases) worldwide occur in China and over 350,000 cases die of HCC each year, which severely threatens human health and life [4, 5]. Multiple factors

may be involved in the occurrence of HCC, such as virus infection including hepatitis b virus and hepatitis c virus, hepatic cirrhosis, aflatoxin, smoking, drinking and genes [6]. It has been demonstrated that genetic factor could affect individual susceptibility to HCC.

Interleukin-6 (*IL*-6), a multi-functional cytokine, has been intensively studied in recent years. *IL*-6 gene locates on short arm of chromosome 7 and consists of 5 exons and 4 introns [7]. Up-regulation level of *IL*-6 has been observed in serum of HCC patients, which suggested that *IL*-6 might be related with the risk of HCC [8-11].

Moreover, studies have showed that the polymorphisms in promoter region of *IL*-6 involved in the pathogenesis of several diseases [12-14]. However, there were few studies about the relationship of *IL*-6 rs2069837 and rs17147230 polymorphisms with HCC risk. So we adopted the case-control design and explored the cor-

Table 1. Primer sequence of rs2069837 and rs17147230

Primer	Forward/Reverse	Primer sequence
rs2069837	Forward	5'-CTTCCTGCT GGAACATTCTATGGC-3'
	Reverse	5'-TTTCTGCCAGTGCCTCTTTGC-3'
rs17147230	Forward	5'-AAAAGGGCAAGGAAGGGAGGTA-3'
	Reverse	5'-CACGA GTCATTTGAGCCATCTTTG-3'

Table 2. Genotype and allele distribution of *IL*-6 rs2069837 andrs17147230

Genotype/allele	Case (%)	Control (%)	X ²	Р	OR (95% CI)
rs2069837					
AA	92/40.7	111/50.5	-	-	1.00
AG	113/50	98/44.5	2.806	0.094	1.391 (0.945-2.048)
GG	21/9.3	11/5	4.565	0.033	2.303 (1.056-5.025)
А	297/65.7	320/72.7	-	-	1.00
G	155/34.3	120/27.3	5.151	0.023	1.392 (1.046-1.852)
rs17147230					
AA	80/35.4	90/44.5	-	-	1.00
AT	107/47.3	109/49.6	0.234	0.629	1.104 (0.739-1.651)
TT	39/17.3	21/5.9	5.717	0.017	2.089 (1.135-3.845)
А	267/59.1	289/65.7	-	-	1.00
Т	185/40.9	151/34.3	4.150	0.042	1.326 (1.010-1.740)

relation of rs2069837 and rs17147230 polymorphisms and HCC susceptibility.

Materials and methods

Study objects

The age of 226 HCC patients including 140 men and 86 women ranged from 41 to 70, and the mean age was 57.6 \pm 7.7. All the patients were confirmed with histopathological examination in our Hospital. Meantime, 220 unrelated healthy people aged from 32 to 76 (mean age 55.8 \pm 7.2) were enrolled as controls, with normal blood routine, no tumors and hereditary diseases.

Genomic DNA extraction and genotyping

3 ml of fasting venous blood extracted from the subjects were shaken up in the EDTA anticoagulant tube and temporarily stored at -20°C in the fridge. Polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) method was adopted to detect rs2069837 and rs17147230 polymorphisms.

The primer sequence was shown in Table 1. PCR reaction mixture included ddH_0 40.5 μ l,

10× PCR Buffer (containing 1.5 mmol/L Mg²⁺) 5 μ l, 10 mmol/L dNTP 1 µl, Taq enzyme 0.5 μ l (5 u/ μ L), forward primer 1 μ l (20 μ mol/L), reverse primer 1 µl (20 µ mol/L), DNA template 1 µl (500 mg). The PCR reaction was conducted according to the following conditions: initial denaturation at 94°C for 2 min, 30 cycles of denaturation at 94°C for 40 s, annealing at 61.2°C for 40 s, annealing at 61°C for 40 s and extension at 72°C for 40 s; finally extension at 72°C for 10 min. 3 µL of PCR amplification products was detected by agarose gel electrophoresis.

The PCR products were mixed with 10× Buffer 0.5 μ L, Mbi I restriction

enzyme (Fermentas, Burlington, Canada) 0.5 μ L and ultrapure water 4 μ L. Then the digestion mixture was processed under 37°C for 6 h. Finally, the digestion products were observed and photographed with MKodak gel imaging analysis system. For genotyping analysis, we determined AA, AG and GG genotypes within rs2069837, and AA, AT and TT genotypes within in rs17147230.

Statistical analysis

Statistical data were analyzed with SPSS18.0 software. Odds ratio (OR) with 95% confidence interval (95% CI), calculated by Chi-square test was applied to estimate the association of *IL*-6 polymorphisms and HCC susceptibility. The significance level was set at P < 0.05.

Results

Correlation analysis

For rs2069837, the frequency of GG genotype in case group significantly differed from that of control group (9.3% vs. 5%, P < 0.05). The results showed that individuals with GG genotype suffered from higher risk of HCC, compared with AA genotype (OR = 2.303, 95% CI =

	1 11				
Haplotype Locus	Case (n)	Control (n)	X ²	Р	OR (95% CI)
A-A	175	191	-	-	1.00
A-T	122	129	0.037	0.847	1.032 (0.748-1.424)
G-A	92	98	0.018	0.892	1.025 (0.721-1.455)
G-T	63	22	19.149	0.000	3.125 (1.845-5.294)
Total	452 (100.0)	440 (100.0)			

Table 3. LD and haplotypes of rs2069837 and rs17147230

1.056-5.025, P = 0.033). Similarly, the G allele was also related with increased risk for HCC (OR = 1.392, 95% CI = 1.046-1.852, P = 0.023). For rs17147230, compared with AA genotype, the risk was increased by 2.541 folds in TT carriers (OR = 2.089, 95% CI = 1.135-3.845, P = 0.017). In the study, we also found that frequency of T allele was significantly higher in cases than controls (OR = 1.326, 95% CI = 1.010-1.740, P = 0.042) (Table 2).

Linkage disequilibrium test and haplotype analysis

With Haploview online software, we figured out the linkage disequilibrium (LD) coefficient of rs2069837 and rs17147230 (D' = 0.89, r² = 0.16), suggesting there was significant LD between the two polymorphisms, so four haplotypes (A-A, A-T, G-A, G-T) were constituted. As shown in **Table 3**, frequencies of G-T haplotype differed between groups with statistical significance (P < 0.05). Moreover, G-T haplotype was associated with increased risk for HCC (OR = 3.125, 95% CI = 1.845-5.294, P = 0.000). In contrast, no statistically significant results were found in A-A, A-T and G-A haplotypes (P > 0.05).

Discussion

IL-6 is a cytokine with complex biological functions in vivo and plays a vital role in immune response, acute phase response and hematopoiesis regulation, thus it is considered as a core member in cytokine family [15]. Recent studies have found disorder of *IL*-6 expression was associated with some diseases, such as systemic lupus erythematosus, rheumatoid, multiple myeloma, and diabetes [16, 17]. Furthermore, *IL*-6 exerts influence on occurrence and development of various tumors though interfering with adhesion and mobility of cells, thrombosis, expression of tumor specific antigens and proliferation of tumor cells [11, 18]. A large number of cells can synthesize and secrete *IL*-6, including activated lymphocytes, mononuclear macrophages, bone marrow cells as well as a few tumor cells. Besides, *IL*-6 has been reported to regulate proliferation and differentiation of cells, immune defense system and hemopoiesis [19, 20].

The occurrence of HCC involves extremely complicated pathogenesis and its occurrence, development and metastasis are closely correlated with genetic mutations, cell signal pathways and abnormal proliferation of new vessels [21]. *IL*-6, also called hepatocyte stimulating factor, B cell stimulating factor or B cell differentiation factor, has been identified to have a vital part in regulating biological effects of hepatocytes [22]. As a hepatocyte stimulating factor, *IL*-6 is able to induce the synthesis of acute phase response proteins in acute inflammatory reactions caused by infections or injuries [18, 23].

The present study explored the association of rs2069837 and rs17147230 polymorphisms within *IL*-6 gene with HCC susceptibility and concluded that rs2069837 and rs17147230 polymorphisms both were related with increased risk for HCC. Further analysis indicated that G-T haplotype was also a genetic-susceptibility factor for HCC.

In summary, *IL*-6 gene rs2069837 and rs-17147230 polymorphisms were associated with HCC susceptibility. Due to the complicated pathogenesis of HCC, the definite mechanism though which *IL*-6 influences susceptibility to HCC is still unknown. Given this, more largescale and rigorous researches are necessary to investigate the issue.

Disclosure of conflict of interest

None.

Address correspondence to: Aiguang Wang, Department of Oncology, Qianfoshan Hospital Affili-

ated to Shandong University, NO. 16766 Jingshi Road, Jinan 250014, Shandong, China. E-mail: wag1974@126.com

References

- Parkin DM. The global health burden of infection-associated cancers in the year 2002. Int J Cancer 2006; 118: 3030-3044.
- [2] Oh S, Kim N, Yoon H, Choi YJ, Lee JY, Park KJ, Kim HJ, Kang KK, Oh DH, Seo AY, Lee JW, Shin CM, Park YS, Oh JC, Lee DH and Jung HC. Risk factors of atrophic gastritis and intestinal metaplasia in first-degree relatives of gastric cancer patients compared with age-sex matched controls. J Cancer Prev 2013; 18: 149-160.
- [3] Eagon PK, Francavilla A, DiLeo A, Elm MS, Gennari L, Mazzaferro V, Colella G, Van Thiel DH and Strazl TE. Quantitation of estrogen and androgen receptors in hepatocellular carcinoma and adjacent normal human liver. Dig Dis Sci 1991; 36: 1303-1308.
- [4] Parkin DM, Bray F, Ferlay J and Pisani P. Global cancer statistics, 2002. CA Cancer J Clin 2005; 55: 74-108.
- [5] Call DR, Borucki MK and Loge FJ. Detection of bacterial pathogens in environmental samples using DNA microarrays. J Microbiol Methods 2003; 53: 235-243.
- [6] Zhu K, Moriarty C, Caplan LS and Levine RS. Cigarette smoking and primary liver cancer: a population-based case-control study in US men. Cancer Causes Control 2007; 18: 315-321.
- [7] Zhang Y, Hayes A, Pritchard A, Thaker U, Haque MS, Lemmon H, Harris J, Cumming A, Lambert JC, Chartier-Harlin MC, St Clair D, Iwatsubo T, Mann DM and Lendon CL. Interleukin-6 promoter polymorphism: risk and pathology of Alzheimer's disease. Neurosci Lett 2004; 362: 99-102.
- [8] Inui A. [Pathogenesis and treatment of cancer anorexia-cachexia, with special emphasis on aged patients]. Nihon Ronen Igakkai Zasshi 2004; 41: 460-467.
- [9] Argiles JM, Busquets S, Felipe A and Lopez-Soriano FJ. Molecular mechanisms involved in muscle wasting in cancer and ageing: cachexia versus sarcopenia. Int J Biochem Cell Biol 2005; 37: 1084-1104.
- [10] Jiang YF, Zhao PW, Tan Y, Liu LH, Li MH, Matsuzaki Y and Niu JQ. [Molecular mechanisms of DHEA and DHEAs on apoptosis and cell cycle arrest via Akt pathway in hepatoma cell lines]. Zhonghua Gan Zang Bing Za Zhi 2007; 15: 441-444.

- [11] Jang Y, Kim OY, Hyun YJ, Chae JS, Koh SJ, Heo YM, Choi D, Shin DJ, Huttner K and Lee JH. Interleukin-6-572C>G polymorphism-association with inflammatory variables in Korean men with coronary artery disease. Transl Res 2008; 151: 154-161.
- [12] Nishimura M, Matsuoka M, Maeda M, Mizuta I, Mita S, Uchino M, Matsui M, Kuroda Y, Kawakami H, Kaji R, Adachi A and Uchiyama T. Association between interleukin-6 gene polymorphism and human T-cell leukemia virus type I associated myelopathy. Hum Immunol 2002; 63: 696-700.
- [13] Ota N, Nakajima T, Nakazawa I, Suzuki T, Hosoi T, Orimo H, Inoue S, Shirai Y and Emi M. A nucleotide variant in the promoter region of the interleukin-6 gene associated with decreased bone mineral density. J Hum Genet 2001; 46: 267-272.
- [14] Argiles JM, Busquets S and Lopez-Soriano FJ. Cytokines as mediators and targets for cancer cachexia. Cancer Treat Res 2006; 130: 199-217.
- [15] Graves DT and Kayal RA. Diabetic complications and dysregulated innate immunity. Front Biosci 2008; 13: 1227-1239.
- [16] Hong DS, Angelo LS and Kurzrock R. Interleukin-6 and its receptor in cancer: implications for translational therapeutics. Cancer 2007; 110: 1911-1928.
- [17] Xu H, Xiong C, He L, Wu B, Peng L, Cheng Y, Jiang F, Tan L, Tang L, Tu Y, Yang Y, Liu C, Gao Y, Li G, Zhang C, Liu S, Xu C, Wu H and Liang S. Trans-Resveratrol Attenuates High Fatty Acid-Induced P2X7 Receptor Expression and IL-6 Release in PC12 Cells: Possible Role of P38 MAPK Pathway. Inflammation 2015; 38: 327-37.
- [18] Arcone R, Fontaine V, Coto I, Brakenhoff JP, Content J and Ciliberto G. Internal deletions of amino acids 29-42 of human interleukin-6 (IL-6) differentially affect bioactivity and folding. FEBS Lett 1991; 288: 197-200.
- [19] Cussigh A, Falleti E, Fabris C, Bitetto D, Cmet S, Fontanini E, Bignulin S, Fornasiere E, Fumolo E, Minisini R, Pirisi M and Toniutto P. Interleukin 6 promoter polymorphisms influence the outcome of chronic hepatitis C. Immunogenetics 2011; 63: 33-41.
- [20] Fabris C, Toniutto P, Bitetto D, Fattovich G, Falleti E, Fontanini E, Cussigh A, Minisini R, Occhino G and Pirisi M. Gene polymorphism at the interleukin 6-174 G >C locus affects the outcome of chronic hepatitis B. J Infect 2009; 59: 144-145.
- [21] Romporn S, Hirankarn N, Tangkijvanich P and Kimkong I. Association of IFNAR2 and IL10RB genes in chronic hepatitis B virus infection. Tissue Antigens 2013; 82: 21-25.

- [22] Trim N, Morgan S, Evans M, Issa R, Fine D, Afford S, Wilkins B and Iredale J. Hepatic stellate cells express the low affinity nerve growth factor receptor p75 and undergo apoptosis in response to nerve growth factor stimulation. Am J Pathol 2000; 156: 1235-1243.
- [23] Dubuisson L, Desmouliere A, Decourt B, Evade L, Bedin C, Boussarie L, Barrier L, Vidaud M and Rosenbaum J. Inhibition of rat liver fibrogenesis through noradrenergic antagonism. Hepatology 2002; 35: 325-331.