# Original Article Polymorphic variants of FGFR-4 associated with clinical characteristics in urothelial cell carcinoma

Wei-Chun Weng<sup>1,2</sup>, Yu-Hui Huang<sup>3,4</sup>, Ying-Erh Chou<sup>3,5</sup>, Shian-Shiang Wang<sup>1,6</sup>, Ming-Ju Hsieh<sup>1,7</sup>, Shun-Fa Yang<sup>1,5</sup>

<sup>1</sup>Institute of Medicine, Chung Shan Medical University, Taichung, Taiwan; <sup>2</sup>Division of Urology, Department of Surgery, Tungs' Taichung MetroHarbor Hospital, Taichung, Taiwan; <sup>3</sup>School of Medicine, Chung Shan Medical University, Taichung, Taiwan; <sup>4</sup>Department of Physical Medicine & Rehabilitation, Chung Shan Medical University Hospital, Taichung, Taiwan; <sup>5</sup>Department of Medical Research, Chung Shan Medical University Hospital, Taichung, Taiwan; <sup>6</sup>Division of Urology, Department of Surgery, Taichung Veterans General Hospital, Taichung, Taiwan; <sup>7</sup>Cancer Research Center, Changhua Christian Hospital, Changhua, Taiwan

Received December 9, 2015; Accepted March 19, 2016; Epub June 15, 2016; Published June 30, 2016

**Abstract:** Background: Fibroblast growth factor receptor-4 (FGFR-4) polymorphic variants have an important role on the development of various cancers. However, the association between FGFR-4 polymorphic variants and the risk of urothelial cell carcinoma (UCC) has not been determined. In this study, we studied the associations of FGFR-4 polymorphic variants with UCC susceptibility and its clinicopathological features. Materials and methods: Four SNPs in FGFR-4 were analyzed among 558 participants comprising 279 controls and 279 patients with UCC by performing a real-time genotype PCR assay. Results: Our results showed that UCC patients who carried at least one A genotype at rs1966265 exhibited a higher risk of invasive tumor stage compared with those carrying the wild-type genotype (P < 0.05). Patients with UCC who carried at least one G genotype (AG and GG) at rs2011077 also exhibited a higher risk of invasive tumor stage in Taiwanese patients in FGFR-4 rs1966265 and rs2011077 may be associated with the risk of invasive tumor stage in Taiwanese patients with UCC. This is the first study to determine the association between FGFR-4 polymorphic variants and UCC tumor stage progression.

Keywords: Polymorphic variants, fibroblast growth factor receptor 4, urothelial cell carcinoma

#### Introduction

Urothelial cell carcinoma (UCC) is the most common malignancy of the urinary tract. In Taiwan, UCC is the ninth leading malignancy among men and the 16th leading malignancy among women. The most widely known risk factors for UCC are tobacco consumption and exposure to aromatic amines [1-3]. Increasingly more evidence regarding the impact of genetic factors in the risk of UCC has been published [4-6].

Fibroblast growth factors (FGFs) are effective regulators of cell proliferation, and cell differentiation. The function are critical in normal development, tissue maintenance angiogenesis, embryonic development, and wound repair [7]. At least 20 FGF members exist, designated FGF-1 through FGF-20, but acidic FGF and basic FGF are names commonly used for FGF-1 and FGF-2, and keratinocyte growth factor (KGF) is the name generally used for FGF-7. FGFs represent a large family of secreted molecules [8]. The FGF receptor (FGFR) family comprises 4 family members, is a subsequent formation of various complexes to initiate downstream signal transduction [9-11]. Aberrant FGFR signaling is mainly contributed to several mechanisms involving genetic variants, gene amplification and chromosomal translocation [12-14].

Fibroblast growth factor receptor-4 (FGFR-4) overexpression is commonly detected in numerous types of human cancers, for example breast cancers, ovarian cancer, bladder cancer, lung cancer, and gastric cancers [15]. Moreover, previous studies have reported that for FGFR-4, the G to A polymorphic variants in rs1966265 (Ile10Val), and the C to T polymorphic variants

Vesiele	
variable	Patients ( $N = 279$ )
Age (yrs)	Mean ± S.D.
	68.75 ± 12.06
Gender	n (%)
Male	182 (65.2%)
Female	97 (34.8%)
Tobacco consumption	
No	203 (72.8%)
Yes	76 (27.2%)
Stage	
Superficial tumor (pTa-pT1)	170 (60.9%)
Invasive tumor (pT2-pT4)	109 (39.1%)
Tumor T status	
ТО	76 (27.2%)
T1-T4	203 (72.8%)
Lymph node status	
NO	251 (90.0%)
N1+N2	28 (10.0%)
Metastasis	
MO	275 (98.6%)
M1	4 (1.4%)
Histopathologic grading	
Low grade	40 (14.3%)
High grade	239 (85.7%)

Table 1. The distributions of demographica	al
characteristics in 279 patients with LICC	

in rs351855 (Gly388Arg) may alters protein expression [16-18]. Additionally, these polymorphic variants have a pathophysiological impact on tumor development in prostate or breast cancers [19, 20]. However, no report has been published on the relationship between FGFR-4 polymorphic variants and UCC risks. Thus, this study explored the associations among these 4 polymorphic variants of FGFR-4 and the risk of UCC, and evaluated the impact of these polymorphic variants on the clinicopathological features of UCC.

#### Materials and methods

#### Subjects and specimen collection

279 patients (182 men and 97 women, with a mean age of 68.75 years) were recruited in 2010-213 at Taichung Veterans General Hospital (TVGH) in Taichung, Taiwan. All 279 patients have pathology proved as urothelial cell carcinoma of bladder or upper urinary tract. The control group was enrolled from the physical examination during the hospital. The approval was obtained from the Institutional Review Board of TVGH, and informed written consent to patients was obtained from each person before commencement of this study.

## FGFR-4 polymorphic variants genotyping

Genomic DNA was extracted using AxyPrep Blood Genomic DNA Miniprep Kit (Axygen Biosciences, Union City, CA, USA). The FGFR-4 rs351855, rs1966265, rs7708357 and rs20-11077 polymorphic variants were assessed by using the TaqMan assay with an ABI Step-OnePlus<sup>™</sup> Real-Time PCR System as previously described [21].

## Statistical analysis

The AOR and 95% Cls of the association between genotype frequencies and risk or clinicopathological features were estimated using multiple logistic regression models, after controlling for age, gender and tobacco. Differences between the 2 groups were significant if p values were < 0.05. We analyzed all data with SPSS software (version 17.0, SPSS Inc., Chicago, IL, USA).

## Results

The distributions of demographics of 279 patients with UCC are shown in Table 1. Compared with the control group, no significant differences were found in age (68.8  $\pm$  12.1 vs  $67.9 \pm 10.8$  y, P = 0.375), sex, and tobacco consumption. The frequencies of FGFR-4 genes were all in the Hardy-Weinberg equilibrium among the control group. The distribution of FGFR-4 genotypes is shown in Table 2. The most frequent alleles were heterozygous G/A, C/T, and A/G for rs1966265, rs351855, and rs2011077, respectively, and homozygous G/G for rs7708357. After adjusting for age, sex, and tobacco consumption, no difference in genotype frequencies for rs1966265, rs351855, rs2011077, and rs7708357 of the FGFR-4 gene was observed between the case and control groups (Table 2).

The distribution of the pathological status and FGFR-4 genotypes in patients with UCC were estimated to clarify the role of FGFR-4 polymorphic variants in the pathological clinical status of patients with UCC. Pathological status

Variable	Controls (N = 279) n (%)	Patients (N = 279) n (%)	OR (95% CI)	AOR (95% CI)	
rs1966265					
GG	73 (26.2%)	75 (26.9%)	1.00	1.00	
GA	143 (51.2%)	140 (50.2%)	0.953 (0.640-1.418)	0.893 (0.568-1.404)	
AA	63 (22.6%)	64 (22.9%)	0.989 (0.615-1.589)	0.766 (0.443-1.324)	
GA+AA	206 (73.8%)	204 (73.1%)	0.964 (0.662-1.404)	0.853 (0.555-1.309)	
rs351855					
CC (Gly/Gly)	76 (27.2%)	84 (30.1%)	1.00	1.00	
CT (Gly/Arg)	149 (53.4%)	136 (48.7%)	0.826 (0.560-1.217)	0.789 (0.508-1.226)	
TT (Arg/Arg)	54 (19.4%)	59 (21.2%)	0.989 (0.610-1.601)	0.773 (0.455-1.342)	
CT+TT	203 (72.8%)	195 (69.9%)	0.869 (0.602-1.255)	0.784 (0.516-1.192)	
rs2011077					
AA	72 (25.8%)	76 (27.2%)	1.00	1.00	
AG	143 (51.3%)	139 (49.9%)	0.921 (0.619-1.371)	0.868 (0.552-1.364)	
GG	64 (22.9%)	64 (22.9%)	0.947 (0.590-1.521)	0.727 (0.421-1.256)	
AG+GG	207 (74.2%)	203 (72.8%)	0.929 (0.638-1.353)	0.823 (0.536-1.263)	
rs7708357					
GG	269 (96.4%)	272 (97.5%)	1.00	1.00	
GA	10 (3.6%)	7 (2.5%)	0.692 (0.260-1.845)	0.749 (0.246-2.282)	
AA	0 (0%)	0 (0%)			
GA+AA	10 (3.6%)	7 (2.5%)	0.692 (0.260-1.845)	0.749 (0.246-2.282)	

Table 2. Distribution frequency of FGFR-4 genotypes in 279 controls and 279 UCC patients

**Table 3.** Distribution frequency of the clinical status and FGFR-4 rs1966265 genotype frequencies in279 patients with UCC

Veriable	FGFR-4 (rs1966265)					
Variable	GG (%) ( <i>N</i> = 75)	GA+AA (%) ( <i>N</i> = 204)	p value	p value		
Stage						
Superficial tumor (pTa-pT1)	53 (70.7%)	117 (57.4%)	1.00			
Invasive tumor (pT2-pT4)	22 (29.3%)	87 (42.6%)	1.791 (1.014-3.165)	P=0.029*		
Tumor T status						
то	19 (25.3%)	57 (27.9%)	1.00			
T1-T4	56 (74.7%)	147 (72.1%)	0.875 (0.478-1.600)	P = 0.393		
Lymph node status						
NO	71 (94.7%)	180 (88.2%)	1.00			
N1+N2	4 (5.3%)	24 (11.8%)	2.367 (0.793-7.064)	P = 0.082		
Metastasis						
MO	74 (98.7%)	201 (98.5%)	1.00			
M1	1 (1.3%)	3 (1.5%)	1.104 (0.113-10.785)	P = 0.707		
Histopathologic grading						
Low grade	9 (12.0%)	31 (15.2%)	1.00			
High grade	66 (88.0%)	173 (84.8%)	0.761 (0.344-1.684)	P = 0.321		

\*p value < 0.05 as statistically significant.

assessments included the stage, tumor size and lymph node statuses, as well as metastasis and histopathologic grading. No significant differences were observed between other FGFR-4 genotypic frequencies and tumor and lymph node statuses or metastasis and histopathologic grading (**Tables 3** and **4**). However, for FGFR-4 rs1966265, patients with the het-

Verieble	FGFR-4 (rs2011077)					
variable	AA (%) ( <i>N</i> = 76)	AG+GG (%) ( <i>N</i> = 203)	p value	p value		
Stage						
Superficial tumor (pTa-pT1)	54 (71.1%)	116 (57.1%)	1.00			
Invasive tumor (pT2-pT4)	22 (28.9%)	87 (42.9%)	1.841 (1.043-3.250)	P=0.023*		
Tumor T status						
ТО	19 (25.0%)	57 (28.1%)	1.00			
T1-T4	57 (75.0%)	146 (71.9%)	0.854 (0.467-1.560)	P = 0.362		
Lymph node status						
NO	72 (94.7%)	179 (88.2%)	1.00			
N1+N2	4 (5.3%)	24 (11.8%)	2.413 (0.809-7.202)	P = 0.076		
Metastasis						
MO	75 (98.7%)	200 (98.5%)	1.00			
M1	1 (1.3%)	3 (1.5%)	1.125 (0.115-10.984)	P=0.701		
Histopathologic grading						
Low grade	9 (11.8%)	31 (15.3%)	1.00			
High grade	67 (88.2%)	172 (84.7%)	0.745 (0.337-1.649)	P = 0.302		

Table 4. Distribution frequency of the clinical status and FGFR-4 rs2011077 genotype frequencies ir
279 patients with UCC

\**p* value < 0.05 as statistically significant.

Table 5. The estimated haplotype frequencies of four examined polymorphic variants in FGFR-4 gene
and the corresponding risk for UCC

Variable		Controls Patients					
rs1966265	rs351855	rs2011077	rs7708357	(N = 558)	(N = 558)	OR (95% CI)	p value
G/A	T/A	A/G	G/T	n (%)	n (%)		
G	С	А	G	279 (50.0%)	286 (51.3%)	Reference	
А	Т	G	G	247 (44.2%)	248 (44.3%)	0.979 (0.769-1.247)	0.866
А	С	G	G	16 (2.9%)	12 (2.2%)	0.732 (0.340-1.575)	0.423
Others#				16 (2.9%)	12 (2.2%)	0.732 (0.340-1.575)	0.423

#Others: ATGA: 7 cases; GTAG: 6 cases; GCAA: 5 cases; ACGA: 5 cases; GTGG: 3 cases; ACAG: 2 cases.

erozygous G/A or A/A genotype exhibited a high risk of UCC with invasive tumor stage (P < 0.05) compared with those with the homozygous G/G genotype, as shown in Table 3. In addition, patients with the heterozygous A/G or G/G genotype also exhibited a high risk of UCC with invasive tumor stage (P < 0.05) compared with those with the homozygous A/A genotype for FGFR-4 rs2011077 (Table 4). No significant differences were observed between FGFR-4 rs351855 and rs7708357 genotypic and invasive tumor clinical status. The relationship of FGFR-4 haplotypes with the risk of developing UCC was also evaluated. The frequency distributions of three common FGFR-4 rs1966265. rs351855, rs2011077, and rs7708357 haplotypes are shown in Table 5, with the most frequent haplotype in the controls (GCAG) being chosen as the reference. We found that all these common FGFR-4 haplotypes exhibit no significant associations with increased susceptibility to UCC (**Table 5**).

#### Discussion

This is the first study to determine a significant association between FGFR-4 polymorphic variants and UCC risk. The results showed that patients carrying the G/A or A/A genotype of the FGFR-4 rs1966265 polymorphic variant exhibited a significantly increased risk of UCC invasive tumor stage. Compared with the A/A genotype of the FGFR-4 rs2011077, the A/G + G/G genotype also exhibited a higher risk of UCC invasive tumor stage.

FGFRs consist of 4 closely related genes (FGFR1-4) associated with the development

of several types of human cancers, for example breast cancers, ovarian cancer, bladder cancer, lung cancer, and gastric cancers [22, 23]. Bang et al reported that the FGFR-4 rs351855 polymorphic variant was associated with metastasis in prostate and breast cancer patients [24]. Moreover, Sugiyama et al., also reported the FGFR-4-R388 allele could increase tumor invasion by decreasing MT1-MMP degradation, thus exerts to poor cancer prognosis [25]. Aberrant expression and polymorphic variant of fibroblast growth factor receptor 4 (FGFR-4) has been linked to tumor progression and anticancer drug resistance. Knockdown of FGFR-4-R388 or MT1-MMP through RNA interference blocked tumor cell invasion and growth in collagen. This was coupled with impaired phosphorylation of FGFR substrate 2 and Src, upregulation of E-cadherin, and suppression of cadherin-11 and N-cadherin [26]. However, few studies have reported the expression of FGFR-4 and its associated polymorphic variants with UCC. However, no significant differences were observed between FGFR-4 rs351855 genotypic and invasive tumor clinical status in our study.

We observed that polymorphic variants in FGFR-4 rs1966265 and FGFR-4 rs2011077 were associated with a high risk of invasive tumor stage in patients with UCC. FGFR-4 polymorphic variant rs1966265 exhibited an association with both respiratory distress and bronchopulmonary dysplasia [16]. FGFR-4 G alleles of the rs2011077 polymorphic variant have a significant impact on the development of prostate cancer and benign prostate hyperplasia [17]. Although the rs2011077 polymorphic variant is located in the intron, it may affect mRNA splicing or transcription activity. It is suggested that the rs2011077 SNP may increase the activity or expression of FGFR-4 at the UCC progressive stage. These findings may explain the relationship between the G alleles of the rs2011077 of FGFR-4 and the increased invasive tumor status of UCC observed in the present study. However, the mechanism by which this SNP modulates the tumor development of UCC should be further investigated in future studies.

In conclusion, our results show that an association between FGFR-4 rs1966265 and rs2011077 was detected with a high risk of invasive tumor in patients with UCC. These find-

ings indicate a novel genetic predisposition to urothelial cell tumorigenesis.

## Acknowledgements

This study was supported by a research grant from Chung Shan Medical University and Tungs' Taichung Metro Harbor Hospital (CSMU-TTM-104-01).

## Disclosure of conflict of interest

None.

Address correspondence to: Drs. Shun-Fa Yang and Ming-Ju Hsieh, Institute of Medicine, Chung Shan Medical University, Taichung 402, Taiwan. Tel: +886-4-24739595 Ext. 34253; Fax: +886-4-24723229; E-mail: ysf@csmu.edu.tw (SFY); 170780@cch.org.tw (MJH)

#### References

- [1] Colin P, Koenig P, Ouzzane A, Berthon N, Villers A, Biserte J, Roupret M. Environmental factors involved in carcinogenesis of urothelial cell carcinomas of the upper urinary tract. BJU Int 2009; 104: 1436-1440.
- [2] Stern MC, Lin J, Figueroa JD, Kelsey KT, Kiltie AE, Yuan JM, Matullo G, Fletcher T, Benhamou S, Taylor JA, Placidi D, Zhang ZF, Steineck G, Rothman N, Kogevinas M, Silverman D, Malats N, Chanock S, Wu X, Karagas MR, Andrew AS, Nelson HH, Bishop DT, Sak SC, Choudhury A, Barrett JH, Elliot F, Corral R, Joshi AD, Gago-Dominguez M, Cortessis VK, Xiang YB, Gao YT, Vineis P. Sacerdote C. Guarrera S. Polidoro S. Allione A, Gurzau E, Koppova K, Kumar R, Rudnai P, Porru S, Carta A, Campagna M, Arici C, Park SS, Garcia-Closas M; International Consortium of Bladder Cancer. Polymorphisms in DNA repair genes, smoking, and bladder cancer risk: Findings from the international consortium of bladder cancer. Cancer Res 2009; 69: 6857-6864.
- [3] Rouissi K, Stambouli N, Marrakchi R, Slama MR, Cherif M, Sfaxi M, Chebil M, Elgaaied AB, Ouerhani S. Smoking and polymorphisms in folate metabolizing genes and their effects on the histological stage and grade for bladder tumors. Bull Cancer 2011; 98: E1-10.
- [4] Wang L, Wang G, Lu C, Feng B, Kang J. Contribution of the -160c/a polymorphism in the e-cadherin promoter to cancer risk: A meta-analysis of 47 case-control studies. PLoS One 2012; 7: e40219.
- [5] Wang YH, Chiou HY, Lin CT, Hsieh HY, Wu CC, Hsu CD, Shen CH. Association between sur-

vivin gene promoter -31 c/g polymorphism and urothelial carcinoma risk in taiwanese population. Urology 2009; 73: 670-674.

- [6] Wang SS, Liu YF, Ou YC, Chen CS, Li JR, Yang SF. Impacts of ca9 gene polymorphisms on urothelial cell carcinoma susceptibility and clinicopathologic characteristics in taiwan. PLoS One 2013; 8: e82804.
- [7] Galzie Z, Kinsella AR, Smith JA. Fibroblast growth factors and their receptors. Biochem Cell Biol 1997; 75: 669-685.
- [8] Thisse B, Thisse C. Functions and regulations of fibroblast growth factor signaling during embryonic development. Dev Biol 2005; 287: 390-402.
- [9] Turner N, Lambros MB, Horlings HM, Pearson A, Sharpe R, Natrajan R, Geyer FC, van Kouwenhove M, Kreike B, Mackay A, Ashworth A, van de Vijver MJ, Reis-Filho JS. Integrative molecular profiling of triple negative breast cancers identifies amplicon drivers and potential therapeutic targets. Oncogene 2010; 29: 2013-2023.
- [10] Su WC, Kitagawa M, Xue N, Xie B, Garofalo S, Cho J, Deng C, Horton WA, Fu XY. Activation of stat1 by mutant fibroblast growth-factor receptor in thanatophoric dysplasia type ii dwarfism. Nature 1997; 386: 288-292.
- [11] Hart KC, Robertson SC, Kanemitsu MY, Meyer AN, Tynan JA, Donoghue DJ. Transformation and stat activation by derivatives of fgfr1, fgfr3, and fgfr4. Oncogene 2000; 19: 3309-3320.
- [12] Wong A, Lamothe B, Lee A, Schlessinger J, Lax I. Frs2 alpha attenuates fgf receptor signaling by grb2-mediated recruitment of the ubiquitin ligase cbl. Proc Natl Acad Sci U S A 2002; 99: 6684-6689.
- [13] Jean S, Mikryukov A, Tremblay MG, Baril J, Guillou F, Bellenfant S, Moss T. Extendedsynaptotagmin-2 mediates fgf receptor endocytosis and erk activation in vivo. Dev Cell 2010; 19: 426-439.
- [14] Yusoff P, Lao DH, Ong SH, Wong ES, Lim J, Lo TL, Leong HF, Fong CW, Guy GR. Sprouty2 inhibits the ras/map kinase pathway by inhibiting the activation of raf. J Biol Chem 2002; 277: 3195-3201.
- [15] Tiong KH, Mah LY, Leong CO. Functional roles of fibroblast growth factor receptors (fgfrs) signaling in human cancers. Apoptosis 2013; 18: 1447-1468.
- [16] Rezvani M, Wilde J, Vitt P, Mailaparambil B, Grychtol R, Krueger M, Heinzmann A. Association of a fgfr-4 gene polymorphism with bronchopulmonary dysplasia and neonatal respiratory distress. Dis Markers 2013; 35: 633-640.
- [17] Ma Z, Tsuchiya N, Yuasa T, Inoue T, Kumazawa T, Narita S, Horikawa Y, Tsuruta H, Obara T,

Saito M, Satoh S, Ogawa O, Habuchi T. Polymorphisms of fibroblast growth factor receptor 4 have association with the development of prostate cancer and benign prostatic hyperplasia and the progression of prostate cancer in a japanese population. Int J Cancer 2008; 123: 2574-2579.

- [18] Xu W, Li Y, Wang X, Chen B, Wang Y, Liu S, Xu J, Zhao W, Wu J. Fgfr4 transmembrane domain polymorphism and cancer risk: A meta-analysis including 8555 subjects. Eur J Cancer 2010; 46: 3332-3338.
- [19] FitzGerald LM, Karlins E, Karyadi DM, Kwon EM, Koopmeiners JS, Stanford JL, Ostrander EA. Association of fgfr4 genetic polymorphisms with prostate cancer risk and prognosis. Prostate Cancer Prostatic Dis 2009; 12: 192-197.
- [20] Agarwal D, Pineda S, Michailidou K, Herranz J, Pita G, Moreno LT, Alonso MR, Dennis J, Wang Q, Bolla MK, Meyer KB, Menéndez-Rodríguez P, Hardisson D, Mendiola M, González-Neira A, Lindblom A, Margolin S, Swerdlow A, Ashworth A, Orr N, Jones M, Matsuo K, Ito H, Iwata H, Kondo N; kConFab Investigators; Australian Ovarian Cancer Study Group, Hartman M, Hui M, Lim WY, Iau PT, Sawyer E, Tomlinson I, Kerin M, Miller N, Kang D, Choi J-, Park SK, Noh D-, Hopper JL, Schmidt DF, Makalic E, Southey MC, Teo SH, Yip CH, Sivanandan K, Tay W-, Brauch H, Brüning T, Hamann U; GENICA Network, Dunning AM, Shah M, Andrulis IL, Knight JA, Glendon G, Tchatchou S, Schmidt MK, Broeks A, Rosenberg EH, van't Veer LJ, Fasching PA, Renner SP, Ekici AB, Beckmann MW, Shen C-, Hsiung C-, Yu J-, Hou M-, Blot W, Cai Q, Wu AH, Tseng C-, Van Den Berg D, Stram DO, Cox A, Brock IW, Reed MW, Muir K, Lophatananon A, Stewart-Brown S, Siriwanarangsan P, Zheng W, Deming-Halverson S, Shrubsole MJ, Long J, Shu X-, Lu W, Gao Y-, Zhang B, Radice P, Peterlongo P, Manoukian S, Mariette F, Sangrajrang S, McKay J, Couch FJ, Toland AE; TNBCC, Yannoukakos D, Fletcher O, Johnson N, dos Santos Silva I, Peto J, Marme F, Burwinkel B, Guénel P, Truong T, Sanchez M, Mulot C, Bojesen SE, Nordestgaard BG, Flyer H, Brenner H, Dieffenbach AK, Arndt V, Stegmaier C, Mannermaa A, Kataja V, Kosma V-, Hartikainen JM, Lambrechts D, Yesilyurt BT, Floris G, Leunen K, Chang-Claude J, Rudolph A, Seibold P, Flesch-Janys D, Wang X, Olson JE, Vachon C, Purrington K, Giles GG, Severi G, Baglietto L, Haiman CA, Henderson BE, Schumacher F, Marchand LL, Simard J, Dumont M, Goldberg MS, Labréche F, Wingvist R, Pylkäs K, Jukkola-Vuorinen A, Grip M, Devilee P, Tollenaar RA, Seynaeve C, García-Closas M, Chanock SJ, Lissowska J, Figueroa

JD, Czene K, Eriksson M, Humphreys K, Darabi H, Hooning MJ, Kriege M, Collée JM, Tilanus-Linthorst M, Li J, Jakubowska A, Lubinski J, Jaworska-Bieniek K, Durda K, Nevanlinna H, Muranen TA, Aittomäki K, Blomqvist C, Bogdanova N, Dörk T, Hall P, Chenevix-Trench G, Easton DF, Pharroah PD, Arias-Perez JI, Zamora P, Benítez J, Milne RL. Fgf receptor genes and breast cancer susceptibility: results from the Breast Cancer Association Consortium. Br J Cancer 2014; 110: 1088-1100.

- [21] Sheu MJ, Hsieh MJ, Chiang WL, Yang SF, Lee HL, Lee LM, Yeh CB. Fibroblast growth factor receptor 4 polymorphism is associated with liver cirrhosis in hepatocarcinoma. PLoS One 2015; 10: e0122961.
- [22] Jacquemier J, Adelaide J, Parc P, Penault-Llorca F, Planche J, deLapeyriere O, Birnbaum D. Expression of the fgfr1 gene in human breast-carcinoma cells. Int J Cancer 1994; 59: 373-378.
- [23] Jaakkola S, Salmikangas P, Nylund S, Partanen J, Armstrong E, Pyrhonen S, Lehtovirta P, Nevanlinna H. Amplification of fgfr4 gene in human breast and gynecological cancers. Int J Cancer 1993; 54: 378-382.

- [24] Bange J, Prechtl D, Cheburkin Y, Specht K, Harbeck N, Schmitt M, Knyazeva T, Muller S, Gartner S, Sures I, Wang H, Imyanitov E, Haring HU, Knayzev P, Iacobelli S, Hofler H, Ullrich A. Cancer progression and tumor cell motility are associated with the fgfr4 arg (388) allele. Cancer Res 2002; 62: 840-847.
- [25] Sugiyama N, Varjosalo M, Meller P, Lohi J, Chan KM, Zhou Z, Alitalo K, Taipale J, Keski-Oja J, Lehti K. Fgf receptor-4 (fgfr4) polymorphism acts as an activity switch of a membrane type 1 matrix metalloproteinase-fgfr4 complex. Proc Natl Acad Sci U S A 2010; 107: 15786-15791.
- [26] Sugiyama N, Varjosalo M, Meller P, Lohi J, Hyytiainen M, Kilpinen S, Kallioniemi O, Ingvarsen S, Engelholm LH, Taipale J, Alitalo K, Keski-Oja J, Lehti K. Fibroblast growth factor receptor 4 regulates tumor invasion by coupling fibroblast growth factor signaling to extracellular matrix degradation. Cancer Res 2010; 70: 7851-7861.