Original Article Correlation between GATA4 gene polymorphism and congenital heart disease

Xue-Yong Yang, Xiao-Yong Jing, Zhe Chen, Ying-Long Liu

The Fourth Ward, Beijing Anzhen Hospital, Capital Medical University, Beijing 100029, China

Received May 16, 2015; Accepted July 6, 2015; Epub September 15, 2015; Published September 30, 2015

Abstract: Objective: The correlation between GATA4 gene polymorphism and congenital heart disease (CHD) was analyzed. Method: Clinical data and blood samples were collected from 350 CHD patients who were treated at the Department of Cardiology in Beijing Anzhen Hospital. The control group consisted of 350 healthy subjects receiving physical examination at our hospital during the same period. Polymorphism was detected by polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) analysis for locus rs11392441 of GATA4 gene. Results: Polymorphism of locus rs1139244 of GATA4 gene was detected in CHD patients. The distribution frequencies of GG genotype and G allele were significantly higher than those of the control group. Conclusion: Polymorphism of locus rs1139244 of GATA4 gene was correlated with CHD.

Keywords: Congenital heart disease (CHD), GATA4 gene, polymorphism

Introduction

Congenital heart disease (CHD) is the most common birth defect that seriously threatens the infants' health. Having the main manifestations of atrial septal defect (ASD), ventricular septal defect (VSD) and patent ductus arteriosus (PDA), CHD presents as the abnormality in the structure and function of heart and great vessels caused by embryonic development disorders [1-3]. The incidence of CHD in foreign reports is 5.4%-16.1% [4], while that of China is 2.51% [5]. The pathogenesis of CHD is highly complex and is not fully understood yet. It is currently accepted that CHD is a polygenic disease resulting from the interaction between environmental and genetic factors [6]. Connections have been established between normal development of heart and multiple transcription factors. Any mutation in transcription factor genes may lead to abnormal cardiac development and CHD. Researchers have found that CHD may be related to NKX2.5 [7], TBX5 [8] and JARID2 genes [9]. Zinc finger transcription factor GATA4 is the zinc finger domain that binds to specific DNA. GATA4 is highly conserved during evolution and implicated in cardiac stem cell differentiation, cyclization, heart block and conduction. GATA4 gene is localized to 8p23.1 with full length of cDNA being 3371 bp, consisting of 6 exons and encoding 422 amino acids [10]. GATA4 gene expression has a direct impact on the embryonic development of heart. Rat fetus with CHD dies at the gestational age of 13.5-16.5 d. GATA4 gene mutation has a close connection with ASD [11], an autosomal dominant disease. Some patients may be combined with VSD, pulmonary artery stenosis (PS), severe diastolic dysfunction and atrioventricular valve regurgitation, but not with contractile dysfunction or lesions of the cardiac conduction system and coronary heart disease [11-13].

We investigated the single nucleotide polymorphism (SNP) locus P66A (rs1139244) in the coding region of GATA4 gene and detected for its connections with CHD risk.

Materials and methods

General data

Three hundred and fifty CHD patients who were treated at Department of Cardiology at Anzhen Hospital from January 2005 to December 2014 were selected to constitute the CHD group.

Locus	Primer sequence	Products (bp)	Tm (°C)	Endonuclease
Rs1139224	Sense: 5'-ACGCGGCCCCACCGGAGCTGCCGC-3'	314	56	SFaNI
	Antisense: 5'-TCGTTGTTGCCGTGGTTTTC-3'			

Groups	Ν	Genotype (n, %)			Р	Allele (n, %)		OR (95% CI)	Р
		CC	CG	GG	-	С	G		
Control group	350	51 (14.6)	165 (47.1)	134 (38.3)	0.002	267 (38.1)	433 (61.9)	1.33 (1.06-1.65)	0.011
CHD group	350	22 (6.3)	178 (50.9)	150 (42.8)		222 (31.7)	478 (68.3)		

Table 3. Frequencies of GG genotype in various types of CHD and in the controls

Groups	N	Genotype			Р	Allele		Р
		CC	CG	GG	-	С	G	-
Control group	350	51 (14.6)	165 (47.1)	134 (38.3)		267 (38.1)	433 (61.9)	
ASD	121	10 (8.3)	75 (62.0)	36 (29.7)	0.015	95 (39.2)	147 (60.8)	0.759
VSD	132	8 (6.0)	78 (59.0)	46 (35.0)	0.013	94 (35.6)	170 (64.4)	0.468
PDA	68	2 (2.9)	15 (22.1)	51 (75.0)	<0.001	19 (13.9)	117 (86.1)	<0.001
PS	29	2(6.9)	10 (34.5)	17 (58.6)	0.089	14 (24.1)	44 (75.9)	0.034

They were 137 males and 213 females with an average age of 10.4 ± 8.2 years old. The normal control group had 350 subjects, including 161 males and 189 females with an average age of 9.6±6.3 years old. All cases were matched for age and had no blood relationship with each other. The two groups did not differ in age and sex ratio significantly (*P*>0.05).

Inclusion criteria: (1) Presenting with clinical features of CHD and having echocardiographic evidences or surgical records; (2) Signing of informed consent.

Exclusion criteria: (1) Presenting with clinical features of CHD but having no echocardio-graphic evidences or other examination results; (2) Confirmed chromosomal abnormalities or syndrome-associated cardiovascular abnormality.

The protocol conformed to medical ethics and was approved by Medical Ethics Committee. Informed consent was obtained from all subjects.

Primer design

SNP database was searched for SNPs in the coding region of GATA4 gene. SNP locus P66A (rs11392441) was selected and primers were designed using Primer 5. The primer sequences are shown in **Table 1**.

Genome DNA extraction

For all cases 2-ml of venous blood was collected with citrate anticoagulation and preserved at -20°C. Genome DNA extraction was performed conventionally using Tris saturated phenol-chloroform method described previously [14].

PCR amplification of target fragment

PCR amplification was carried out conventionally. The 25 μ l reaction system consisted of genome DNA (20 ng/ μ l) 2 μ l, 10× buffer (MgCl²⁺ 15 mmoL/L) 2.5 μ l, dNTP 1.0 μ l, upstream primer 0.5 μ l, downstream primer 0.5 μ l, TaqDNA polymerase 0.2 μ l and sterilized water 18.3 μ l. PCR reaction conditions: denaturation at 94°C for 5 min, 94°C for 45 s, 56-60°C for 45 s, 72°C for 45 s, 35 cycles, final extension at 72°C for 10 min. Genotyping was performed using PCR-RFLP. Restriction enzyme is SFaNI, the conditions and products of enzyme digestion for the genotyping of locus rs11392441 in GATA4 gene was described previously [15].

Statistical methods

Statistical analysis was carried out using SPSS software 17.0. Whether the genotype frequency satisfied Hardy-Weinberg equilibrium was examined with X^2 test. Differences in genotype

frequency and allele frequency were compared by using X^2 test between the groups, and P<0.05 was considered as statistically significant.

Results

Genotype frequency and allele frequency at locus rs1139244 in GATA4 gene

Genotype frequencies and allele frequencies at locus P66A (rs1139244) in GATA4 gene for CHD group and control group are shown in Table 2. It can be seen that the frequencies satisfied Hardy-Weinberg equilibrium. The frequency of homozygous GG genotype in GATA4 gene was 38.3% and 42.8% in the control group and CHD group, respectively, showing significant differences (P<0.05). The frequency of G allele in GATA4 gene was 61.9% and 68.3% in the control group and CHD group, respectively. A difference of statistical significance was found between the two groups (P<0.05). The CHD risk was significant increased 1.33 fold in the carriers of C allele compared to G allele carriers (OR=1.33, 95% CI: 1.06-1.65; P=0.011).

Frequencies of GG genotype in various types of CHD and in the controls

The frequencies of GG genotype at locus rs1139244 in GATA4 gene were significantly higher in cases with atrial septal defect (ASD), ventricular septal defect (VSD), and PDA than in the controls (**Table 3**).

Discussion

In the present study, we found rs1139244 in GATA4 gene were significantly associated with CHD in Chinese population, especially for ASD, VSD, and FDA.

GATA4 is a candidate gene for the occurrence of cardiovascular abnormalities, belonging to GATA transcription gene family. Containing zinc finger domain, GATA4 is a highly conserved transcription gene [16] and interacts with other transcriptional regulatory gene during early stage of heart development. Over 30 gene promoters in myocardial cells are directly regulated by GATA4. GATA4-related signaling transduction pathway plays a key role in specific differentiation and transport of cardiac stem cells

after heart formation [17]. GATA4-knockout mice failed to develop primitive heart tube [18]. This indicates that GATA4 mainly controls the formation of cardiac progenitor cells in the endodermal layer. However, functional deficit of GATA4 may lead to abnormal differentiation of endodermal cells and hence heart defect. Some foreign studies have reported chromosome 8p deletion in syndromes associated with heart defect, and GATA4 deletion may be the pathogenic reason. Mutations in GATA4 can cause abnormal cardiac development, including E359dd (frameshift mutation), G296S (missense mutation) and 1074delC (frameshift mutation) [19]. But the correlation between GATA4 gene polymorphism and CHD has not been reported. We performed PCR-RFLP for SNP locus P66A (rs1139244) in the coding region of GATA4 gene among 350 CHD cases and 350 normal controls. SNPs and haplotypes were analyzed. Results showed that the frequencies conformed to Hardy-Weinberg equilibrium and were representative of the population. GG genotype frequency in CHD cases and normal controls was 42.8% and 38.3%, respectively, and G allele frequency was 68.3% and 61.9%, respectively, both frequencies showing significant differences between the groups. It was indicated that G allele is a susceptible factor of CHD, and its mutation can lead to CHD. Moreover, the genotype frequency at locus P66A (rsll39244) was significantly higher in cases with ASD, VSD, and PDA than in normal controls (P<0.05). No obvious differences were found for other types of CHD compared with the control.

In conclusion, the polymorphism at locus P66A (rs1139244) in GATA4 gene is related to congenital ASD, VSD, and PDA. Therefore, GATA4 genetic polymorphism can be useful marker for the diagnosis of CHD. The findings of the present study need to be further verified by largesample trials.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Ying-Long Liu, The Fourth Ward, Beijing Anzhen Hospital, Capital Medical University, 2 Anzhen Road, Chaoyang District, Beijing 100029, China. Tel: +86-1064456534; Fax: +86-1064456534; E-mail: liuy_long@163.com

References

- Zhang QJ, Liu ZP. Histone methylations in heart development, congenital and adult heart diseases. Epigenomics 2015; 7: 321-30.
- [2] Lee KJ, Seto W, Benson L, Chaturvedi RR. Pharmacokinetics of sirolimus-eluting stents implanted in the neonatal arterial duct. Circ Cardiovasc Interv 2015; 8: e002233.
- [3] Colen T, Smallhorn JF. Three-dimensional echocardiography for the assessment of atrioventricular valves in congenital heart disease: past, present and future. Semin Thorac Cardiovasc Surg Pediatr Card Surg Annu 2015; 18: 62-71.
- [4] Sadowski SL. Congenital cardiac disease in the newborn infant: past, present, and future. Crit Care Nurs Clin North Am 2009; 21: 37-48.
- [5] Arias López I, Martínez Tallo E, Campo Sanpedro F, Cardesa García JJ. Incidence and clinical characteristics of congenital heart disease in Badajoz Province, Spain. An Pediatr (Barc) 2008; 69: 23-7.
- [6] Wang X, Li P, Chen S, Xi L, Guo Y, Guo A, Sun K. Influence of genes and the environment in familial congenital heart defects. Mol Med Rep 2014; 9: 695-700.
- [7] Qu XK, Qiu XB, Yuan F, Wang J, Zhao CM, Liu XY, Zhang XL, Li RG, Xu YJ, Hou XM, Fang WY, Liu X, Yang YQ. A novel NKX2.5 loss-of-function mutation associated with congenital bicuspid aortic valve. Am J Cardiol 2014; 114: 1891-5.
- [8] Hoffmann AD, Yang XH, Burnicka-Turek O, Bosman JD, Ren X, Steimle JD, Vokes SA, McMahon AP, Kalinichenko VV, Moskowitz IP. Foxf genes integrate tbx5 and hedgehog pathways in the second heart field for cardiac septation. PLoS Genet 2014; 10: e1004604.
- [9] Barth JL, Clark CD, Fresco VM, Knoll EP, Lee B, Argraves WS, Lee KH. Jarid2 is among a set of genes differentially regulated by Nkx2.5 during outflow tract morphogenesis. Dev Dyn 2010; 239: 2024-33.
- [10] Orjuela Quintero DC, Núñez F, Caicedo V, Pachón S, Salazar Salazar M. Mutations in the GATA4 gen in patients with non-syndromic congenital heart disease. Invest Clin 2014; 55: 207-16.
- [11] Xiang R, Fan LL, Huang H, Cao BB, Li XP, Peng DQ, Xia K. A novel mutation of GATA4 (K319E) is responsible for familial atrial septal defect and pulmonary valve stenosis. Gene 2014; 534: 320-3.

- [12] Warburton D, Ronemus M, Kline J, Jobanputra V, Williams I, Anyane-Yeboa K, Chung W, Yu L, Wong N, Awad D, Yu CY, Leotta A, Kendall J, Yamrom B, Lee YH, Wigler M, Levy D. The contribution of de novo and rare inherited copy number changes to congenital heart disease in an unselected sample of children with conotruncal defects or hypoplastic left heart disease. Hum Genet 2014; 133: 11-27.
- [13] Xiong F, Li Q, Zhang C, Chen Y, Li P, Wei X, Li Q, Zhou W, Li L, Shang X, Xu X. Analyses of GATA4, NKX2.5, and TFAP2B genes in subjects from southern China with sporadic congenital heart disease. Cardiovasc Pathol 2013; 22: 141-5.
- [14] Maw MT, Yamaguchi T, Kasanga CJ, Terasaki K, Fukushi H. A practical tissue sampling method using ordinary paper for molecular detection of infectious bursal disease virus RNA by RT-PCR. Avian Dis 2006; 50: 556-60.
- [15] Orjuela Quintero DC, Núñez F, Caicedo V, Pachón S, Salazar Salazar M. Mutations in the GATA4 gen in patients with non-syndromic congenital heart disease. Invest Clin 2014; 55: 207-16.
- [16] Shaw-Smith C, De Franco E, Lango Allen H, Batlle M, Flanagan SE, Borowiec M, Taplin CE, van Alfen-van der Velden J, Cruz-Rojo J, Perez de Nanclares G, Miedzybrodzka Z, Deja G, Wlodarska I, Mlynarski W, Ferrer J, Hattersley AT, Ellard S. GATA4 mutations are a cause of neonatal and childhood-onset diabetes. Diabetes 2014; 63: 2888-94.
- [17] Bisping E, Ikeda S, Kong SW, Tarnavski O, Bodyak N, McMullen JR, Rajagopal S, Son JK, Ma Q, Springer Z, Kang PM, Izumo S, Pu WT. Gata4 is required for maintenance of postnatal cardiac function and protection from pressure overload-induced heart failure. Proc Natl Acad Sci U S A 2006; 103: 14471-6.
- [18] Looyenga BD, Hammer GD. Origin and identity of adrenocortical tumors in inhibin knockout mice: implications for cellular plasticity in the adrenal cortex. Mol Endocrinol 2006; 20: 2848-63.
- [19] Schluterman MK, Krysiak AE, Kathiriya IS, Abate N, Chandalia M, Srivastava D, Garg V. Screening and biochemical analysis of GATA4 sequence variations identified in patients with congenital heart disease. Am J Med Genet A 2007; 143A: 817-23.