

## Original Article

# Pathological changes of cochlear in deaf mice at different time after mouse cytomegalovirus infection

Yongyuan Tian, Xinguo Liu, Hongjian Liu, Jinyan Xing

Department of Ear-Norse-Throat, The Zhumadian City Center Hospital, No. 747 Zhonghua Road, Zhumadian 463000, P. R. China

Received January 17, 2015; Accepted April 28, 2015; Epub May 15, 2015; Published May 30, 2015

**Abstract:** Objective: This study aims to observe the pathological changes of inner ear in deaf mice at different time after mouse cytomegalovirus infection. Methods: A total of 60 BALB/C mice were divided into 2 groups randomly: model group (A) and control group (B). In model group, 10  $\mu$ l of MCMV was injected into the brain of each mouse while 10  $\mu$ l of physiological saline was injected in control group. 10 cochlear samples were taken from 5 mice selected from each group randomly after infection for 1, 3, 5, 7, 14 and 21 days respectively. They were detected with PCR and HE staining methods. Auditory brain stem response was determined. The apoptosis of spiral ganglion (SGN) cells was detected by apoptosis assay kit. The levels of Bcl-2 and Bax were detected by RT-PCR and western blotting methods. Results: In group A, PCR results were negative after infection for 1 day, they were positive after infection for 3 days to 21 days. In group B, PCR results were negative in the experimental period. Compared with group B, ABR I wave latency and threshold increased while ABR I wave decreased in group A. There were no obvious hyperemia and inflammatory cells infiltration in group B, In group A, hemorrhage of scala tympani and scala vestibule appeared and reached highest peak after infection for 3 days accompanied by inflammatory cell infiltration; the vestibular membrane thickened after infection for 5 days; cell gap of SGN cells widened, arranged more sparsely with cell edema after infection for 7 days accompanied by infiltration of plasma cells; fibroblast proliferation and fibrosis appeared after infection for 14 days. Conclusions: MCMV infection occurred in cochlear after MCMV infection for 3 days and could sustain, the continues pathological changes of inner will bring difficulties to the treatment of CMV deafness, further studies on the specific mechanism of SGN changes caused by CMV infection will provide an important target for the treatment of CMV deafness.

**Keywords:** Mouse cytomegalovirus (MCMV), cochlea, MCMV-DNA, inflammatory reaction

## Introduction

Cytomegalovirus (CMV) is a member of herpes virus, its infection rate is about 0.2%-2.3% in the newborn [1, 2]. Congenital CMV infection will be affected by maternal age, race, economic status and other social factors [3]. Almost all the people faced the risk of CMV infection and the infection rate in populous and economically undeveloped regions is higher. The clinical symptoms such as skin ecchymosis, jaundice, hepatosplenomegaly, chorioretinitis or cerebellar malformation could appear in children with congenital infection or acquired infection of human cytomegalovirus (HCMV) [4, 5]. Dahle [6] first reported progressive hearing loss in children after congenital HCMV infection in 1974, and suggested that subclinical congeni-

tal HCMV infection may be one of the causes of sensorineural hearing loss (SNHL) in infant. At present, congenital HCMV infection is thought to be the main cause of infantile non hereditary SNHL [7-11]. Deafness or hearing loss caused by CMV infection can be unilateral or bilateral, they had undulatory, progressive and delayed characteristics, and began in a few months even years after birth [7, 12]. In this study we observed the pathological changes of inner ear in deaf mice at different time with MCMV-induced hearing loss mouse model [13].

## Materials and methods

### *Experimental animals and cells*

NIH/3T3 cells and MCMV Smith virus were gifted from Shandong Provincial Academy of

Medical Sciences. A total of 60 SPF grade adult healthy BALB/c mice weighing 20 to 30 g were obtained from the animal experimental center of Shandong University. MCMV Smith virus infected NIH/3T3 cells and the TCID<sub>50</sub> was 104 IU/0.1 ml using cytopathic virus titration [15]. The mice were randomly divided into model group (A, n=30) and control (uninfected) group (B, n=30). In model group, 10 µl of MCMV was injected into the brain of each mouse while 10 µl of physiological saline was injected in control group.

### *Determination of auditory brain stem response (ABR)*

Determination of ABR was performed according to reference [13]. The mice were anaesthetized with ether after infection for 3 weeks. ABR was determined in the acoustic electric shielding state. The detection electrode was placed under the skin of test ear mastoid, the reference electrode was placed under the skin of the middle of head, the ground electrode was placed on the toe. They were stimulated with rarefaction wave and short sound. The stimulus frequency was 13 Hz and stimulus intensity was 80 dB, scanning speed was 1 ms/D.

### *Cochlea samples*

10 cochlear samples were taken from 5 mice selected from each group randomly after infection for 1, 3, 5, 7, 14 and 21 days respectively. They were detected with PCR, HE staining and apoptosis of spiral ganglion (SGN) cells.

### *PCR detection*

DNA was isolated from the samples using DNA isolation kit. Primers sequence was follows. Forward: 5'-TCAGCCATCAACTCTGCTACCAAC-3'; Reverse: 5'-ATCTGAAACAGCCGTATATCATCTTG-3' [14]. The reaction conditions were 94°C 5 min; 94°C 30 s, 55°C 30 s and 72°C 30 s, 30 cycles; 72°C 10 min. Following amplification for 35 cycles, the products were detected using 1.5% agarose gel electrophoresis and viewed with GIS-2008 digital gel imaging system.

### *HE staining*

Cochlear samples were fixed with 10% formalin for 3 days and decalcified with EDTA for 1 week. They were embedded with paraffin and sliced.

They were observed under microscope after routine HE staining.

### *Detection of apoptosis*

The apoptosis of SGN cells was detected by apoptosis assay kit according to the manual. Apoptosis related genes of Bcl-2 and Bax were detected by RT-PCR and western blotting methods. Total RNA was extracted from the cochlear samples using the RNA extraction kit according to the protocol. Samples (1 µg per sample from a total volume of 20 µL) were quantified by spectrophotometry. The expression levels of Bcl-2 and Bax were determined by RT-PCR kit following the manufacturer's instructions. All values obtained were normalized to mouse β-actin. The sequences of PCR primers were as follows: Bcl-2 forward: 5'-GGATTGTGGCCTTCTTGAG-3', reverse: 5'-CCAAACTGAGCAGAGTCTTC-3'; Bax forward: 5'-TCCACCAAGAAGCTGAGCGAG-3', reverse: 5'-GTCCAGCCCATGATGGTCTCT-3'; β-actin forward: 5'-CTGCCGCATCCTCTTCTC-3', reverse: 5'-CTGTCGCCTTACCGTTCC-3'. The protein levels of Bcl-2 and Bax were determined by western blotting method. The samples were lysed in RIPA lysis buffer, and the lysates were harvested by centrifugation (12,000 rpm) at 4°C for 5 min. Protein samples were then separated by electrophoresis in a 10% sodium dodecyl sulfate polyacrylamide gel and were transferred onto a polyvinylidene fluoride membrane. After blocking the nonspecific binding sites for 60 min with 5% nonfat milk, the membranes were incubated overnight at 4°C with required primary antibodies. The membranes were then washed three times with Tris-buffered saline with Tween-20 (TBST) for 10 min and were probed with the horseradish peroxidase (HRP)-conjugated goat anti-mouse IgG antibody at room temperature for 1 h. After three washes with TBST, the membranes were developed using an enhanced chemiluminescence system (Applygen Technologies Inc, China). The protein levels were normalized to the level of GAPDH detected using goat anti-mouse GAPDH monoclonal antibody.

## Results

### *ABR determination*

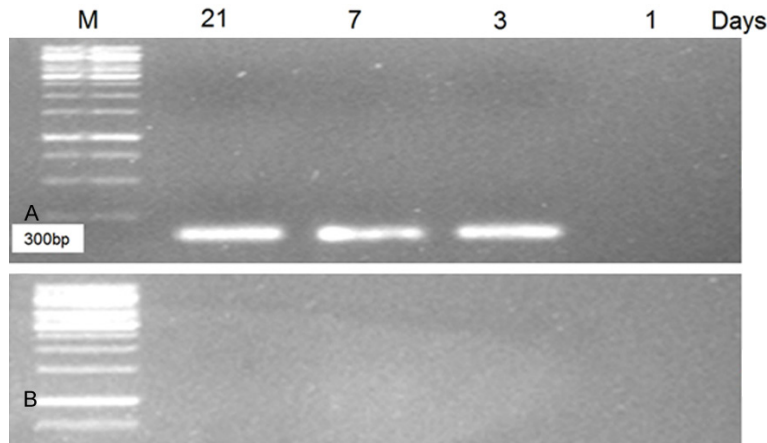
Compared with group B after infection for 3 weeks, ABR I wave latency and threshold

## Pathological changes of cochlear in deaf mice after MCMV infection

**Table 1.** Comparison of ABR I wave latency, amplitude and threshold in different group after infection for 3 weeks ( $\bar{x} \pm s$ )

Group	N	I wave latency (ms)	I wave amplitude (uv)	Threshold (dB)
A	30	1.93±0.11*	1.82±0.35*	66.26±4.19*
B	30	1.27±0.13	4.91±0.23	24.52±3.82

\* $P < 0.05$  vs B group.



**Figure 1.** PCR results of MCMV infection in different groups. A: Group A; B: Group B. In group A, PCR results were negative after infection for 1 day, they were positive after infection for 3 days to 21 days. In group B, PCR results were negative in the experimental period.

increased while ABR I wave decreased in group A ( $P < 0.05$ ) (Table 1).

### PCR results

In group A, PCR results were negative after infection for 1 day, they were positive after infection for 3 days to 21 days. In group B, PCR results were negative in the experimental period. These showed that MCMV infection occurred in cochlear after intracranial injection of MCMV suspension for 3 days and could sustain to 3rd week (Figure 1).

### Pathological changes of cochlear in different time and groups

There were no obvious hyperemia and inflammatory cells infiltration in group B, no thickening of the vestibular membrane, SGN cells arranged densely without widening cell gap and no fibrosis. In group A, there were no obvious changes after infection for 1 day, massive hemorrhage of scala tympani and scala vestibule appeared and reached highest peak after infection for 3 days accompanied by inflammatory cell infiltration. The vestibular membrane

thickened after infection for 5 days; cell gap of SGN cells widened, arranged more sparsely with cell edema after infection for 7 days accompanied by infiltration of plasma cells; fibroblast proliferation and fibrosis appeared after infection for 14 days (Figure 2).

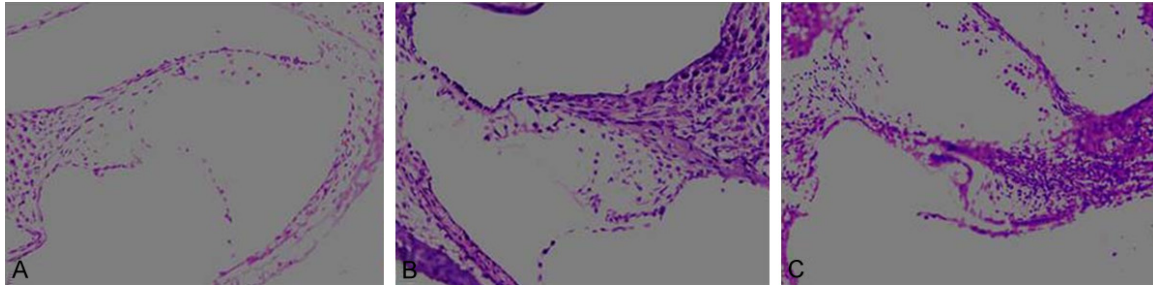
### Apoptosis results

As shown in Figure 3, there were few apoptotic cells and no obvious changes in each time point in control group, while the apoptotic cells increased significantly in each time point after MCMV infection in group A compared to the control group ( $P < 0.05$ ), it reached the peak after MCMV infection for 7 days. RT-PCR and western blotting results showed that the levels of Bcl-2 decreased and the levels of Bax increased in group

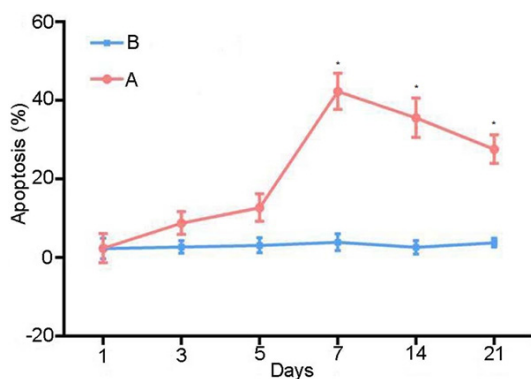
A after MCMV infection for 3 days when compared to group B ( $P < 0.05$ , Figures 4 and 5).

### Discussion

The mechanism of SNHL caused by congenital CMV infection is not very clear. The delayed and progressive characteristics may be due to the persistent inflammation mediated by virus copy or host immune cells. Persistent inflammation such as the cell loss of outer and inner hair, cochlear ROS, increased TNF- $\alpha$  and interleukins could appear in cochlear after inner ear infected by CMV [15-17]. It has been confirmed that, CMV virus can be found in human and animal model of cochlea by PCR or immunohistochemistry [18-21]. In this study we found that MCMV infection occurred in cochlear after intracranial injection of MCMV suspension for 3 days and could sustain to 3<sup>rd</sup> week. The pathological changes of cochlear also occurred from 3<sup>rd</sup> day after intracranial injection of MCMV suspension, which including hemorrhage of scala vestibuli and scala tympani with infiltrating inflammatory cells, stria vascular hyperemia, vestibular membrane thickening, spiral ganglion (SGN) cell gap widened and the scala tym-



**Figure 2.** HE staining results ( $\times 400$ ). A: HE staining results after infection for 5 day in group A, the vestibular membrane thickened after infection for 5 days; B: HE staining results after infection for 1 day in group A, there was no obvious change after infection for 1 day; C: HE staining results in group B, there were no obvious hyperemia and inflammatory cells infiltration in group B.



**Figure 3.** Apoptosis results of SGN cells. A: Group A; B: Group B. There were few apoptotic cells and no obvious changes in each time point in control group, while the apoptotic cells increased significantly in each time point after MCMV infection in group A compared to group B. \*Compared with group B,  $P < 0.05$ .

pani fibrosis. Hemorrhage of scala vestibuli and scala tympani with infiltrating inflammatory cells was more serious after infection for 3-5 days which maybe because of the acute inflammatory reaction in cochlea caused by MCMV infection. However, other studies [22, 23] showed that these occurred after infection for 8 days.

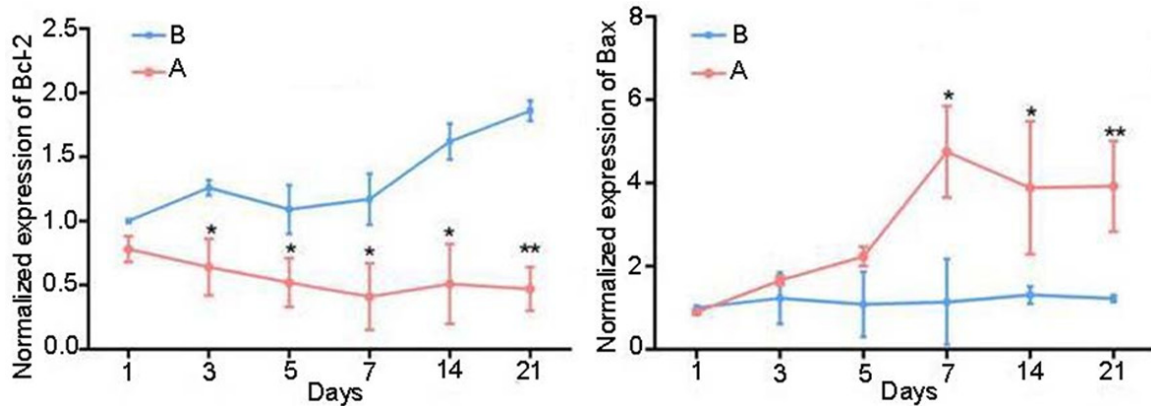
Previous studies showed that SGN and perilymphatic epithelial cells were the main parts of virus infection, while the inner and outer hair cells were not main infection parts [17]. In this study, we found that the intercellular gap of SGN cells changed after infection for 7 days with cellular edema. This was caused by persistent inflammation in the cochlea or by the cytopathic effect (CPE) of MCMV remained unclear. There were few apoptotic cells and no obvious changes in each time point in control group,

while the apoptotic cells increased significantly in each time point after MCMV infection in group A. RT-PCR and western blotting results showed that the levels of Bcl-2 decreased and the levels of Bax increased in group A after MCMV infection for 3 days when compared to group B. The relationship between CMV and host cells is very complex, its infection in the body not only can promote the apoptosis of host cells but also can inhibit the apoptosis of the host cells, which maybe correlated with the infection dose [25]. It inhibits the apoptosis of the host cells in the early stage of infection and promotes the apoptosis of host cells in the later stage of infection [26]. The differences existed in different studies because of different modeling methods and different strains of CMV activity.

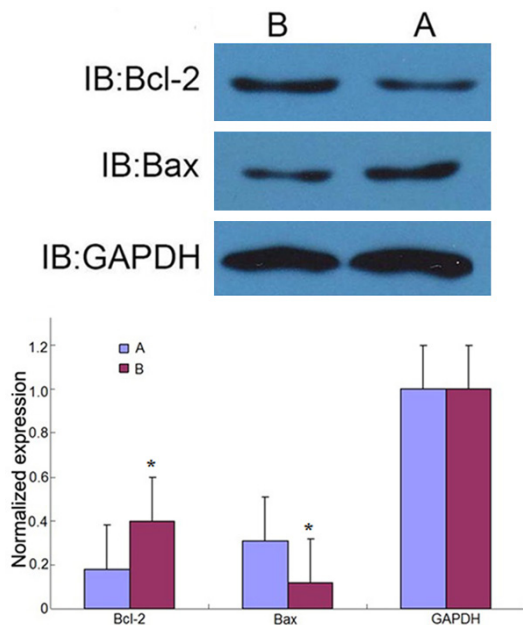
The most effective treatment is artificial cochlear implantation for the pre language children with bilateral severe or very severe SNHL. Studies have found that cochlear fibrosis occurred after long-term CMV infection in human [16] and guinea pig [18, 27] and fibrosis could develop into ossification. Although the fibrosis and ossification of cochlear is not a contraindication for cochlear implantation, it is difficult for electrode placement. We found cochlear fibrosis in one mouse after infection for 14 days and in the other mouse after infection for 21 days without obvious ossification. The target cells of artificial cochlea were SGN cells, so the residual SGN cells' number and function in patients determined cochlear transplantation effect. In this study we found morphological changes appeared in SGN cells after infection for 7 days and the cause was not clear. At present, the curative effect of antiviral



## Pathological changes of cochlear in deaf mice after MCMV infection



**Figure 4.** RT-PCR results of Bcl-2 and Bax expression. A: Group A; B: Group B. The levels of Bcl-2 decreased and the levels of Bax increased in group A after MCMV infection for 3 days when compared to group B. \*Compared with group B,  $P < 0.05$ .



**Figure 5.** Western blotting results of Bcl-2 and Bax proteins. A: Group A; B: Group B. The levels of Bcl-2 decreased and the levels of Bax increased in group A after MCMV infection for 3 days when compared to group B. \*Compared with group B,  $P < 0.05$ .

therapy alone for CMV deafness was unsatisfactory. Further research on the specific mechanism of SGN changes caused by CMV infection will provide an important target for the treatment of CMV deafness.

In a word, MCMV infection occurred in cochlear after intracranial injection of MCMV suspension for 3 days and could sustain, the continues pathological changes of inner will bring dif-

ficulties to the treatment of CMV deafness, further studies on the specific mechanism of SGN changes caused by CMV infection will provide an important target for the treatment of CMV deafness.

### Disclosure of conflict of interest

None.

**Address correspondence to:** Yongyuan Tian, Department of Ear-Norse-Throat, The Zhumadian City Center Hospital, No. 747 Zhonghua Road, Zhumadian 463000, P. R. China. Tel: 86-15103809757; E-mail: yonyuantian@126.com

### References

- [1] Schlesinger Y, Halle D, Eidelman AI, Reich D, Dayan D, Rudensky B, Raveh D, Branski D, Kaplan M, Shefer V and Miron D. Urine polymerase chain reaction as a screening tool for the detection of congenital cytomegalovirus infection. *Arch Dis Child Fetal Neonatal* Ed 2003; 88: F371-374.
- [2] Zucca C, Binda S, Borgatti R, Triulzi F, Radice L, Butte C, Barkhaus PE and Barbi M. Retrospective diagnosis of congenital cytomegalovirus infection and cortical maldevelopment. *Neurology* 2003; 61: 710-712.
- [3] Kimberlin DW, Lin CY, Sanchez PJ, Demmler GJ, Dankner W, Shelton M, Jacobs RF, Vaudry W, Pass RF, Kiell JM, Soong SJ and Whitley RJ. National Institute of A and Infectious Diseases Collaborative Antiviral Study G. Effect of ganciclovir therapy on hearing in symptomatic congenital cytomegalovirus disease involving the central nervous system: a randomized, controlled trial. *J Pediatr* 2003; 143: 16-25.

- [4] Oliver SE, Cloud GA, Sanchez PJ, Demmler GJ, Dankner W, Shelton M, Jacobs RF, Vaudry W, Pass RF, Soong SJ, Whitley RJ and Kimberlin DW. National Institute of Allergy IDCASG. Neurodevelopmental outcomes following ganciclovir therapy in symptomatic congenital cytomegalovirus infections involving the central nervous system. *J Clin Virol* 2009; 46 Suppl 4: S22-26.
- [5] Engman ML, Lewensohn-Fuchs I, Mosskin M and Malm G. Congenital cytomegalovirus infection: the impact of cerebral cortical malformations. *Acta Paediatr* 2010; 99: 1344-1349.
- [6] Dahle AJ, McCollister FP, Hamner BA, Reynolds DW and Stagno S. Subclinical congenital cytomegalovirus infection and hearing impairment. *J Speech Hear Disord* 1974; 39: 320-329.
- [7] Fowler KB, McCollister FP, Dahle AJ, Boppana S, Britt WJ and Pass RF. Progressive and fluctuating sensorineural hearing loss in children with asymptomatic congenital cytomegalovirus infection. *J Pediatr* 1997; 130: 624-630.
- [8] Harris S, Ahlfors K, Ivarsson S, Lernmark B and Svanberg L. Congenital cytomegalovirus infection and sensorineural hearing loss. *Ear Hear* 1984; 5: 352-355.
- [9] Fowler KB, Dahle AJ, Boppana SB and Pass RF. Newborn hearing screening: will children with hearing loss caused by congenital cytomegalovirus infection be missed? *J Pediatr* 1999; 135: 60-64.
- [10] Grosse SD, Ross DS and Dollard SC. Congenital cytomegalovirus (CMV) infection as a cause of permanent bilateral hearing loss: a quantitative assessment. *J Clin Virol* 2008; 41: 57-62.
- [11] Misono S, Sie KC, Weiss NS, Huang ML, Boeckh M, Norton SJ and Yueh B. Congenital cytomegalovirus infection in pediatric hearing loss. *Arch Otolaryngol Head Neck Surg* 2011; 137: 47-53.
- [12] Peckham CS, Stark O, Dudgeon JA, Martin JA and Hawkins G. Congenital cytomegalovirus infection: a cause of sensorineural hearing loss. *Arch Dis Child* 1987; 62: 1233-1237.
- [13] Qiao Y, Meng L, Wang J and Meng H. Effect of Ganciclovir on murine cytomegalovirus-induced hearing loss in a mouse model. *Cell Biochem Biophys* 2011; 61: 407-412.
- [14] Tang-Feldman YJ, Wojtowicz A, Lochhead GR, Hale MA, Li Y and Pomeroy C. Use of quantitative real-time PCR (qRT-PCR) to measure cytokine transcription and viral load in murine cytomegalovirus infection. *J Virol Methods* 2006; 131: 122-129.
- [15] Yuehua Q, Longzhen Z, Kailin X, Lingyu Z, Lingjian M, Jun W and Hong M. Inflammatory lesions of cochlea in murine cytomegalovirus-infected mice with hearing loss. *Cell Biochem Biophys* 2012; 62: 281-287.
- [16] Rarey KE and Davis LE. Temporal bone histopathology 14 years after cytomegalic inclusion disease: a case study. *Laryngoscope* 1993; 103: 904-909.
- [17] Schachtele SJ, Mutnal MB, Schleiss MR and Lokensgard JR. Cytomegalovirus-induced sensorineural hearing loss with persistent cochlear inflammation in neonatal mice. *J Neurovirol* 2011; 17: 201-211.
- [18] White DR, Choo DI, Stroup G and Schleiss MR. The effect of cidofovir on cytomegalovirus-induced hearing loss in a Guinea pig model. *Arch Otolaryngol Head Neck Surg* 2006; 132: 608-615.
- [19] Bauer PW, Parizi-Robinson M, Roland PS and Yegappan S. Cytomegalovirus in the perilymphatic fluid. *Laryngoscope* 2005; 115: 223-225.
- [20] Di Nardo W, Cattani P, Scorpecci A, Giannantonio S, D'Onghia S, Fadda G and Paludetti G. Cytomegalovirus DNA retrieval in the inner ear fluids of a congenitally deaf child one month after primary infection: a case report. *Laryngoscope* 2011; 121: 828-830.
- [21] Sugiyama S, Yoshikawa T, Nishiyama Y, Morishita Y, Sato E, Hattori T and Nakashima T. Detection of human cytomegalovirus DNA in perilymph of patients with sensorineural hearing loss using real-time PCR. *J Med Virol* 2003; 69: 72-75.
- [22] Harris JP, Woolf NK, Ryan AF, Butler DM and Richman DD. Immunologic and electrophysiological response to cytomegaloviral inner ear infection in the guinea pig. *J Infect Dis* 1984; 150: 523-530.
- [23] Woolf NK, Harris JP, Ryan AF, Butler DM and Richman DD. Hearing loss in experimental cytomegalovirus infection of the guinea pig inner ear: prevention by systemic immunity. *Ann Otol Rhinol Laryngol* 1985; 94: 350-356.
- [24] Reuter JD. Cytomegalovirus induces T-cell independent apoptosis in brain during immunodeficiency. *J Clin Virol* 2005; 32: 218-223.
- [25] Reuter JD, Wilson JH, Idoko KE and van den Pol AN. CD4+ T-cell reconstitution reduces cytomegalovirus in the immunocompromised brain. *J Virol* 2005; 79: 9527-9539.
- [26] Goldmacher VS. Cell death suppression by cytomegaloviruses. *Apoptosis* 2005; 10: 251-265.
- [27] Nomura Y, Hara M and Kurata T. Experimental herpes simplex virus and cytomegalovirus labyrinthitis. *Acta Otolaryngol Suppl* 1989; 457: 57-66.
- [28] Juanjuan C, Yan F, Li C, Haizhi L, Ling W, Xinrong W, Juan X, Tao L, Zongzhi Y and Suhua C. Murine model for congenital CMV infection and hearing impairment. *Virol J* 2011; 8: 70.