Original Article Association of TLR3 gene polymorphisms with age-related macular degeneration

Qin Wang

Department of Ophthalmology, Qianxinanzhou People's Hospital, No. 95, Yanan Road, Xingyi 562400, Guizhou, China

Received July 27, 2015; Accepted September 28, 2015; Epub February 15, 2016; Published February 29, 2016

Abstract: Purpose: In this study, we selected rs3775291 and rs3775296 polymorphisms of from toll-like receptor 3 (TLR3) gene to investigate their association with the susceptibility of age-related macular degeneration (ARMD or AMD). Haplotypes of the two polymorphisms were also detected in AMD patients. Methods: A hospital-based multiple-center case-control study was designed. 110 AMD cases and 108 healthy controls were enrolled in this study. χ^2 test was applied to count and analyze the genotype and allele frequencies in case and control groups. The relationship between TLR3 polymorphisms and the susceptibility of AMD was presented by odds ratios (ORs) and 95% confidence intervals (Cls). Results: The genotype distributions of TLR3 rs3775291 and rs3775296 polymorphisms in control group were in accordance with Hardy-Weinberg equilibrium (HWE) (P>0.05). The distributions of rs3775291 polymorphism AG genotype in case and control groups had statistically significant differences (P<0.05), and AG genotype could increase the onset risk of AMD by 1.89 times when compared with GG genotype (OR=1.89, 95% CI=1.09-3.28). In addition, rs3775296 polymorphism TT genotype was related to the occurrence of AMD (OR=4.70, 95% CI=1.26-17.46), and T allele could also increase the risk of AMD (OR=1.64, 95% CI=1.06-2.55). Linkage disequilibrium (LD) and haplotype analysis suggested that rs3775291 and rs3775296 polymorphisms formed 4 haplotypes, and the distribution differences of one of them, namely T-A haplotype, between case and control groups were statistically significant (P<0.05). Conclusions: TLR3 polymorphisms can increase the onset risk of AMD.

Keywords: Age-related macular degeneration (AMD), toll-like receptor 3 (TLR3) gene, haplotypes

Introduction

Age-related macular degeneration (ARMD or AMD) is a serious irreversible blinding eye disease. It mainly implicates the multi-layer tissues of retinal pigment epithelium (RPE), photoreceptor cell layer and choroid [1]. Epidemiological investigation results showed that AMD usually occurs in people over the age of 50, and the lesions invade both eves of some patients [2]. AMD remains the leading cause of blindness among the aged populations in America and other developed countries [3, 4]. There are about 50 million AMD patients all over the world and the morbidity is rapidly increasing along with the aging of the population. The older the age, the higher the incidence. About 6%~8% of the people over 75 years old in America have advanced macular degeneration that can lead to severe visual impairment [5, 6]. In China, the morbidity of AMD is approximately 5% in people over the age of 50, 8.8% in people aged 50~64 years, and 12% in people over 75 years old. AMD is gradually becoming one of the main eye diseases that can result in blindness [7].

Although most scientists believe that family history, heredity, smoking, dysimmunity, chronic photopathy, hyperopia, iris color, alimentary deficiency, obesity, chronic diseases and some other factors may have something to do with AMD, the specific pathogenesis of the disease is still cannot be determined at present [8]. There are two kinds of AMD: dry AMD and wet AMD, the latter of which does a greater harm to the eyesight. Various family studies indicated that AMD had a heritability as high as 71%, and it demonstrated that genetic factors played important roles in the occurrence of the disease [9, 10]. With the development of molecular biological technology, related researches on

Table 1. Primer sequences of rs3775291 and rs3775296
polymorphisms

Locus	Forward/Reverse	Primer sequence		
rs3775291 A/G	5291 A/G Forward 5'-TATTCCAGGCATAAAAAG-3			
	Reverse	5'-TATTGCTTTTTATGCCTG-3'		
rs3775296 C/T	Forward	5'-CATTTGAAAGCCATCTGC-3'		
	Reverse	5'-ATAGCAGATGGCTTTCAA-3'		

toll-like receptor 3 (*TLR3*) gene become a hot spot. *TLR3* gene widely express in the mammalian body, and has a close relationship with variety of signal transductions in the body [11, 12]. In the last few years, studies on the relationship between Toll-like receptors (*TLRs*) and AMD have been gradually increasing. The latest study conducted among Caucasian populations in Europe and America pointed out that *TLR3* rs3775291 single nucleotide polymorphism (SNP) was associated with the occurrence and features of AMD [13, 14].

This study genotyped *TLR3* rs3775291 and rs3775296 SNPs using polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) method. Relationship between the two SNPs and AMD, association between the haplotypes of the two SNPs and AMD were analyzed in this study too. We hoped that it would provide a scientific basis for the early prevention of AMD.

Materials and methods

Clinical information

According to the diagnostic criteria of AMD [15], we collected the clinical information of 110 diagnosed wet AMD patients (48 males and 62 females) who were sporadically hospitalized in the ophthalmology department of Qianxinanzhou People's Hospital. The cases were proved to have choroidal neovascularization (CNV) by fluorescence fundus angiography (FFA) and they were excluded the possibilities of having neovascularization diseases, high myopia and diabetic retinopathy, with a mean age of 65 years. 108 healthy persons in the same hospital volunteered to take part in the study. The healthy controls were 45 males and 63 females with a mean age of 60. Their eye examination results were normal, and they did not suffer from systemic blood diseases like hypertension and diabetes, and had no family history of AMD. The cases and controls were all Han population of China living in the local area for a certain time, and they were not connected with each other by blood. The subjects were fully informed the study contents and they agreed to sign informed consent.

Sample collection process

4 ml fasting elbow venous blood of each participant was collected after 12 hours fasting. We injected the blood samples into anti-coagulative tubes and preserved the samples in refrigerator at -20°C until to use.

PCR-RFLP process and genotype determination

We applied polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) method to detect the genotypes of TLR3 gene rs3775291 and rs3775296 polymorphisms. Primer sequences used for amplification are listed in **Table 1**. The PCR process was occurred in a total of 30 µL systems, which consisted of 10× Buffer 3 µL, dNTPs 2.4 µL (final concentration 0.2 mM), forward and reverse primers 0.9 µL each (final concentration 0.3 µM), Tag DNA polymerase 0.24 µL (final concentration 0.04 mM), DNA template 2 µL and ddH_aO 20.56 µL. PCR reaction conditions were: at first, 94°C initial denaturation for 5 min; then 35 cycles of 94°C for 30 s, 55°C for 30 s, 72°C for 30 s; finally 72°C extension for 10 min.

Restriction enzyme Taq I (TthHB8I) was used to digest the above PCR products, with a recognition sequence as TJCGA. The total enzymatic system was 20 μ L, including 10× Taq I Basal Buffer 2 μ L, 10 u/ μ L Taq I 1 μ L, 0.1% BSA 2 μ L, PCR products 10 μ L and dd H₂O 5 μ L. Such enzymatic system was mixed well, centrifuged instantaneously, and then incubated in a water bath at 65°C for 30 min. Digested products were put in 2% agarose gel (containing 0.5 g/ mL ethidium bromide) for electrophoresis. The results were determined using gel imaging system.

Statistical analysis

The data were processed by SPSS18.0 statistical software. Hardy-Weinberg equilibrium (HWE) was used to detect the representativeness of the subjects. Linkage disequilibrium

130110200 poi	ymorphism	5			
Genotype/Allele	Case (n=110)	Control (n=108)	X ²	Ρ	OR (95% CI)
rs3775291					
GG	42/38.2	58/53.7	-	-	1.00
AG	63/57.3	46/42.6	5.21	0.03	1.89 (1.09-3.28)
AA	5/4.5	4/3.7	0.62	0.50	1.73 (0.44-6.82)
G	147/66.8	162/75	-	-	1.00
А	73/33.2	54/25	3.53	0.07	1.49 (0.98-2.26)
rs3775296					
GG	57/51.8	67/62	-	-	1.00
GT	41/37.3	38/35.2	0.68	0.47	1.27 (0.72-2.23)
TT	12/10.9	3/2.8	6.20	0.02	4.70 (1.26-17.49)
G	155/70.5	172/79.6	-	-	1.00
Т	65/29.5	44/20.4	4.89	0.03	1.64 (1.06-2.55)

Table 2. Genotype and allele distributions of *TLR3* rs3775291 andrs3775296 polymorphisms

Table 3. Linkage disequilibrium and haplotype analysis on

 rs3775291 and rs3775296 polymorphisms

Haplotype Locus1-Locus2	Case 2 (n=220)	Control 2 (n=216)	X ²	Ρ	OR (95% CI)	
G-A	56/25.5	50/23.1	-	-	1.00	
G-T	99/45	122/56.5	1.86	0.19	0.73 (0.46-1.15)	
T-A	17/7.7	4/1.9	5.67	0.03	3.80 (1.20-12.03)	
T-G	48/21.8	40/18.5	0.06	0.89	1.07 (0.61-1.89)	

Notes: locus1, rs3775291; locus2, rs3775296.

and the haplotypes between *TLR3* gene polymorphisms were assessed by Haploview. Each difference between case and control groups was assessed by χ^2 test. Odds ratios (ORs) with 95% confidence intervals (Cls) were used to calculate the relationship between *TLR3* polymorphisms and AMD susceptibility. When P<0.05, differences between groups were considered with statistical significance.

Results

HWE detection

We inspected the HWE examination of the genotypes of *TLR3* rs3775291 and rs3775296 polymorphisms in case and control groups. Then we found that the genotype distributions in control group met the genetic equilibrium (P>0.05), which suggested that the controls had representativeness.

Genotype and allele frequency distributions of TLR3 polymorphisms

The distributions and comparisons results of genotypes and alleles in *TLR3* polymorphisms

groups were given in Table 2. As we could see, for rs3775291 polymorphism, AG genotype was more frequently in case group than that in control group, and the distribution difference was statistically significant (P< 0.05). Besides, AG genotype increased the onset risk of AMD when compared with GG genotype (OR=1.89, 95% CI=1.09-3.28). In rs3775296 polymorphism, TT genotype had a higher frequency in case group than that in control group and the distribution difference between groups was obviously (P<0.05). TT genotype could also increase the incidence of AMD (OR= 4.70, 95% CI=1.26-17.49). In the mean time, T allele of rs3775296 significantly associated with the AMD susceptibility (P=0.03, OR=1.64, 95% CI=1.06-2.55).

between case and control

Linkage disequilibrium and haplotype analysis on TLR3

polymorphisms

Analysis of linkage disequilibrium was performed by Haploview online software, then we discovered that there existed a high linkage disequilibrium between rs3775291 and rs-3775296 polymorphisms of TLR3. The two polymorphisms could form four kinds of haplotypes: G-A, G-G, T-A and T-G. Haplotype analysis results were shown in Table 3. It could be observed that the distributions of T-A halplotype in case and control groups were significantly different (P<0.05), and T-A could increase the risk of AMD when compared with G-A haplotype (x²=5.67, P=0.03, OR=3.80, 95% CI=1.20-12.03). Additionally, the distributions of G-A, G-G and T-G haplotypes in case and control groups were not statistically significant (P>0.05). Thus we concluded that T-A was a susceptible haplotype to AMD.

Discussion

In recent years, the incidence of AMD in middle and old aged people has been increasing year by year all over the world. AMD is the most common reason for irreversible loss of vision and it has been the main cause of blindness in people over 50 years old in the developed countries [16, 17]. In America, over 8 million people have suffered from the disease [3]. China has developed into an aging stage, and the vision problems of the elderly, especially age-related eye diseases have gradually become a social concern. Researches carried out in different regions showed that the incidence of AMD in China was about 3%-15.5%, which seriously affected the life quality of the elderly and increased the economic burden of the society [18].

Related studies about TLR3 polymorphisms and eye diseases at present all have confirmed the existence of a close relationship between them, and TLR3 can promote the occurrence and development of AMD [19-21]. TLRs play important roles in the human body. It can identify a variety of pathogenic modes. TLR3 gene locates in human chromosome 4q35 region, which included 5 exons and 4 introns. The rs3775291 polymorphism of TLR3 results in the 412th encoding amino acid mutating from leucine into phenylalanine. In the year 2009, Cho et al. undertook a study about the correlation between TLR3 rs3775291 polymorphism and TLR4 rs4986790 polymorphism and AMD, but they failed to find an obvious positive result [22]. The next year, Sng et al. compared the frequency distributions of genotypes and alleles of TLR3 rs3775291 polymorphism in 246 Singaporean Chinese people who had exudative AMD. They found that the correlation between TLR3 rs3775291 polymorphism and the onset of AMD among Singaporean Chinese people was not significant [23]. However, Maloney et al. indicated that TLR3 protein might be a receptor of the human wet AMD lesions [24]. Zhu et al. discovered that the mRNA and protein expressions of TLR2 and TLR3 from the peripheral blood mononuclear cells (PBMCs) were higher in case group than in control group, and they inferred that TLR3 might be related to the pathogenesis of wet AMD [25].

The present study ascertained that *TLR3* polymorphisms (rs3775291 and rs3775296) were associated with AMD. As the detecting results indicated, rs3775291 polymorphism AG genotype was highly distributed in the case group

and it was a susceptible factor to AMD; rs3775296 polymorphism TT genotype and T allele respectively increased 4.7 times and 1.64 times of the onset risk of AMD. The result suggested a significant association of rs3775296 polymorphism with the incidence of AMD in Chinese Han population. Furthermore, linkage disequilibrium and haplotype analysis found that the frequencies of T-A haplotype of the two SNPs of *TLR3* gene in case and control groups were significantly different. T-A haplotype had 3.80 times increased risk of developing AMD, which manifested that T-A was a susceptible haplotype for AMD.

Because of the racial differences, the research results among different races are not consistent with each other. Genetic influences of *TLR3* gene polymorphisms on AMD are quite different between Chinese people and white people, while the pathogenesis has not been fully understood. Even so, we have suggested that *TLR3* polymorphisms might associate with the susceptibility of wet AMD in China. With the development of research and application of molecular biology, we hope that the further studies will provide certain directive significance for the exploration of the pathogenesis and treatment of AMD.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Qin Wang, Department of Ophthalmology, Qianxinanzhou People's Hospital, No. 95, Yanan Road, Xingyi 562400, Guizhou, China. Tel: +86-13885908769; E-mail: wanggwing@sina.com

References

- [1] de Jong PT. Age-related macular degeneration. N Engl J Med 2006; 355: 1474-1485.
- [2] Spaide RF, Armstrong D and Browne R. Continuing medical education review: choroidal neovascularization in age-related macular degeneration-what is the cause? Retina 2003; 23: 595-614.
- [3] Friedman DS, O'Colmain BJ, Munoz B, Tomany SC, McCarty C, de Jong PT, Nemesure B, Mitchell P and Kempen J. Prevalence of age-related macular degeneration in the United States. Arch Ophthalmol 2004; 122: 564-572.
- [4] Evans J and Wormald R. Is the incidence of registrable age-related macular degeneration increasing? Br J Ophthalmol 1996; 80: 9-14.

- [5] Klein R, Peto T, Bird A and Vannewkirk MR. The epidemiology of age-related macular degeneration. Am J Ophthalmol 2004; 137: 486-495.
- [6] Klein R, Klein BE and Linton KL. Prevalence of age-related maculopathy. The Beaver Dam Eye Study. Ophthalmology 1992; 99: 933-943.
- [7] Li Y, Xu L, Jonas JB, Yang H, Ma Y and Li J. Prevalence of age-related maculopathy in the adult population in China: the Beijing eye study. Am J Ophthalmol 2006; 142: 788-793.
- [8] Ambati J, Ambati BK, Yoo SH, Ianchulev S and Adamis AP. Age-related macular degeneration: etiology, pathogenesis, and therapeutic strategies. Surv Ophthalmol 2003; 48: 257-293.
- [9] Klaver CC, Wolfs RC, Assink JJ, van Duijn CM, Hofman A and de Jong PT. Genetic risk of agerelated maculopathy. Population-based familial aggregation study. Arch Ophthalmol 1998; 116: 1646-1651.
- [10] Yates JR and Moore AT. Genetic susceptibility to age related macular degeneration. J Med Genet 2000; 37: 83-87.
- [11] Takeda K, Kaisho T and Akira S. Toll-like receptors. Annu Rev Immunol 2003; 21: 335-376.
- [12] Lafon M, Megret F, Lafage M and Prehaud C. The innate immune facet of brain: human neurons express TLR-3 and sense viral dsRNA. J Mol Neurosci 2006; 29: 185-194.
- [13] Yang Z, Stratton C, Francis PJ, Kleinman ME, Tan PL, Gibbs D, Tong Z, Chen H, Constantine R, Yang X, Chen Y, Zeng J, Davey L, Ma X, Hau VS, Wang C, Harmon J, Buehler J, Pearson E, Patel S, Kaminoh Y, Watkins S, Luo L, Zabriskie NA, Bernstein PS, Cho W, Schwager A, Hinton DR, Klein ML, Hamon SC, Simmons E, Yu B, Campochiaro B, Sunness JS, Campochiaro P, Jorde L, Parmigiani G, Zack DJ, Katsanis N, Ambati J and Zhang K. Toll-like receptor 3 and geographic atrophy in age-related macular degeneration. N Engl J Med 2008; 359: 1456-1463.
- [14] Edwards AO, Chen D, Fridley BL, James KM, Wu Y, Abecasis G, Swaroop A, Othman M, Branham K, Iyengar SK, Sivakumaran TA, Klein R, Klein BE and Tosakulwong N. Toll-like receptor polymorphisms and age-related macular degeneration. Invest Ophthalmol Vis Sci 2008; 49: 1652-1659.
- [15] Age-Related Eye Disease Study Research Group. The Age-Related Eye Disease Study (AREDS): design implications. AREDS report no. 1. Control Clin Trials 1999; 20: 573-600.
- [16] Pascolini D, Mariotti SP, Pokharel GP, Pararajasegaram R, Etya'ale D, Negrel AD and Resnikoff S. 2002 global update of available data on visual impairment: a compilation of population-based prevalence studies. Ophthalmic Epidemiol 2004; 11: 67-115.

- [17] Congdon N, O'Colmain B, Klaver CC, Klein R, Munoz B, Friedman DS, Kempen J, Taylor HR and Mitchell P. Causes and prevalence of visual impairment among adults in the United States. Arch Ophthalmol 2004; 122: 477-485.
- [18] Sundelin S, Wihlmark U, Nilsson SE and Brunk UT. Lipofuscin accumulation in cultured retinal pigment epithelial cells reduces their phagocytic capacity. Curr Eye Res 1998; 17: 851-857.
- [19] Kleinman ME, Kaneko H, Cho WG, Dridi S, Fowler BJ, Blandford AD, Albuquerque RJ, Hirano Y, Terasaki H, Kondo M, Fujita T, Ambati BK, Tarallo V, Gelfand BD, Bogdanovich S, Baffi JZ and Ambati J. Short-interfering RNAs induce retinal degeneration via TLR3 and IRF3. Mol Ther 2012; 20: 101-108.
- [20] Zhou P, Fan L, Yu KD, Zhao MW and Li XX. Tolllike receptor 3 C1234T may protect against geographic atrophy through decreased dsRNA binding capacity. FASEB J 2011; 25: 3489-3495.
- [21] Wornle M, Merkle M, Wolf A, Ribeiro A, Himmelein S, Kernt M, Kampik A and Eibl-Lindner KH. Inhibition of TLR3-mediated proinflammatory effects by Alkylphosphocholines in human retinal pigment epithelial cells. Invest Ophthalmol Vis Sci 2011; 52: 6536-6544.
- [22] Cho Y, Wang JJ, Chew EY, Ferris FL 3rd, Mitchell P, Chan CC and Tuo J. Toll-like receptor polymorphisms and age-related macular degeneration: replication in three case-control samples. Invest Ophthalmol Vis Sci 2009; 50: 5614-5618.
- [23] Sng CC, Cackett PD, Yeo IY, Thalamuthu A, Venkatraman A, Venkataraman D, Koh AH, Tai ES, Wong TY, Aung T and Vithana EN. Toll-like receptor 3 polymorphism rs3775291 is not associated with choroidal neovascularization or polypoidal choroidal vasculopathy in Chinese subjects. Ophthalmic Res 2011; 45: 191-196.
- [24] Maloney SC, Antecka E, Orellana ME, Fernandes BF, Odashiro AN, Eghtedari M and Burnier MN Jr. Choroidal neovascular membranes express toll-like receptor 3. Ophthalmic Res 2010; 44: 237-241.
- [25] Zhu Y, Liang L, Qian D, Yu H, Yang P, Lei B and Peng H. Increase in peripheral blood mononuclear cell Toll-like receptor 2/3 expression and reactivity to their ligands in a cohort of patients with wet age-related macular degeneration. Mol Vis 2013; 19: 1826-1833.