

## Original Article

# Expression of matrix metalloproteinase-9, cyclooxygenase-2 and vascular endothelial growth factor are increased in gastrointestinal stromal tumors

Naiqing Liu\*, Jianguo Huang\*, Shuxiang Sun, Zhongjin Zhou, Jingyu Zhang, Fahui Gao, Qinli Sun

Department of General Surgery, Yishui Central Hospital, Linyi 276400, P. R. China. \*Equal contributors.

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**Abstract:** This study is to investigate the expression of matrix metalloproteinase-9 (MMP-9), cyclooxygenase-2 (COX-2) and vascular endothelial growth factor (VEGF) in gastrointestinal stromal tumor (GIST). Immunohistochemistry was performed to detect the expression of MMP-9, COX-2 and VEGF. The expression of MMP-9, COX-2 and VEGF was compared among different clinicopathological features of GIST. Spearman rank correlation analysis was conducted to analyze the correlation among MMP-9, COX-2 and VEGF. The positive expression rates of MMP-9, COX-2 and VEGF were 76.9%, 84.6% and 82.7%. The expression levels of MMP-9, COX-2 and VEGF were significantly different among the clinicopathological features of growth pattern, tumor diameter, metastasis, mitotic count and central necrosis ( $P < 0.05$ ). Their expression levels were higher in GIST tissues with higher levels of malignancy, tumor size, metastasis, mitotic count and central necrosis. However, their expression levels were not significantly different among age, gender, primary tumor site or CD117 expression. Additionally, there were positive correlations between COX-2 and VEGF ( $r = 0.612, P < 0.01$ ), between COX-2 and MMP-9 ( $r = 0.592, P < 0.05$ ), and between MMP-9 and VEGF ( $r = 0.690, P < 0.01$ ). MMP-9, COX-2 and VEGF expression levels are increased in GIST tissues and related with clinicopathological features of GIST.

**Keywords:** Gastrointestinal stromal tumor, matrix metalloproteinase-9, cyclooxygenase-2, vascular endothelial growth factor

## Introduction

Cyclooxygenase-2 (COX-2) is the rate-limiting enzyme for prostaglandin synthesis. It is not expressed in normal tissues, but up-regulated in malignant tumors such as esophageal cancer and colon cancer [1, 2]. COX-2 can promote tumor progression through inhibiting cell apoptosis and anti-tumor immunity and promoting tumor angiogenesis [3]. Dohadwala et al reported that COX-2 promoted the invasion and metastasis of non-small cell lung cancer through the paracrine and autocrine pathways of PGE2 [4]. Nakase et al found that COX-2 played important roles in the transition process from liver cirrhosis to liver cancer and in the progression of liver cancer, and that COX-2 was related to the poor prognosis of liver cancer [5].

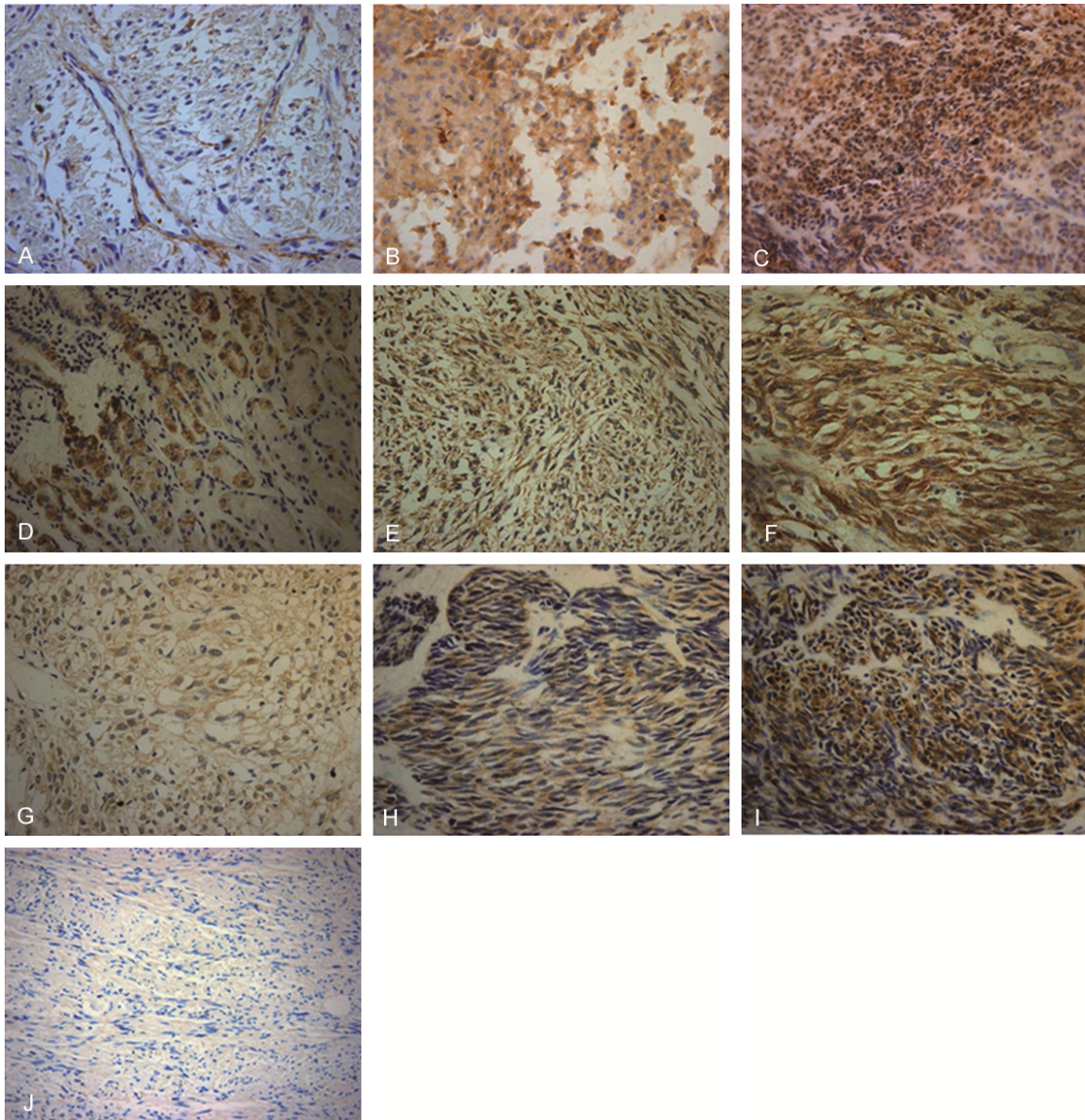
Matrix metalloproteinases (MMPs) can promote tumor invasion and metastasis through degrading the extracellular matrix. MMPs can also promote angiogenesis [6, 7]. MMP-9 is

one of the important MMPs, which can specifically degrade type IV collagen, the main component of basement membrane [8] and induce tumor angiogenesis. Vascular endothelial growth factor (VEGF) is an important angiogenic factor. It can regulate the generation of blood vessels and lymphatic vessels, and is closely related to tumor growth, invasion and metastasis [9, 10].

Gastrointestinal stromal tumor (GIST) is a common gastrointestinal cancer, which is originated in the mesenchymal tissue and mainly occurs in the gastrointestinal tract [11]. The histopathological features of GIST are similar to those of Cajal stromal cells [12, 13]. Currently, surgery is still the best means for the treatment of GIST without metastasis.

In this study, we detected the expression of COX-2, MMP-9 and VEGF in GIST, and analyzed their relationship with the clinicopathological features of GIST.

## Expression of MMP-9, COX-2 and VEGF in GIST



**Figure 1.** Representative immunohistochemical staining images of MMP-9, COX-2 and VEGF. Expression of MMP-9, COX-2 and VEGF in GIST tissues was detected by immunohistochemistry. Representative immunohistochemical results were shown. Magnification:  $\times 400$ . Cells with brown articles were defined as positive. The overall degree of staining was defined as follows: negative staining (-), score 0; weak positive staining (+), score 1-3; positive staining (++) , score 4-6; strong positive staining (+++), score 7-9. A. MMP-9 weak positive staining (+); B. MMP-9 positive staining (++) ; C. MMP-9 strong positive staining (+++); D. COX-2 weak positive staining (+); E. COX-2 positive staining (++) ; F. COX-2 strong positive staining (+++); G. VEGF weak positive staining (+); H. VEGF positive staining (++) ; I. VEGF strong positive staining (+++); J. Negative control.

### Materials and methods

#### Specimens

GIST tissues were collected from 52 patients who were admitted to Department of General Surgery, Yishui Central Hospital of Linyi from January 1998 to December 2005. They were

diagnosed as GIST by pathological examination. These patients included 28 males and 24 females. Their age ranged from 16 to 78 years, with an average age of 56.7 years. There were 14 cases with benign GIST, 11 cases with borderline GIST and 27 cases with malignant GIST. Prior written and informed consent were obtained from every patient and the study was

## Expression of MMP-9, COX-2 and VEGF in GIST

**Table 1.** The expression of MMP-9, COX-2 and VEGF in GIST

	Cases	Staining results				Positive rate (%)
		-	+	++	+++	
MMP9	52	12	9	18	13	76.9%
COX-2	52	8	9	20	15	84.6%
VEGF	52	9	9	16	18	82.7%

approved by the ethics review board of Yishui Central Hospital.

### *Immunohistochemical staining*

The paraffin blocks of GIST tissues were successively sectioned into slices (4  $\mu$ m in thickness). The expression levels of MMP-9, COX-2 and VEGF were detected by S-P immunohistochemical kit (Beijing Zhong Shan-Golden Bridge Biological Technology CO., Ltd., Beijing, China) according to manufacturer's instructions. Briefly, sections were dewaxed and rehydrated in graded alcohols. Then sections were incubated with 0.3% hydrogen peroxide to inactivate endogenous peroxidase activity. Antigen retrieval was achieved by incubating with sodium citrate (PH 6.0). After blocking, sections were incubated with primary antibodies at 37°C in the dark for 1 h. After washing with PBS, secondary antibodies were added and incubated in dark for 30 minutes. Then sections were developed with DAB chromogenic reagent. Finally, sections were counterstained with haematoxylin. Rabbit anti-human MMP-9 monoclonal antibody, mouse anti-human COX-2 monoclonal antibody, and rabbit anti-human VEGF monoclonal antibody were purchased from Beijing Zhong Shan-Golden Bridge Biological Technology CO., Ltd. PBS instead of primary antibodies was used as a negative control.

### *Evaluation of immunohistochemical staining results*

Cells with brown granules in the cytoplasm were determined as positive. Based on the staining intensity, immunohistochemistry staining results were scored as follows: score 0, no staining; score 1, light yellow; score 2, brownish yellow and score 3, tan. Based on the percentage of positive staining, immunohistochemistry staining results were scored as follows: score 0, 0% of positive staining; score 1, percentage of positive cells < 30%; score 2, percentage of

positive cells between 30% and 70%; and score 3, percentage of positive cells > 70%. The degree of staining was calculated by multiplying the score evaluated based on staining intensity and that on the percentage of positive staining. And the overall degree of staining was defined as follows: negative staining (-), score 0; weak positive staining (+), score 1-3; positive staining (++), score 4-6; strong positive staining (+++), score 7-9.

### *Statistical analysis*

Data was analyzed by SPSS 17.0 software (SPSS Inc., Chicago, Illinois, USA). The  $\chi^2$  test was used to analyze count data. The correlation among MMP-9, COX-2 and VEGF was analyzed by Spearman rank correlation analysis.  $P < 0.05$  was considered as statistically significant.

## Results

### *The positive expression of MMP-9, COX-2 and VEGF in GIST tissues*

To determine the expression levels of MMP-9, COX-2 and VEGF in GIST tissues, immunohistochemical staining was performed. The results of representative immunohistochemical staining were shown in **Figure 1**. The positive expression of MMP-9, COX-2 and VEGF all showed as diffuse or scattered brown particles in the cytoplasm of GIST cells. Some particles were dispersed in the mesenchyma of GIST tissues. Based on staining intensity and percentage of positive cells, the degree of staining was evaluated. MMP-9 weak positive staining (+), positive staining (++) and strong positive staining (+++) was shown in **Figure 1A-F** showed the weak positive staining (+) of COX-2, positive staining (++) of COX-2, and strong positive staining (+++) of COX-2, respectively. VEGF weak positive staining (+), positive staining (++) , strong positive staining (+++) was shown in **Figure 1G-I**. The staining result of negative control was shown in **Figure 1J**. As shown in **Table 1**, there were 40 cases with MMP9 positive expression, 44 cases with COX-2 positive expression and 43 cases with VEGF positive expression. The positive expression rate for MMP-9, COX-2 and VEGF was 76.9%, 84.6% and 82.7%, respectively. These results indicate that expression levels of MMP9, COX-2 and VEGF are relatively high in GIST tissues.



## Expression of MMP-9, COX-2 and VEGF in GIST

**Table 2.** Relationship between the expressions of MMP-9, COX-2 and VEGF in GIST and its clinicopathological features

		Cases	MMP-9		COX-2		VEGF	
			Positive cases (rate %)	P	Positive cases (rate %)	P	Positive cases (rate %)	P
Gender	Male	28	21 (75.0%)	0.722	23 (82.0%)	0.882	22 (78.6%)	0.358
	Female	24	19 (79.2%)		21 (87.5%)		22 (91.7%)	
Age (years)	< 65	19	15 (78.9%)	1.000	17 (89.5%)	0.736	15 (74.9%)	0.872
	≥ 65	33	25 (75.8%)		27 (81.8%)		28 (84.8%)	
Growth pattern	Benign	14	5 (35.7%)	0.042	8 (57.1%)	0.090	8 (57.1%)	0.156
	Borderline	11	9 (81.8%)	0.000	10 (90.9%)	0.004	10 (90.9%)	0.021
	Malignant	27	26 (96.3%)	0.196	26 (96.3%)	0.501	25 (92.6%)	1.000
Metastasis	No	21	12 (57.1%)	0.014	14 (66.7%)	0.010	14 (66.7%)	0.032
	Yes	31	28 (90.3%)		30 (96.8%)		29 (93.5%)	
Tumor diameter (cm)	D < 2 cm	9	2 (22.2%)	0.070	4 (44.4%)	0.160	4 (44.4%)	0.160
	2 ≤ D < 5 cm	11	8 (72.7%)	0.000	9 (81.8%)	0.001	9 (81.8%)	0.003
	D > 5 cm	32	30 (93.8%)	0.183	31 (96.9%)	0.156	30 (94.1%)	0.566
Mitotic count	n < 1/10 HP	18	10 (55.6%)	0.021	12 (66.7%)	0.027	11 (61.1%)	0.009
	n > 1/10 HP	34	30 (88.4%)		32 (94.1%)		32 (94.1%)	
Central necrosis	No	20	11 (55.0%)	0.009	13 (65.0%)	0.007	12 (60.0%)	0.002
	Yes	32	29 (90.6%)		31 (96.9%)		31 (96.9%)	
Primary site	Stomach	25	17 (68.0%)	0.481	19 (76.0%)	0.500	19 (77.8%)	0.500
	Intestine	18	14 (77.8%)	0.077	16 (88.9%)	0.162	16 (90.5%)	0.644
	Mesentery	9	9 (100%)	0.268	9 (100%)	0.538	8 (83.3%)	1.000
CD117	Positive	39	31 (82.1%)	0.254	34 (87.2%)	0.657	33 (84.2%)	0.657
	Negative	13	9 (61.5%)		10 (76.9%)		10 (81.3%)	

**Table 3.** The correlation among VEGF, MMP-9 and COX-2

	VEGF	MMP9
COX-2	r = 0.612 P = 0.0001	r = 0.592 P = 0.0001
MMP9	r = 0.690 P = 0.0001	-

### *Relationship between the expressions of MMP-9, COX-2 and VEGF in GIST and clinicopathological features of GIST*

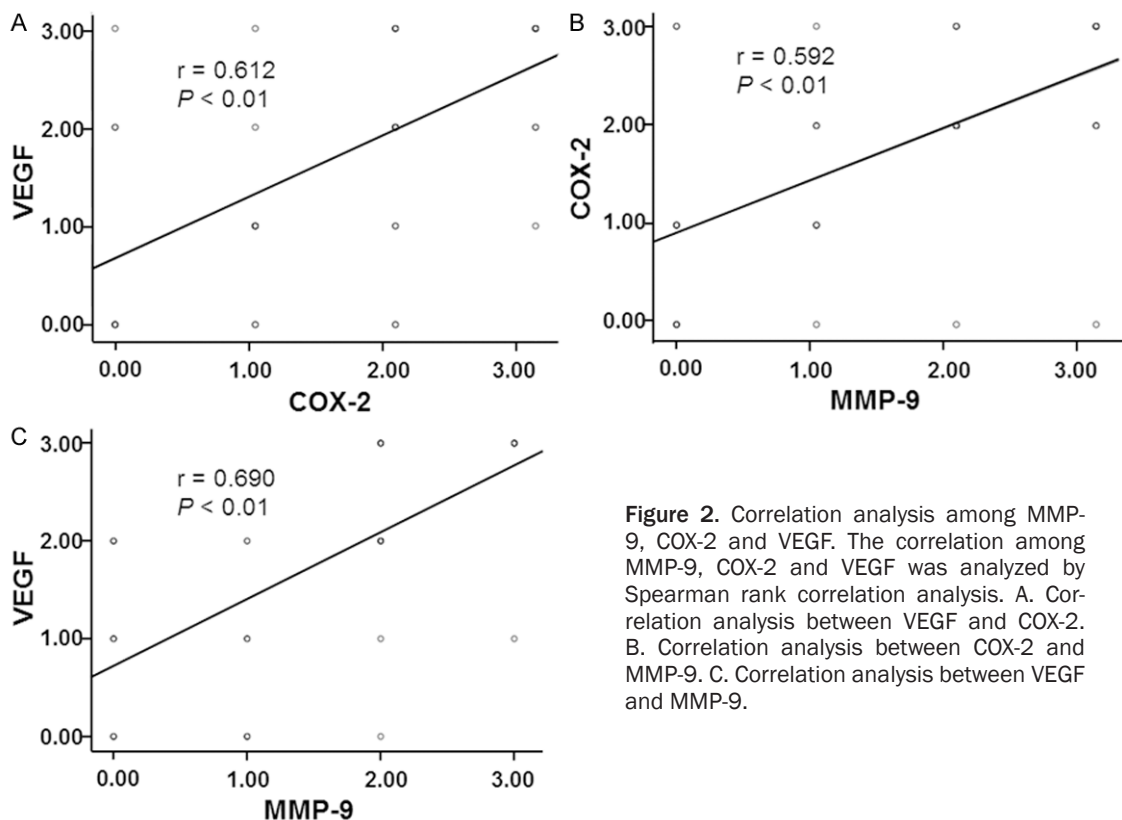
To determine the relationship between the expressions of MMP-9, COX-2 and VEGF in GIST and clinicopathological features of GIST, the positive rates of MMP-9, COX-2 and VEGF were compared among different clinicopathological features. The analyzed clinicopathological features included gender, age, growth pattern, metastasis, tumor diameter, mitotic count, central necrosis, primary site, and CD117 expression. As shown in **Table 2**, the positive rates of MMP-9, COX-2 and VEGF were significantly different among the clinicopathological features of growth pattern, tumor diameter, metastasis,

mitotic count and central necrosis ( $P < 0.05$ ). Compared with borderline GIST, malignant GIST had significantly expression levels of MMP-9 ( $P = 0.000$ ), COX-2 ( $P = 0.004$ ) and VEGF ( $P = 0.021$ ). The positive rates of MMP-9, COX-2 and VEGF in tumor diameter > 5 cm were significantly higher than those in tumor diameter < 2 cm ( $P = 0.000, 0.001, 0.003$  for MMP-9, COX-2 and VEGF respectively). GIST with metastasis, higher mitotic count central necrosis also had significantly higher expression levels of MMP-9, COX-2 and VEGF ( $P < 0.05$ ). However, there were no significant differences in the positive rates of MMP-9, COX-2 and VEGF among the clinicopathological features of gender, age, primary tumor site or CD117 expression ( $P > 0.05$ ). These results suggest that positive expression of MMP-9, COX-2 and VEGF are related with clinicopathological features of growth pattern, metastasis, tumor diameter, mitotic count, and central necrosis.

### *Correlation among VEGF, MMP-9 and COX-2*

Spearman rank correlation analysis was performed to analyze the correlation among the expression of MMP-9, COX-2 and VEGF. As

## Expression of MMP-9, COX-2 and VEGF in GIST



**Figure 2.** Correlation analysis among MMP-9, COX-2 and VEGF. The correlation among MMP-9, COX-2 and VEGF was analyzed by Spearman rank correlation analysis. A. Correlation analysis between VEGF and COX-2. B. Correlation analysis between COX-2 and MMP-9. C. Correlation analysis between VEGF and MMP-9.

shown in **Table 3** and **Figure 2**, there were positive correlations between the expression of COX-2 and VEGF ( $r = 0.612$ ,  $P < 0.01$ ), the expression of COX-2 and MMP-9 ( $r = 0.592$ ,  $P < 0.01$ ) and the expression of MMP-9 and VEGF ( $r = 0.690$ ,  $P < 0.01$ ). These results indicate that there is positive correlation among MMP-9, COX-2 and VEGF.

### Discussion

It is reported that MMP-9, COX-2 and VEGF are highly expressed in many tumor tissues [14, 15]. They are involved in tumor angiogenesis, local invasion and metastasis [16, 17], and are closely related to the tumorigenesis and tumor development [18, 19]. This study revealed that the positive expression rate of COX-2 in GIST was 84.6% and that COX-2 expression was higher in malignant GIST tissues than that in benign GIST tissues. Our result was consistent with a previous study [20], indicating that COX-2 may be used as an indicator to evaluate the malignancy of GIST. We also found that the expression of COX-2 was closely related to tumor metastasis, mitotic count and central necrosis of GIST, which suggests that COX-2

might be involved in the progression and metastasis of GIST.

In this study, the positive expression rates of MMP-9 and VEGF in GIST were 76.9% and 82.7%, respectively. And, expression levels of MMP-9 and VEGF were significantly higher in malignant GIST than that in benign GIST. Their expression levels were associated with the GIST tumor size, invasion and metastasis, mitotic count and central necrosis. In gastric cancer, the positive expression of MMP9 and VEGF is associated with tumor size, degree of invasion, and lymph node metastasis [21]. These results suggest both of MMP-9 and VEGF may be involved in GIST angiogenesis, thus promoting its invasion and metastasis.

Further investigation showed that there were positive correlations among the expressions of COX-2, VEGF and MMP-9 in GIST. Tsujii et al reported that VEGF expression in colon cancer cell lines was up-regulated after transfection with COX-2, and the COX-2 specific inhibitor NS398 could block this effect [22]. This suggests that COX-2 might promote tumor angiogenesis through VEGF pathway. In a study on

## Expression of MMP-9, COX-2 and VEGF in GIST

breast cancer cells, selective inhibitors of COX-2 could significantly decrease the expression of MMP and reduce cell proliferation and invasiveness of tumor cells [23]. One study also showed that selective inhibitors of COX-2 reduced the viability of head and neck squamous cell, and inhibited their invasion and adhesion through down-regulating the expressions of MMP-9, MMP-2 and VEGF [24]. This might be because that over expression of COX-2 in tumor cells induces the upregulation of MMP-9 and VEGF, which further promote tumor growth, invasion and metastasis through degrading the extracellular matrix and promoting angiogenesis [25, 26].

In summary, expression levels of COX-2, VEGF and MMP-9 are increased in GIST tissues. They may be used as indicators to assess the malignancy grades of GIST. COX-2 could promote tumor angiogenesis, invasion and metastasis through upregulation of MMP-9 and VEGF. The application of COX-2 inhibitor could provide new strategies for the clinical treatment of GIST.

### Disclosure of conflict of interest

None.

**Address correspondence to:** Qinli Sun, Department of General Surgery, Yishui Central Hospital, No. 17 Jiankang Road, Linyi 276400, P. R. China. Tel: 13854952789; Fax: 0539-2251647; E-mail: sun-qinli0808@163.com

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## Expression of MMP-9, COX-2 and VEGF in GIST

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