Original Article

MicroRNA26 increases sensitivity of cutaneous squamous cell carcinoma to dacarbazine

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Abstract: Cutaneum carcinoma threatens health and lives of patients. Dacarbazine is one of the most important therapeutics for cutaneum carcinoma. Current study was focused on molecular mechanism of sensibilization effect of microRNA26 on sensitivity of cutaneous squamous cell carcinoma to dacarbazine. Colon16 (cell line of cutaneous squamous cell carcinoma to dacarbazine. Colon16 (cell line of cutaneous squamous cell carcinoma were collected to compare expression of microRNA26. Synthetic microRNA26 and control microRNA were transfected into Colon16 with lipofection transfection, respectively. All Colon16 cells were treated with dacarbazine. The growth and apoptosis of Colon16 were examined by flow cytometry. RT-PCR was performed to examine expression level of microRNA26. Growth of Colon16 was significantly inhibited after dacarbazine treatment, while apoptosis and expression of microRNA26 increased. Compared with para-carcinoma tissue, cutaneous squamous cell carcinoma had a higher level of microRNA26. Overexpression of microRNA26 promoted dacarbazine-induced apoptosis of Colon16. Dacarbazine induced apoptosis of cutaneous squamous cell carcinoma via increasing expression of microRNA26, and microRNA26 increased sensitivity of cutaneous squamous cell carcinoma to dacarbazine.

Keywords: Dacarbazine, microRNA26, Colon16, apoptosis, cutaneous squamous cell carcinoma

Introduction

Cutaneous squamous cell carcinoma is a malignant skin disease caused by mutations and malignant hyperplasia of skin cells [1]. Cutaneum carcinoma threatens health and lives of patients [2]. Recent study indicated underlying mechanism of cutaneous squamous cell carcinoma included cell prosoplasia, increasing proliferation and inhibited apoptosis [3]. Cutaneous squamous cell carcinoma not only results in aberrant accumulation of cutaneous squamous cells ,but also lead to infiltration of cancer cells in multiple organs, including vagina, cervix, esophagus, lips and oral cavity [4, 5]. Surgery is the main treatment for early-stage cutaneous squamous cell carcinoma, while advanced cutaneous squamous cell carcinoma is treated by non-operation therapy, including chemotherapy, radiotherapy and comprehensive therapy. The therapeutic mechanism of chemotherapy is inhibiting cancer proliferation and promoting apoptosis via chemicals intervention [6].

Dacarbazine (DTIC) is an important chemotherapeutic widely used in clinical scenario, and has promising efficacy for malignant lymphoma, melanoma and soft tissue tumor [7]. Latest research showed combination of dacarbazine and other adjuvant therapies had an ideal efficacy for cutaneous squamous cell carcinoma, meanwhile, animal experiment demonstrated combination of dacarbazine and specific microRNA increased sensitivity of cutaneous squamous cells to dacarbazine [8]. However, molecular mechanism underlying dacarbazine efficacy for cutaneous squamous cell carcinoma was still unclear.

MicroRNA is a kind of small RNA fragments with a length of 17 bp to 27 bp. Although microRNA does not encode any proteins, it has many biology functions [9, 10], including cell viability, proliferation [11], apoptosis [12], cell cycle [13], autophagy [14], and intracellular signal transduction [15]. In the field of cutaneous squamous cell carcinoma, studies about microRNA are still scarce [16].

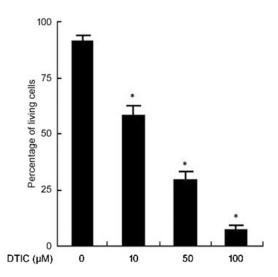


Figure 1. Growth of Colon16 was inhibited after dacarbazine treatment. DTIC represents dacarbazine. *P<0.05 versus control group (leftmost panel). All the experiments were performed three times yielding similar results and comprised of biological replicates. For all the figures, the mean ± standard deviation (s.d.) is represented unless otherwise noted.

Current study was aimed to explore not only regulatory mechanism underlying effect of dacarbazine on Colon16, but alsomolecular mechanism of how microRNA26 influenced sensitivity of Colon16 to dacarbazine so as to provide theory basis for clinical practice.

Materials and methods

Reagents and materials

Skin squamous carcinoma cells Colon16 were preserved in the laboratory. MTT used for detection of cell viability, as well as high glucose culture DMEM and fetal bovine serum were acquired from the Beijing Dingguo Biological Co., Ltd (Beijing, China). FITC-Annexin-V used for analysis of apoptosis and cell transfection reagents liposomes were provided by Beyotime biological technological research institute (Jiangsu, Haimen). Reverse transcription kit was purchased from Tiangen biological co., LTD. microRNA26 and control microRNA were synthesized by Jima biological co., LTD, sequences were shown as 5'ACAGTACGACG-TGTAGCA3' and 5'ATGACGACAGACACGACT3'. Skin squamous cell carcinoma tissue and paracarcinorma tissue were provided by dermatology in our hospital and approved by our hospital ethics committee; the informed forms were

signed by all skin squamous cell carcinoma patients.

Culture of skin squamous carcinoma cells Colon16

Skin squamous carcinoma cells were quickly transferred from liquid nitrogen to 37°C water bath for thawing, then centrifuging at 1000 rpm for 5 min, removing supernatant and resuspended in new DMEM and cultured at 37°C , 5% CO_{2} .

Transfection of skin squamous carcinoma cells Colon16

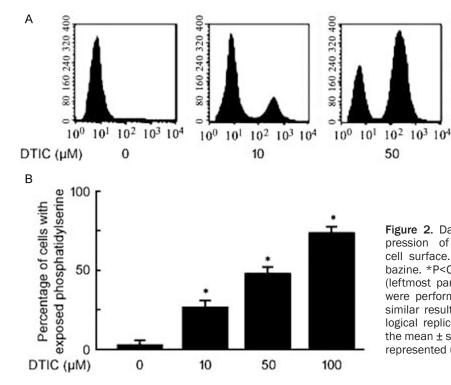
Skin squamous carcinoma cells cultured at 37° C, 5% CO $_2$ were reseeded in 96 wells plate (103/well) before transfection 24 h, when the density of cells reached at 80%, microRNA26 and control microRNA were diluted in culture by ratio of 1:100, mixing fully and stewing for 6 min. In the end, the mixture above was dropped in cell culture plate and shaken gently for fully reaction. The culture was changed after 24 h transfection.

MTT cell viability assay for skin squamous carcinoma cells Colon16

Preliminary test was performed based working concentration of dacarbazine [16], 10 μ M, 50 μ M and 100 μ M dacarbazine were used respectively to treat skin squamous carcinoma cells Colon16 for 14 h. Three parallel traits were designed. MTT (5 mg/ml, 30 μ l) reaction solution was added into three different skin squamous carcinoma cells Colon16, culturing at 37°C for 3 h. Washing cell above 3 times, and then DMSO was added into squamous carcinoma cells Colon16 to terminate reaction for 10 min. In the end, the optical density was detected for squamous carcinoma cells Colon16 at 492 nm in microplate reader.

Cell apoptosis assay

Skin squamous carcinoma cells Colon16 transfected with microRNA or treated with dacarbazine were selected, and cell apoptosis was detected by cytometer and caspase activity respectively. Cytometer were performed according to the introduction of kit, showed as following: adjusting the treated cell concentration at 105/ml, cell suspension was mixed with reac-



240 320 400 160 80 100 101 102 103 104 100

Figure 2. Dacarbazine increased expression of phosphatidylserine on cell surface. DTIC represents dacarbazine. *P<0.05 versus control group (leftmost panel). All the experiments were performed three times yielding similar results and comprised of biological replicates. For all the figures, the mean ± standard deviation (s.d.) is represented unless otherwise noted.

tion solution and FITC-Annexin V reagents by ratio of 250:50:1, reacting for 15 min at room temperature. Then expression of phosphatidylserine was detected in different skin squamous carcinoma cells Colon16, emission and absorb wavelength were 488 nm and 625 nm, respectively.

Activity assay were performed based the introduction of kit, specific steps shown as following: adjusting the treated cell concentration at 105/ml, cells transfected microRNA or treated with dacarbazine were lysed, then chromophoric substrate were added into the lysis, culturing 30 min at 37°C, in the end, absorbance value were assayed at 490 nm, which indicated the relative activity of caspase-3.

RT-PCR

Skin squamous carcinoma cells Colon16 transfected microRNA or treated with dacarbazine and Skin squamous carcinoma cells were collected, then total RNA was extracted by Trizol on the basis of the instruction of kit, then reversed transcript into cDNA, processed RT-PCR, microRNA26 and control microRNA primers were treated as primers, then the microRNA26 in skin squamous carcinoma cells Colon16 were detected.

Statistical method

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All results were analyzed with SPSS12.0. All the experiments were performed three times yielding similar results and comprised of biological replicates. For all the figures, the mean ± standard deviation (s.d.) is represented unless otherwise noted. Unpaired Student's t test or ANOVA with Tukey's comparison were used for paired and group comparisons, respectively. Single factor variance was applied in the intergroup analysis. P value<0.05 was considered to be statistically significant.

Results

Dacarbazine inhibited growth of Colon16

Our finding on MTT experiment demonstrated dacarbazine inhibited growth of Colon16 in a dose-dependent manner. Compared with Colon16 untreated by dacarbazine, growth of all dacarbazine-treated Colon16 cell lines was significantly inhibited (Figure 1). What is more, dacarbazine concentration was negative with growth of Colon16 (P<0.05).

Dacarbazine induced apoptosis of Colon16

Experiment results of Colon16 showed that dacarbazine resulted in activation of caspase-3

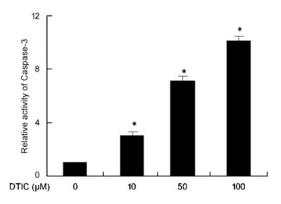


Figure 3. Dacarbazine resulted in activation of caspase-3. DTIC represents dacarbazine. *P<0.05 versus control group (leftmost panel). All the experiments were performed three times yielding similar results and comprised of biological replicates. For all the figures, the mean \pm standard deviation (s.d.) is represented unless otherwise noted.

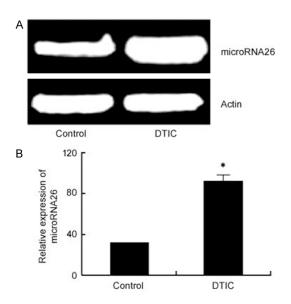


Figure 4. Expression level of microRNA26 was significantly increased in dacarbazine-treated Colon16. A. RT-PCR images. B. Analysis of microRNA26 expression. DTIC represents dacarbazine. The concentration of dacarbazine is 10 μ M. *P<0.05 versus control group (left panel). All the experiments were performed three times yielding similar results and comprised of biological replicates. For all the figures, the mean \pm standard deviation (s.d.) is represented unless otherwise noted.

and increased expression of phosphatidylserine on cell surface. Such effect of dacarbazine was also induced in a dose-dependent manner. Compared with Colon16 untreated by dacarbazine, apoptosis of all dacarbazine-treated Colon16 cell lines was significantly enhanced (P<0.05, Figures 2 and 3).

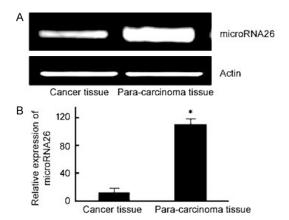


Figure 5. MicroRNA26 expression significantly decreased in cancer tissues from cutaneous squamous cell carcinoma patients. A. RT-PCR images. B. Analysis of microRNA26 expression. DTIC represents dacarbazine. *P<0.05 versus control group (left panel). All the experiments were performed three times yielding similar results and comprised of biological replicates. For all the figures, the mean ± standard deviation (s.d.) is represented unless otherwise noted.

Dacarbazine increased expression level of microRNA26 in Colon16

RT-PCR result showed, compared with control group, expression level of microRNA26 in Colon16 was significantly elevated after dacarbazine treatment (P<0.05, Figure 4). Such data indicated that microRNA26 increase might mediate dacarbazine-induced apoptosis of Colon16 (Figure 4).

MicroRNA26 expression significantly decreased in cancer tissues from cutaneous squamous cell carcinoma patients

Compared with para-carcinoma tissue, microR-NA26 expression of cutaneous squamous cell carcinoma tissues significantly decreased, verified by RT-PCR examination (P<0.05, Figure 5). Such data suggested low expression level of microRNA26 was associated with disease progress of cutaneous squamous cell carcinoma.

MicroRNA26 transfection enhanced dacarbazine-induced apoptosis of Colon16

Our finding on transfection experiment showed both microRNA26 transfection and control microRNA transfection did not cause apoptosis of Colon16 at baseline. However, compared with control microRNA transfection, dacarba-

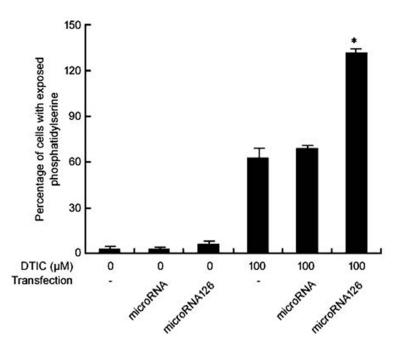


Figure 6. Overexpression of microRNA26 enhanced dacarbazine-induced apoptosis. DTIC represents dacarbazine. *P<0.05 versus control group (leftmost panel). All the experiments were performed three times yielding similar results and comprised of biological replicates. For all the figures, the mean \pm standard deviation (s.d.) is represented unless otherwise noted.

zine induced a higher apoptosis degree in microRNA26 transfected Colon16 (P<0.05, Figure 6), suggesting dacarbazine-induced apoptosis was associated with microRNA and overexpression of microRNA26 could enhance dacarbazine-induced apoptosis.

Discussion

Cutaneous squamous cell carcinoma is a world-wide disease, seriously impacting human health and lives [17]. Thus, to study this disease is of both theoretical significance and clinical value [18].

Dacarbazine plays a key role in clinical antineoplastic therapy [19]. However, it is unclear that how dacarbazine regulated cutaneous squamous cell carcinoma. Our study explored regulatory mechanism underlying effect of dacarbazine on cutaneous squamous cell carcinoma, verified by experiment conducted in cutaneous squamous cell carcinoma Colon16 cell line. We found that dacarbazine treatment inhibited growth of Colon16, induced apoptosis of Colon16. Moreover, such apoptosis inhibiting effect is dose-independent with dacarbazine, consistent with related studies in other types of cancers or other chemotherapeutics [20]. Three main findings were elucidated in our study. (1) Dacarbazine treatment suppressed growth of Colon16 and induced significant apoptosis of Colon16 with a marked increase of microRNA26 expression. (2) In patients with cutaneous squamous cell carcinoma, microRNA26 might be used as a predicting factor. (3) Overexpression of microR-NA26 further induced apoptosis of cutaneous squamous cell carcinoma with clinical significance. These findings indicated that dacarbazine not only limited cancer growth, but also induced apoptosis of cutaneous squamous cell carcinoma via increasing expression of microRNA26, which finally increased sensitivity of cutaneous squamous cell carcinoma to dacarbazine.

Though similar with previous studies [21], our study stilled demonstrated disagreement. We observed that overexpression of microRNA26 alone cannot inhibit Colon16, while enhance inhibiting effect when combined with dacarbazine. One possibility is that every type of microRNA regulated different cancer with different effect [22], and some microRNA can induce opposite effect under different condition [23, 24]. Meanwhile, details of experimental condition differ among studies [25-27]. Further studies are warranted to answer these questions.

Our study had three limitations. (1) Only cell model was established to explore effect of dacarbazine, thus, animal experiment need to be conducted in cutaneous squamous cell carcinoma model [28]. (2) We found dacarbazine increased expression of microRNA26, but it was not yet to be seen how expression of microRNA26 changed at different stage of cancer. (3) Our study did not elucidated detailed mechanism underlying how microRNA26 increased sensitivity of cutaneous squamous cell carcinoma to dacarbazine.

In conclusion, microRNA26 enhanced sensitivity of cutaneous squamous cell carcinoma to

dacarbazine, and dacarbazine promoted cancer apoptosis via increasing expression of microRNA26.

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Disclosure of conflict of interest

None.

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References

- [1] Yamazaki N, Uhara H, Fukushima S, Uchi H, Shibagaki N, Kiyohara Y, Tsutsumida A, Namikawa K, Okuyama R, Otsuka Y and Tokudome T. Phase II study of the immunecheckpoint inhibitor ipilimumab plus dacarbazine in Japanese patients with previously untreated, unresectable or metastatic melanoma. Cancer Chemother Pharmacol 2015; 76: 969-975.
- [2] Shih V, Ten Ham RM, Bui CT, Tran DN, Ting J and Wilson L. Targeted therapies compared to dacarbazine for treatment of BRAF(V600E) metastatic melanoma: a cost-effectiveness analysis. J Skin Cancer 2015; 2015: 505302.
- [3] Ribas A, Puzanov I, Dummer R, Schadendorf D, Hamid O, Robert C, Hodi FS, Schachter J, Pavlick AC, Lewis KD, Cranmer LD, Blank CU, O'Day SJ, Ascierto PA, Salama AK, Margolin KA, Loquai C, Eigentler TK, Gangadhar TC, Carlino MS, Agarwala SS, Moschos SJ, Sosman JA, Goldinger SM, Shapira-Frommer R, Gonzalez R, Kirkwood JM, Wolchok JD, Eggermont A, Li XN, Zhou W, Zernhelt AM, Lis J, Ebbinghaus S, Kang SP and Daud A. Pembrolizumab versus investigator-choice chemotherapy for ipilimumab-refractory melanoma (KEYNOTE-002): a randomised, controlled, phase 2 trial. Lancet Oncol 2015; 16: 908-918.
- [4] Kandolf-Sekulovic L, Babovic N, Jokic N, Nikolin B, Nikolic D, Janjic Z, Mijugkovic Z, Rajovic M, Novakovic M, Pejcic I, Kovaevic P, Mihajlovic D, Roganovic T, Ferenc V, Nikolic J, Marinkovic M and Bizetic Z. Clinicopathological characteristics, diagnosis and treatment of

- melanoma in Serbia--the Melanoma Focus Study. Vojnosanit Pregl 2015; 72: 312-316.
- [5] Mignot G, Hervieu A, Vabres P, Dalac S, Jeudy G, Bel B, Apetoh L and Ghiringhelli F. Prospective study of the evolution of blood lymphoid immune parameters during dacarbazine chemotherapy in metastatic and locally advanced melanoma patients. PLoS One 2014; 9: e105907.
- [6] Beasley GM, Speicher P, Augustine CK, Dolber PC, Peterson BL, Sharma K, Mosca PJ, Royal R, Ross M, Zager JS and Tyler DS. A multicenter phase I dose escalation trial to evaluate safety and tolerability of intra-arterial temozolomide for patients with advanced extremity melanoma using normothermic isolated limb infusion. Ann Surg Oncol 2015; 22: 287-294.
- [7] Carvajal RD, Wong MK, Thompson JA, Gordon MS, Lewis KD, Pavlick AC, Wolchok JD, Rojas PB, Schwartz JD and Bedikian AY. A phase 2 randomised study of ramucirumab (IMC-1121B) with or without dacarbazine in patients with metastatic melanoma. Eur J Cancer 2014; 50: 2099-2107.
- [8] Klinac D, Gray ES, Freeman JB, Reid A, Bowyer S, Millward M and Ziman M. Monitoring changes in circulating tumour cells as a prognostic indicator of overall survival and treatment response in patients with metastatic melanoma. BMC Cancer 2014; 14: 423.
- [9] Hashimoto N, Tsuboi A, Kagawa N, Chiba Y, Izumoto S, Kinoshita M, Kijima N, Oka Y, Morimoto S, Nakajima H, Morita S, Sakamoto J, Nishida S, Hosen N, Oji Y, Arita N, Yoshimine T and Sugiyama H. Wilms tumor 1 peptide vaccination combined with temozolomide against newly diagnosed glioblastoma: safety and impact on immunological response. Cancer Immunol Immunother 2015; 64: 707-716.
- [10] Hardy KM, Strizzi L, Margaryan NV, Gupta K, Murphy GF, Scolyer RA and Hendrix MJ. Targeting nodal in conjunction with dacarbazine induces synergistic anticancer effects in metastatic melanoma. Mol Cancer Res 2015; 13: 670-680.
- [11] Grob JJ, Amonkar MM, Martin-Algarra S, Demidov LV, Goodman V, Grotzinger K, Haney P, Kampgen E, Karaszewska B, Mauch C, Miller WH Jr, Millward M, Mirakhur B, Rutkowski P, Chiarion-Sileni V, Swann S and Hauschild A. Patient perception of the benefit of a BRAF inhibitor in metastatic melanoma: quality-of-life analyses of the BREAK-3 study comparing dabrafenib with dacarbazine. Ann Oncol 2014; 25: 1428-1436.
- [12] Alrwas A, Papadopoulos NE, Cain S, Patel SP, Kim KB, Deburr TL, Bassett R Jr, Hwu WJ, Bedikian AY, Davies MA, Woodman SE and Hwu P. Phase I trial of biochemotherapy with

- cisplatin, temozolomide, and dose escalation of nab-paclitaxel combined with interleukin-2 and interferon-alpha in patients with metastatic melanoma. Melanoma Res 2014; 24: 342-348.
- [13] Bedikian AY, Garbe C, Conry R, Lebbe C, Grob JJ; Genasense Melanoma Study Group. Dacarbazine with or without oblimersen (a Bcl-2 antisense oligonucleotide) in chemotherapy-naive patients with advanced melanoma and lownormal serum lactate dehydrogenase: 'The AGENDA trial'. Melanoma Res 2014; 24: 237-243.
- [14] Leccia MT, Planchamp F, Sassolas B, Combemale P, Modiano P, Bedane C, Cupissol D, Derrey S, Dygai-Cochet I, Lamant L, Lubrano V, Mirabel X, Mourregot A, Rouge Bugat ME, Siegrist S, Thariat J, Tiffet O, Truc G, Verdoni L and Mazeau-Woynar V. [Management of patients with metastatic cutaneous melanoma: French national guidelines. French National Cancer Institute]. Ann Dermatol Venereol 2014; 141: 111-121.
- [15] Teimouri F, Nikfar S and Abdollahi M. Efficacy and side effects of dacarbazine in comparison with temozolomide in the treatment of malignant melanoma: a meta-analysis consisting of 1314 patients. Melanoma Res 2013; 23: 381-389.
- [16] Koprowska K, Hartman ML, Sztiller-Sikorska M and Czyz ME. Parthenolide enhances dacarbazine activity against melanoma cells. Anticancer Drugs 2013; 24: 835-845.
- [17] Ma C and Armstrong AW. Severe adverse events from the treatment of advanced melanoma: a systematic review of severe side effects associated with ipilimumab, vemurafenib, interferon alfa-2b, dacarbazine and interleukin-2. J Dermatolog Treat 2014; 25: 401-408.
- [18] Robert C, Dummer R, Gutzmer R, Lorigan P, Kim KB, Nyakas M, Arance A, Liszkay G, Schadendorf D, Cantarini M, Spencer S and Middleton MR. Selumetinib plus dacarbazine versus placebo plus dacarbazine as first-line treatment for BRAF-mutant metastatic melanoma: a phase 2 double-blind randomised study. Lancet Oncol 2013; 14: 733-740.
- [19] Ghosh SK, Bandyopadhyay D, Barma KD, Basu S and Roy A. Metastatic melanoma from an unknown primary site presenting as skin-colored nodules and multiple visceral involvement. Skinmed 2012; 10: 396-399.
- [20] Ugurel S, Paschen A and Becker JC. Dacarbazine in melanoma: from a chemotherapeutic drug to an immunomodulating agent. J Invest Dermatol 2013; 133: 289-292.

- [21] Ribas A, Kefford R, Marshall MA, Punt CJ, Haanen JB, Marmol M, Garbe C, Gogas H, Schachter J, Linette G, Lorigan P, Kendra KL, Maio M, Trefzer U, Smylie M, McArthur GA, Dreno B, Nathan PD, Mackiewicz J, Kirkwood JM, Gomez-Navarro J, Huang B, Pavlov D and Hauschild A. Phase III randomized clinical trial comparing tremelimumab with standard-of-care chemotherapy in patients with advanced melanoma. J Clin Oncol 2013; 31: 616-622.
- [22] Ferrucci PF, Minchella I, Mosconi M, Gandini S, Verrecchia F, Cocorocchio E, Passoni C, Pari C, Testori A, Coco P and Munzone E. Dacarbazine in combination with bevacizumab for the treatment of unresectable/metastatic melanoma: a phase II study. Melanoma Res 2015; 25: 239-245.
- [23] Faiao-Flores F, Quincoces Suarez JA, Fruet AC, Maria-Engler SS, Pardi PC and Maria DA. Curcumin analog DM-1 in monotherapy or combinatory treatment with dacarbazine as a strategy to inhibit in vivo melanoma progression. PLoS One 2015; 10: e0118702.
- [24] Maio M, Grob JJ, Aamdal S, Bondarenko I, Robert C, Thomas L, Garbe C, Chiarion-Sileni V, Testori A, Chen TT, Tschaika M and Wolchok JD. Five-year survival rates for treatment-naive patients with advanced melanoma who received ipilimumab plus dacarbazine in a phase III trial. J Clin Oncol 2015; 33: 1191-1196.
- [25] Jiang G, Li RH, Sun C, Liu YQ and Zheng JN. Dacarbazine combined targeted therapy versus dacarbazine alone in patients with malignant melanoma: a meta-analysis. PLoS One 2014; 9: e111920.
- [26] Saldanha G, Elshaw S, Sachs P, Alharbi H, Shah P, Jothi A and Pringle JH. microRNA-10b is a prognostic biomarker for melanoma. Mod Pathol 2016; 29: 112-121.
- [27] Tian Y, Wei W, Li L and Yang R. Down-Regulation of miR-148a Promotes Metastasis by DNA Methylation and is Associated with Prognosis of Skin Cancer by Targeting TGIF2. Med Sci Monit 2015; 21: 3798-3805.
- [28] Skourti E, Logotheti S, Kontos CK, Pavlopoulou A, Dimoragka PT, Trougakos IP, Gorgoulis V, Scorilas A, Michalopoulos I and Zoumpourlis V. Progression of mouse skin carcinogenesis is associated with the orchestrated deregulation of mir-200 family members, mir-205 and their common targets. Mol Carcinog 2016; 55: 1229-1242.