

Original Article

Relationship between *ADH2* Arg47His variation and hepatocellular carcinoma susceptibility: a meta analysis

Shuzhen Liu^{1*}, Yongchun Cui^{2*}, Baohong Yang¹, Ping Chai¹, Zheng Su¹, Qianqian Zhang¹, Dejie Zheng¹, Rui Li¹, Guohua Yu¹

¹Clinical Oncology Department, Weifang People's Hospital, Kuiwen District, Weifang, Shandong, China; ²Department of Drug Clinical Trial Institution of Shandong Cancer Hospital & Institute, Jinan, Shandong, China. *Equal contributors.

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Abstract: Background: To further investigate the relationship between *ADH2* Arg47His variation and hepatocellular carcinoma (HCC) susceptibility through a meta-analysis. Methods: The related articles were searched in PubMed, Embase and CNKI databases. And finally 518 cases and 607 controls were included in our meta-analysis Odds ratios (ORs) with 95% confidence intervals (95% CIs) were used to assess the relationship between *ADH2* Arg47His variation and HCC risk. A fixed-effect model or a random-effect model was applied according to the between-study heterogeneity. Results: Quantitative synthesis demonstrated that no significant association was found between *ADH2* Arg47His variation and HCC susceptibility (His/His vs. Arg/Arg: OR=0.99, 95% CI=0.79-1.25; His/His + Arg/His vs. Arg/Arg: OR=1.01, 95% CI=0.86-1.20; His/His vs. Arg/Arg + Arg/His: OR=0.90, 95% CI=0.74-1.11; His vs. Arg: OR=0.98, 95% CI=0.86-1.11; Arg/His vs. Arg/Arg: OR=1.05, 95% CI=0.82-1.34). Conclusion: Our analysis showed that *ADH2* Arg47His variation may not be associated with HCC susceptibility.

Keywords: *ADH2*, variation, hepatocellular carcinoma

Introduction

Hepatocellular carcinoma (HCC), a main form of liver cancer, is the third leading cause of cancer death in the world. The incidence of HCC is increased rapidly and it has been considered as the major health problem in the worldwide [1-3]. Alcohol drinking, smoking and chronic viral infections including hepatitis B virus and hepatitis C virus may contribute to increase HCC risk [4-9]. Additionally, genetic factors are considered to be associated with the risk for HCC [10, 11].

Alcohol dehydrogenase (ADH) isoenzyme is a metabolic barrier against self-administer ethanol produced in carbohydrates through the fermentation of bacteria [12, 13]. *ADH2*, a well known member of ADH family, locates on chromosome 4q22-23, and includes three alleles, namely *ADH2* 1*, *ADH2* 2* and *ADH2* 3*. *ADH2* gene is mainly responsible for the conversion of ethanol to carcinogenic metabolite during the elimination phase [14-16]. Additionally, *ADH2* expression could result in high blood acetaldehyde levels, which can easily

lead to DNA damage, and finally cause the occurrence of cancer [17-19]. Previous studies have found that *ADH2* gene was associated with some cancers, such as colorectal cancer and esophageal cancer [20, 21].

ADH2 has numerous polymorphic sites, and one of the most studied functional polymorphisms in *ADH2* gene is Arg47His with an acid substitution of arginine (Arg) to histidine (His) at codon 47. In addition, Arg47His shows great impact on enzyme activities, and ultimately causes the occurrence of diseases. Several studies have investigated the association of *ADH2* Arg47His polymorphism with HCC risk [22-24]. Based on the published studies, our meta-analysis was aimed to make clear the relationship between *ADH2* Arg47His polymorphism and HCC risk.

Methods

Search strategy

The related articles were searched in PubMed, Embase and CNKI databases with the terms of

Table 1. Principle characteristics of the studies included in the meta-analysis

First author	Year	Country	Ethnicity	Control source	Genotyping method	Case	Control	HWE
Takeshita_Male	2000	Japan	Asian	Hospital based	RFLP	85	101	0.158
Takeshita_Female	2000	Japan	Asian	Hospital based	RFLP	17	24	0.102
Sakamoto T	2006	Japan	Asian	Hospital based	PCR-CTPP	209	275	0.475
Ding J	2008	China	Asian	Population based	PCR-RFLP	207	207	0.806

RFLP: restriction fragment length polymorphism; PCR-CTPP: PCR-confronting two-pair primers; PCR-RFLP: PCR-restriction fragment length polymorphism; *1*1, *1*2 and *2*2 represent non-risk homozygous genotype, heterozygous genotype, and risk homozygous genotype, respectively; HWE: Hardy-Weinberg equilibrium.

“ADH2” or “alcohol dehydrogenase 2”, “polymorphism” or “variant”, “hepatocellular carcinoma”, “HCC” or “liver”. Studies were included according to the following inclusion criteria: (1) evaluating the association between ADH2 Arg47His and HCC; (2) case-control study; (3) with sufficient data for evaluating odds ratios (ORs) with 95% confidence intervals (95% CIs). When numerous articles were published with overlapping data, only the largest or most recent studies were included. The literature search was updated on December 17, 2014.

Data extraction

All the following data were extracted by two independent investigators: name of first author, publication date, ethnicity, country of origin, number of cases and controls, genotyping methods, genotype frequencies, and Hardy-Weinberg equilibrium (HWE), as displayed in **Table 1**.

Statistical analysis

Crude ORs with 95% confidence intervals (95% CIs) were used to evaluate the correlation between ADH2 Arg47His polymorphism and HCC risk. The pooled ORs were analyzed under the following 5 genetic models: His/His vs. Arg/Arg, His/His + Arg/His vs. Arg/Arg, His/His vs. Arg/Arg + Arg/His, His vs. Arg, and Arg/His vs. Arg/Arg. Z test was used to estimate whether the pooled ORs were significant, and $P < 0.05$ was considered to be statistically significant. Q test was used to testify between-study heterogeneity. The pooled ORs were calculated by the fixed-effects model or random-effects model in the presence ($P \leq 0.10$) or absence ($P > 0.10$) of heterogeneity. Sensitivity analysis was performed to estimate the stability of the results. Begg's funnel plots and Egger's test were used to testify publication bias. HWE was checked by χ^2 test. Statistical analysis was conducted using

STATA version 12.0 (Stata Corporation, College Station, TX, USA).

Results

Study characteristics

A total of 150 studies were identified through databases in which 65 studies were precluded for unrelated titles and abstracts, 56 studies were excluded for duplicate publications, and 25 studies were precluded for no case-control studies. And finally 518 cases and 607 controls were included in our study.

Quantitative synthesis

As displayed in **Table 2** and **Figure 1**, there was no significant association between ADH2 Arg47His polymorphism and HCC risk (His/His vs. Arg/Arg: OR=0.99, 95% CI=0.79-1.25; His/His + Arg/His vs. Arg/Arg: OR=1.01, 95% CI=0.86-1.20; His/His vs. Arg/Arg + Arg/His: OR=0.90, 95% CI=0.74-1.11; His vs. Arg: OR=0.98, 95% CI=0.86-1.11; Arg/His vs. Arg/Arg: OR=1.05, 95% CI=0.82-1.34).

Sensitivity analysis

Sensitivity analysis was conducted to evaluate the influence of each individual study on the pooled ORs. The overall results were not altered when any individual study was excluded, suggesting our results were statistically steady.

Publication bias

Begg's funnel plot and Egger's test were performed to evaluate the publication bias. The shape of the funnel plot seemed symmetrical. Additionally, no significant publication bias was detected by Egger's test in the meta-analysis ($P = 0.64$). Therefore, there existed no apparent publication bias and the results were statistically credible (**Figure 2**).

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Table 2. Association of *ADH2* Arg47His polymorphism and HCC risk

	His/His vs. Arg/Arg		His/His+Arg/His vs. Arg/Arg		His/His vs. Arg/Arg		His vs. Arg		Arg/His vs. Arg/Arg	
	OR (95% CI)	<i>Ph</i>	OR (95% CI)	<i>Ph</i>	OR (95% CI)	<i>Ph</i>	OR (95% CI)	<i>Ph</i>	OR (95% CI)	<i>Ph</i>
Fixed-effects model										
Total	0.99 (0.79, 1.25)	0.989	1.01 (0.86, 1.20)	0.995	0.90 (0.74, 1.11)	0.193	0.98 (0.86, 1.11)	0.830	1.05 (0.82, 1.34)	0.964

Ph: *P*-value of heterogeneity test; *1*1, *1*2 and *2*2 represent non-risk homozygous genotype, heterozygous genotype, and risk homozygous genotype, respectively.

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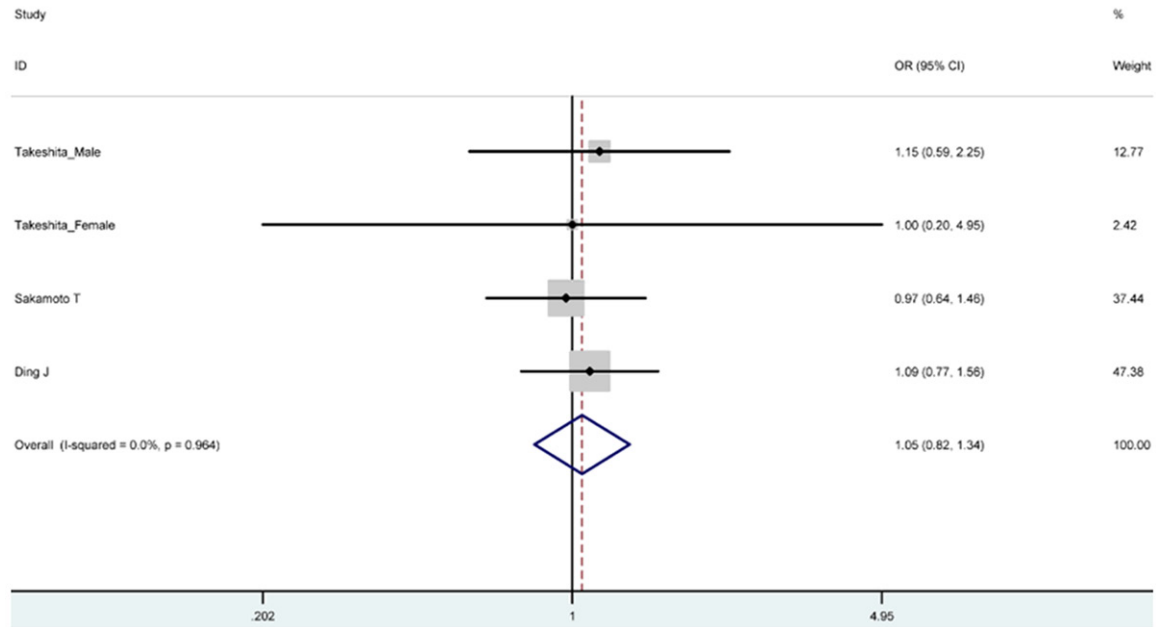


Figure 1. Forest plot analysis of association of *ADH2* Arg47His polymorphism and HCC risk under Arg/His vs. Arg/Arg genetic model.

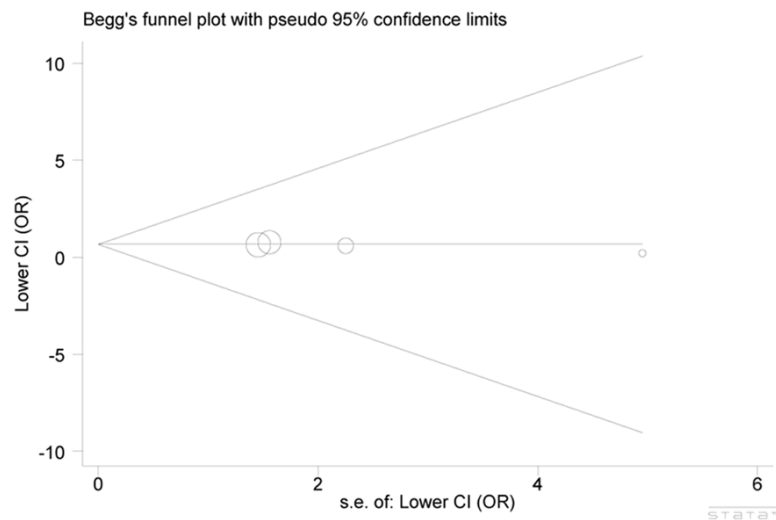


Figure 2. Publication bias test.

Discussion

HCC is one of the most common cancers in Asian populations [25]. Although some measures have been taken to improve the diagnosis of HCC, the results are still unsatisfactory due to the unclear etiology [26-29]. It has been demonstrated that both environmental and genetic factors may play significant roles in the etiology of HCC [4, 11]. The risk of HCC associ-

ated with exposure to exogenous xenobiotics or endogenous substances may be modified by genetic variations in metabolic detoxification activities. Therefore, the effects of genetic polymorphisms on HCC risk cannot be ignored.

Some epidemiological studies have investigated the relationship between *ADH2* Arg47His polymorphism and HCC susceptibility. In a large-scale study conducted by Ding et al., HCC patients were divided into drinkers and non-drinkers. Compared to the non-drink-

ers with active *ADH2* genotypes, the drinkers with inactive *ADH2* genotypes showed no high risk for HCC, thus the results revealed that *ADH2* Arg47His polymorphism was not related with increased risk of HCC [23]. Similar to the above study, Sakamoto T et al. divided HCC patients into non-drinkers, light, moderate, and heavy drinkers and they drew a same conclusion on the relationship of *ADH2* Arg47His and HCC [25].

Our meta-analysis, including 518 cases and 607 controls, was conducted to obtain a more precise assessment on the relationship between *ADH2* Arg47His polymorphism and HCC risk. And the results demonstrated that *ADH2* Arg47His polymorphism was not associated with HCC risk.

Some limitations in our study should be addressed. Firstly, our analysis was performed based on Asians, without considering other ethnic groups. Secondly, our study ignored the effects of gender on HCC risk. Thirdly, the sample size was relatively small. Finally, the results were summarized with unadjusted estimates, which might affect the validity of results. Therefore, further well-designed investigations performed in a larger scale are needed to clarify this point of view.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Guohua Yu, Clinical Oncology Department, Weifang People's Hospital, Kuiwen District, Weifang 261041, Shandong, China. E-mail: ghyry@126.com

References

- [1] Yi HM, Zhang W, Ai X, Li KY and Deng YB. Radiofrequency ablation versus surgical resection for the treatment of hepatocellular carcinoma conforming to the Milan criteria: systemic review and meta-analysis. *Int J Clin Exp Med* 2014; 7: 3150-3163.
- [2] Sabokrouh A, Goodarzi MT, Vaisi-Raygani A, Khatami S and Taghizadeh-Jahed M. Effects of treatment with platinum azidothymidine and azidothymidine on telomerase activity and bcl-2 concentration in hepatocellular carcinoma-induced rats. *Avicenna J Med Biotechnol* 2014; 6: 200-209.
- [3] Bazine A, Fetohi M, Berri MA, Essaadi I, Elbakraoui K, Ichou M and Errihani H. Spinal cord ischemia secondary to transcatheter arterial chemoembolization for hepatocellular carcinoma. *Case Rep Gastroenterol* 2014; 8: 264-269.
- [4] Chuang SC, La Vecchia C and Boffetta P. Liver cancer: descriptive epidemiology and risk factors other than HBV and HCV infection. *Cancer Lett* 2009; 286: 9-14.
- [5] Tanaka K, Hirohata T, Fukuda K, Shibata A, Tsukuma H and Hiyama T. Risk factors for hepatocellular carcinoma among Japanese women. *Cancer Causes Control* 1995; 6: 91-98.
- [6] Tanaka K, Hirohata T, Koga S, Sugimachi K, Kanematsu T, Ohryohji F, Nawata H, Ishibashi H, Maeda Y, Kiyokawa H and et al. Hepatitis C and hepatitis B in the etiology of hepatocellular carcinoma in the Japanese population. *Cancer Res* 1991; 51: 2842-2847.
- [7] Shibata A, Fukuda K, Nishiyori A, Ogimoto I, Sakata R and Tanikawa K. A case-control study on male hepatocellular carcinoma based on hospital and community controls. *J Epidemiol* 1998; 8: 1-5.
- [8] Tsukuma H, Hiyama T, Oshima A, Sobue T, Fujimoto I, Kasugai H, Kojima J, Sasaki Y, Imaoka S, Horiuchi N, et al. A case-control study of hepatocellular carcinoma in Osaka, Japan. *Int J Cancer* 1990; 45: 231-236.
- [9] Fukuda K, Shibata A, Hirohata I, Tanikawa K, Yamaguchi G and Ishii M. A hospital-based case-control study on hepatocellular carcinoma in Fukuoka and Saga Prefectures, northern Kyushu, Japan. *Jpn J Cancer Res* 1993; 84: 708-714.
- [10] Jiang W, Huang H, Ding L, Zhu P, Saiyin H, Ji G, Zuo J, Han D, Pan Y, Ding D, Ma X, Zhang Y, Wu J, Yi Q, Liu JO, Dang Y and Yu L. Regulation of cell cycle of hepatocellular carcinoma by NF90 through modulation of cyclin E1 mRNA stability. *Oncogene* 2014; [Epub ahead of print].
- [11] Hu Q, Lou GG, Liu YC, Qian L and Lv BD. The Tumor Necrosis Factor-alpha-308 and -238 Polymorphisms and Risk of Hepatocellular Carcinoma for Asian Populations: A Meta-Analysis. *Curr Ther Res Clin Exp* 2014; 76: 70-75.
- [12] Jelski W, Orywal K, Laniewska M and Szmitkowski M. The diagnostic value of alcohol dehydrogenase (ADH) isoenzymes and aldehyde dehydrogenase (ALDH) measurement in the sera of gastric cancer patients. *Clin Exp Med* 2010; 10: 215-219.
- [13] Jelski W and Szmitkowski M. Alcohol dehydrogenase (ADH) and aldehyde dehydrogenase (ALDH) in the cancer diseases. *Clin Chim Acta* 2008; 395: 1-5.
- [14] Duell EJ, Sala N, Travier N, Munoz X, Boutron-Ruault MC, Clavel-Chapelon F, Barricarte A, Arriola L, Navarro C, Sanchez-Cantalejo E, Quiros JR, Krogh V, Vineis P, Mattiello A, Tumino R, Khaw KT, Wareham N, Allen NE, Peeters PH, Numans ME, Bueno-de-Mesquita HB, van Oijen MG, Bamia C, Benetou V, Trichopoulos D, Canzian F, Kaaks R, Boeing H, Bergmann MM, Lund E, Ehrnstrom R, Johansen D, Hallmans G, Stenling R, Tjonneland A, Overvad K, Ostergaard JN, Ferrari P, Fedirko V, Jenab M, Nesi G, Riboli E and Gonzalez CA. Genetic variation in alcohol dehydrogenase (ADH1A, ADH1B, ADH1C, ADH7) and aldehyde dehydrogenase (ALDH2), alcohol consumption and gastric cancer risk in the European Prospective Investiga-

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- tion into Cancer and Nutrition (EPIC) cohort. *Carcinogenesis* 2012; 33: 361-367.
- [15] Haseba T, Kameyama K, Mashimo K and Ohno Y. Dose-Dependent Change in Elimination Kinetics of Ethanol due to Shift of Dominant Metabolizing Enzyme from ADH 1 (Class I) to ADH 3 (Class III) in Mouse. *Int J Hepatol* 2012; 2012: 408190.
- [16] Lao-Sirieix P, Caldas C and Fitzgerald RC. Genetic predisposition to gastro-oesophageal cancer. *Curr Opin Genet Dev* 2010; 20: 210-217.
- [17] Norppa H, Tursi F, Pfaffli P, Maki-Paakkanen J and Jarventaus H. Chromosome damage induced by vinyl acetate through in vitro formation of acetaldehyde in human lymphocytes and Chinese hamster ovary cells. *Cancer Res* 1985; 45: 4816-4821.
- [18] Singh NP and Khan A. Acetaldehyde: genotoxicity and cytotoxicity in human lymphocytes. *Mutat Res* 1995; 337: 9-17.
- [19] Aylward B, Lloyd J, Zaffran M, McNair-Scott R and Evans P. Reducing the risk of unsafe injections in immunization programmes: financial and operational implications of various injection technologies. *Bull World Health Organ* 1995; 73: 531-540.
- [20] Chiang CP, Jao SW, Lee SP, Chen PC, Chung CC, Lee SL, Nieh S and Yin SJ. Expression pattern, ethanol-metabolizing activities, and cellular localization of alcohol and aldehyde dehydrogenases in human large bowel: association of the functional polymorphisms of ADH and ALDH genes with hemorrhoids and colorectal cancer. *Alcohol* 2012; 46: 37-49.
- [21] Zhang L, Jiang Y, Wu Q, Li Q, Chen D, Xu L, Zhang C, Zhang M and Ye L. Gene-environment interactions on the risk of esophageal cancer among Asian populations with the G48A polymorphism in the alcohol dehydrogenase-2 gene: a meta-analysis. *Tumour Biol* 2014; 35: 4705-4717.
- [22] Ding J, Li S, Wu J, Gao C, Zhou J, Cao H, Su PS, Liu Y, Zhou X and Chang J. Alcohol dehydrogenase-2 and aldehyde dehydrogenase-2 genotypes, alcohol drinking and the risk of primary hepatocellular carcinoma in a Chinese population. *Asian Pac J Cancer Prev* 2008; 9: 31-35.
- [23] Takeshita T, Yang X, Inoue Y, Sato S and Morimoto K. Relationship between alcohol drinking, ADH2 and ALDH2 genotypes, and risk for hepatocellular carcinoma in Japanese. *Cancer Lett* 2000; 149: 69-76.
- [24] Sakamoto T, Hara M, Higaki Y, Ichiba M, Horita M, Mizuta T, Eguchi Y, Yasutake T, Ozaki I, Yamamoto K, Onohara S, Kawazoe S, Shigematsu H, Koizumi S and Tanaka K. Influence of alcohol consumption and gene polymorphisms of ADH2 and ALDH2 on hepatocellular carcinoma in a Japanese population. *Int J Cancer* 2006; 118: 1501-1507.
- [25] Szumera-Cieckiewicz A, Olszewski WT, Piech K, Glogowski M and Prochorec-Sobieszek M. Endobronchial metastasis from hepatocellular carcinoma—a case description with literature review. *Int J Clin Exp Pathol* 2013; 6: 1942-1947.
- [26] Tsoulfas G, Agorastou P, Tooulis A and Marakis GN. Current and future challenges in the surgical treatment of hepatocellular carcinoma: a review. *Int Surg* 2014; 99: 779-786.
- [27] Li Q, Zhu LZ, Yang RJ and Zhu X. Cytotoxic activity of anticancer drugs on hepatocellular carcinoma cells in hypoxic-hyponutritional culture. *Int Surg* 2014; 99: 745-752.
- [28] Kan H, Guo W, Huang Y and Liu D. MicroRNA-520g induces epithelial-mesenchymal transition and promotes metastasis of hepatocellular carcinoma by targeting SMAD7. *FEBS Lett* 2015; 589: 102-9.
- [29] Zhou L, Rui JA, Wang SB, Chen SG and Qu Q. Risk factors of microvascular invasion, portal vein tumor thrombosis and poor post-resectional survival in HBV-related hepatocellular carcinoma. *Hepatogastroenterology* 2014; 61: 1696-1703.