# Review Article Prognostic role of microRNA 125b invarious cancers: a meta-analysis

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**Abstract:** Objectives: Recently, lots of studies have demonstrated that microRNAs (miRNA) exhibit altered expression in various cancers and maybe a prognostic biomarker of cancers. We performed a meta-analysis to evaluate the impact of miR-125b expressionin solid tumors on patients' overall survival (OS), release-free survival (RFS) and progress-free survival (DFS). Design: Meta-analysis: Data sources and study eligibility criteria: Studies were identified by searching PubMed, Embace, and Cochrane Library and were assessed by further quality evaluation. The pooled hazard ratios (HRs) with 95% confidence intervals (CIs) were calculated to investigate the association between miR-125 bexpression and cancer patients OS, RFS and PFS. Results: Our analysis results showed that miR-125b predicted poorOS (HR = 1.14 (95% CI: 0.77-1.69), RFS (HR = 2.48, 95% CI 1.43-4.30) and PFS (HR = 1.00, 95% CI 0.45-2.23). The subgroups showed miR-125b was significantly associated with worse OS in gastric carcinoma (HR = 1.61, 95% CI 1.05-2.49; *P* < 0.001) and hepatocellular carcinoma (HR = 1.74, 95% CI 1.02-2.97; *P* < 0.001). Conclusion: The findings from this meta-analysis suggest that miR-125b could be a useful clinical prognostic biomarker of human cancers.

Keywords: Cancer, miR-125b, prognosis, meta-analysis

#### Introduction

MicroRNAs are a class of endogenous small noncoding RNAs, which had the length of 19~25 nucleotides and mainly regulate the mRNAs and the expression level of their target proteins by directly binding with the corresponding mRNAs on the 3' UTR or 5' UTR [1]. It is reported that miRNAs have an important role in the development of a variety of human diseases [2-4].

MiR-125b is validated to be transcribed from two loci situated on chromosomes 11q23 and 21q21 and its product is hsa-miR-125b-1 and hsa-miR-125b-2 respectively [5]. It is reported that miR-125b was involved in various tumor development [6-10]. Previous studies show that miR-125b suppresses the proliferation and migration of osteosarcoma cells through down-regulation of STAT3 [11], but promotes proliferation through down-regulation of Bak1 in prostate cancer cells [10]. These data suggest that miR-125b might act as a tumor suppressor or oncogene depending on he cellular context.

The majority of cancers are often a serious problem for the clinical problem, Henceit is imperative for us to identify of predictive biomarkers to improve treatment of patients with various cancers [12]. From the different studies miR-125b maybe act as a significant tumor biomarker and a potential therapeutic target [13]. However, the result from single study is not enough to evaluate whether miR-125b can be considered as a promising biomarker. So we collected the date and performed meta-analysis to assess the prognostic value of miR-125 levels in different cancers.

#### Materials and methods

#### Search strategy

We performed a meta-analysis in accordance with the guidelines of observational studies in epidemiology (MOOSE) [14]. We searched the



studies from PubMed, Embace, and Cochrane Library. The search strategy was "microRNA-125b OR miR-125b" AND "tumor OR neoplasm OR cancer OR "carcinoma" and "survival OR prognosis OR outcome". The database search was carried out by two authors (Quanhui Mei and Ruizheng Shi). The disagreements were resolved by consensus.

# Inclusion and exclusion criteria

Eligible studies included in this meta-analysis according to the following criteria: (1) it reported miR-125b expression by the specific methods in tumor tissue or blood; (2) it invested the association between miR-125b expression and survivaloutcome; (3) it reported sufficient data to estimate the hazard ratio (HR) and 95% confidence intervals (CI) according to miR-125b expression. The candidate articles were screenedby author (Quanhui Mei). Articles were excludeif they were (1) a case reports; (2) letters and reviews; (3) animal trails; (4) hematological malignancies and autoimmune disorders; (5) or lack of important information such as hazard ratio (HR), 95% CI and P value. The full texts of the articles were carefully examined for comprehensive evaluation. The whole process was supervised by Ruizheng Shi.

### Quality assessment

All the included studies was performed independently by two investigators (Quanhui Mei and Ruizheng Shi), based on the critical guidelines of the Dutch Cochrane Centre proposed by MOOSE for prognostic meta-analyses. The articles should included a basic criteria as follows: (1) clear report of study population and country; (2) clear definition of type of cancer; (3) clear demonstrated the outcome assessment; (4) clear definition of measurement of miR-125b. Otherwise the studies were excluded for the reason of the quality of the meta-analysis.

# Data extraction

Two reviewers independently extracted the required information from all eligible studies to rule out any discrepancy. The following data were extracted: first author, study of year, type of carcinoma, source of patients, number of patients, method of testing miR-125b, and HR of miR-125b for overall survival (OS), Releasefree survival (RFS) and Progression free survival (PFS), as well as the corresponding 95% CI. If the HR and CI were not reported directly, the total observed events and the numbers of patients in each group were extracted to calculate HR and its variance indirectly [15]. If only Kaplan-Meier curves are available, data was extracted from the graphical survival plots by Engauge Digitizer version 4.1. Only reported univariate analysis results for survival in eligible studies were considered for the aggregation of the survival data. All the data were resolved by consensus.

# Statistical analysis

Statistical heterogeneity was assessed by visual inspection of forest plots, by performing the Chisquaretest (assessing the *P* value) and calculating the  $I^2$  statistic [16]. If the *P* value was less than 0.05 and/or  $I^2$  exceeded 50%, indicat-

Study	Year	Туре	Country	Sample	Survival	Obtain	Method	HR(CI)
Li WX [17]	2008	Hepatocellular carcinoma	China	75	OS	Original	qRT-PCR	OS: 2.43 (1.13-5.24)
Naohiro Nishida [18]	2010	Colorectal cancer	Japan	89	OS	Engauge	qRT-PCR	OS: 1.6 (0.52-4.90)
Zhang Y [19]	2011	Breast cancer	China	105	OS	Engauge	qRT-PCR	OS: 0.49 (0.19-1.26)
Feng JJ [20]	2012	glioma	America	277	OS	Original	qRT-PCR	OS: 1.08 (0.86-1.35)
Ma YX [21]	2012	Non-small cell lung cancer	China	193	OS	Original	qRT-PCR	OS: 2.46 (1.80-3.38)
Song FJ [22]	2013	Gastric cancer	China	358	OS+PFS	Original	qRT-PCR	OS: 1.8 (1.19-2.72) PFS: 1.76 (1.2-2.58)
M Shiiba [23]	2013	Oral squamous cell carcinoma	Japan	50	OS	Engauge	qRT-PCR	OS: 0.77 (0.1-6.19)
Federico Cappuzzo [24]	2014	Colorectal cancer	Italy	183	OS+PFS	Calculate	microarray	OS: 1.8 (1.19-2.72) PFS: 0.68 (0.43-1.07)
Li XX [25]	2014	Glioma	China	45	OS	Original	qRT-PCR	OS: 2.43(1.13-5.24)
Felice H Tsang [26]	2014	Hepatocellular carcinoma	China	49	OS+RFS	Engauge	qRT-PCR	OS: 1.28 (0.2-8.36) RFS: 1.51 (0.42-5.42)
Wu JG [27]	2014	Gastric cancer	China	149	OS	Engauge	qRT-PCR	OS: 1.03 (0.4-2.67)
Fu Q [28]	2014	Clear-cell renal cell carcinoma	China	259	RFS	Original	qRT-PCR	OS: 1.8 (1.19-2.72) RFS: 2.40 (1.37-3.78)
Yu XZ [29]	2015	None-small cell lung cancer	China	42	PFS	Engauge	qRT-PCR	PFS: 0.94 (0.38-2.36)
Vilquin P [30]	2015	Breast cancer	France	65	RFS	Original	qRT-PCR	RFS: 6.8 (1.35-35.26)

#### Table 1. Characteristics of studies



Figure 2. Meta-analysis of miR-125b expression and solid tumors' overall survival.

ing the presence of heterogeneity, a randomeffects model (the DerSimonian-Laird method) was used. Otherwise, the fixed-effects model (the Mante-Haenszel method) was used. Subgroup analysis was further performed to explore the source of heterogeneity. Heterogeneity was defined as P < 0.10 or  $I^2 > 50\%$ . Subgroup and sensitivity analysis was carried out by investigating the influence of a single study on the overall HR. Furthermore, Begg's test was performed to provide quantitative evidence of publication bias. All analyses were performed using STATAvision 12.0 (Stata Corporation, College Station, TX, USA).

Study Overall Survival of subgroups ID Sorting by type	HR (95% CI)	% Weight
colorectal cancer NAOH IRO NISHIDA (2010) Federico Cappuzzo (2014) Subtotal (I-squared = 47.9%, p = 0.166)	1.60 (0.52, 4.90 0.68 (0.43, 1.07 0.89 (0.41, 1.94	) 5.84 ) 12.51 ) 18.34
gastric cancer Fengju Song (2013) Jian-Guo Wu (2014) Subtotal (I-squared = 10.5%, p = 0.291)	1.80 (1.19, 2.72 1.03 (0.40, 2.67 1.61 (1.05, 2.49	) 13.03 ) 7.13 ) 20.15
gliofna Junjie Feng (2012) Xinxing Li (2014) Subtotal (I-squared = 74.7%, p = 0.047)	1.08 (0.86, 1.35 2.43 (1.13, 5.24 1.49 (0.68, 3.23	) 15.08 ) 8.84 ) 23.92
hepatocellular carcinom a Wenxi Li (2008) Felice H. Tsang (2014) Subtotal (I-squared = 0.0%, p = 0.737)	1.79 (1.02, 3.13 - 1.28 (0.20, 8.36 1.74 (1.02, 2.97	) 11.20 ) 2.74 ) 13.94
breast cancer Yan Zhang (2011) Subtotal (I-squared = .%, p = .)	0.49 (0.19, 1.26 0.49 (0.19, 1.26	) 7.16 ) 7.16
non-small cell lung cancer Ma Yuxia (2012) Subtotal (I-squared = .%, p = .)	2.46 (1.80, 3.38 2.46 (1.80, 3.37	) 14.18 ) 14.18
oral squamous cell carcinoma M Shiiba (2013) Subtotal (I-squared = .%, p = .)	0.77 (0.10, 6.19 0.77 (0.10, 6.06	) 2.31 ) 2.31
Overall (I-s quared = 72.6%, p = 0.000) NOTE: Weights are from random effects analysis	1.34 (0.95, 1.88	) 100.00
.0979 1	10.2	

Figure 3. Subgroups analysis of miR-125b expression and solid tumors' overall survival.

#### Results

#### Study characteristics

According to the criteria mentioned in materials and methods, 499 abstracts were initially selected. However, 452 irrelevant abstracts were excluded by the first choosing. Fortyseven full-text articles were reviewed for further evaluation and twenty-seven were excluded because they were the solid tumor. The remaining twenty articles had further read and six articles were excluded because of not including the OS analysis. At last we selected fourteen articles including 1939 patients, which were published between 2008 and 2015 (Figure 1). The category of cancers included breast cancer (2 studies), gastric cancer (2 studies), hepatocellular carcinoma (2 studies), colorectal cancer (2 studies), non-small cell lung cancer (2 studies), glioma (2 studies), oral squamous cell carcinoma and clear-cell renal cell carcinoma. Quantitative RT-PCR and microarray were used to detect miRNAs expression in all studies. HRs wereestimated in 6 studies by engauge software and directly reported in five studies. Themajor characteristics of the 14 eligible studies are listed in **Table 1**.

# Overall survival (OS) associated with miR-125b expression

For studies evaluating OS for miR-125b, a random-effects model was used to calculate the pooled HR and its 95% CI because of the high significant heterogeneity ( $I^2 = 72.6\%$ , P =0.000). The result demonstrated that high level of miR-125b may predict poorer OS, with the pooled HR being 1.14 (**Figure 2**). The subgroups were analyzed according to the main characteristics such as tumor type. In the subgroup of tumor type, we found the miR-125b was signifi-



Figure 4. Meta-analysis of miR-125b expression and solid tumors' RFS/PFS.



Figure 5. Subgroups analysis of miR-125b expression and solid tumors' RFS/PFS.



Figure 6. Begg's funnel plot of publication bias.

cantly associated with worse OS in gastric carcinoma (HR = 1.61, 95% Cl 1.05-2.49; P < 0.001; fixed-effects model), hepatocellular carcinoma (HR = 1.74, 95% Cl 1.02-2.97; P < 0.001; fixed-effects model), without any heterogeneity in thedata ( $I^2 = 10.5\%$ , P = 0.291;  $I^2 = 0.0\%$ , P = 0.737, resp) (**Figure 3**).

# Tumor progression (RFS/PFS) associated with miR-125b expression

We analyzed tumor progression associated with miR-125b expression. Six studies included the RFS and PFS analysis. Meta-analysis of the eligible studies predicted that high level of miR-125 was significantly associated with poor DFS/PFS (pooled HR = 1.48, 95% Cl: 0.79-2.76). There was significant heterogeneity was observed ( $l^2 = 80.4\%$ , P = 0.000) and the random-effects model was applied (**Figure 4**). In the subgroup of tumor type, we found the miR-125b was significantly associated with worse RFS (HR = 2.48, 95% Cl 1.43-4.30; P < 0.001; fixed-effects model) (**Figure 5**).

#### Heterogeneity and publication bias analysis

Sensitivity analysis was performed by deletion of individual studies using the fixed-effects model. By excluding this study from the analysis, similar pooled HR and significance were obtained.Bgger's test wasused to evaluate the publication bias (**Figure 6**). The *P* values of Begg's tests wasover 0.05 (P = 0.15). Hence, there was noevidence for significant publication bias in themeta-analysis.

#### Discussion

Cancer is a global and growing problem which is potentially life-threatening that should be

recognized immediately with decisive intervention in order to decreased mortality and morbidity. The signs and symptoms of tumor may present at any time. However, despite the advances technology was used to treatment the cancer, the five years survival was also lower mainly due to the late diagnosis and lack of prognostic markers for various cancers. There are few defined prognostic and diagnostic biomarkers available in cancers. So it is imperative for us to identify the newer biomarker of various cancer [31]. MiR-125b exhibits a large range of correlation with different cancers. Acting as tumor suppressor, miR-125b shows a lower expression in hepatocellular carcinoma, chronic lymphocytic leukemia, cutaneous squamous cell carcinoma, melanoma, Ewing's sarcoma, bladder cancer head and neck cancer. As a tumor promoter, miR-125b increased carcinogenesis in B-cell leukemia, myeloid leukemia, non-small cell lung cancer, clear-cell renal carcinoma, glioblastoma, prostate cancer, pancreatic cancer and oligodendroglia, in which miR-125 is overexpressed [28, 32-53]. In a word, miR-125b can be acted as a different role in the tumors. MiR-125b can regulate thetumor cell proliferation, apoptosis, invasion and metastasis. For example, Liu LH et al [11] found miR-125b might inhibit tumor cell proliferation by down-regulating STAT3. MiR-125b also influenced the expression of survivin protein by modulating the STAT3 signaling. Such regulation accelerates tumor cell apoptosis [54].

In terms of this, a total of 1939 participants from 14 studies were included into the metaanalysis. This result showed that high expression of miR-125b maybepredict a unsatisfactory outcome of some cancers (HR = 1.14, 2.48, 1.00 for OS, RFS, and PFS, resp). For OS, the data displayed that miR-125b was anundesirably prognostic marker ingastric carcinoma (HR = 1.61, 95% Cl 1.05-2.49; P < 0.001) and hepatocellular carcinoma (HR = 1.74, 95% Cl 1.02-2.97; P < 0.001). Additionally, there was no obvious riskof publication bias in our metaanalysis. From the aboveresults, we found that high expression of tissue miR-125b was a negative prognostic factor in some cancer patients.

Although the present meta-analysis showed that theexpression of miR-125b maybe play a worse role in the prognosis in several cancers, some limitation was still in this meta-analysis. Firstly, the obvious heterogeneity existedin our meta-analysis. Secondly, several HRs were unreported in the original article that we have to calculate the HR from the survival curve. Thirdly, the number of studies in subgroup analyses was relativelysmall. More studies on these cancers are needed in the future.

In sum, in this meta-analysis, we got a concluded that miR-125b was acted as a biomarker in various carcinomas. Increased miR-125b level incancerous tissues was associated with unsatisfactory OS, PFS and RFS. However, our results should be regarded attention because of the limitations of the present analysis listedabove. There should be moremulticenter studies needed to focus on the relationship between miR-125b and cancer prognosis.

#### Disclosure of conflict of interest

None.

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#### References

- Carthew RW and Sontheimer EJ. Origins and mechanisms of miRNAs and siRNAs. Cell 2009; 136: 642-655.
- [2] Xiao C and Rajewsky K. MicroRNA control in the immune system: basic principles. Cell 2009; 136: 26-36.
- [3] Koralov SB, Muljo SA, Galler GR, Krek A, Chakraborty T, Kanellopoulou C, Jensen K, Cobb BS, Merkenschlager M, Rajewsky N and Rajewsky K. Dicer ablation affects antibody di-

versity and cell survival in the B lymphocyte lineage. Cell 2008; 132: 860-874.

- [4] Lujambio A and Lowe SW. The microcosmos of cancer. Nature 2012; 482: 347-355.
- [5] Hall KN, Wakeman MA, Levy RC and Khoury J. Factors associated with career longevity in residency-trained emergency physicians. Ann Emerg Med 1992; 21: 291-297.
- [6] Bousquet M, Harris MH, Zhou B and Lodish HF. MicroRNA miR-125b causes leukemia. Proc Natl Acad Sci U S A 2010; 107: 21558-21563.
- [7] Klusmann JH, Li Z, Bohmer K, Maroz A, Koch ML, Emmrich S, Godinho FJ, Orkin SH and Reinhardt D. miR-125b-2 is a potential oncomiR on human chromosome 21 in megakaryoblastic leukemia. Genes Dev 2010; 24: 478-490.
- [8] Le MT, Teh C, Shyh-Chang N, Xie H, Zhou B, Korzh V, Lodish HF and Lim B. MicroRNA-125b is a novel negative regulator of p53. Genes Dev 2009; 23: 862-876.
- [9] Zhou M, Liu Z, Zhao Y, Ding Y, Liu H, Xi Y, Xiong W, Li G, Lu J, Fodstad O, Riker Al and Tan M. MicroRNA-125b confers the resistance of breast cancer cells to paclitaxel through suppression of pro-apoptotic Bcl-2 antagonist killer 1 (Bak1) expression. J Biol Chem 2010; 285: 21496-21507.
- [10] Shi XB, Xue L, Yang J, Ma AH, Zhao J, Xu M, Tepper CG, Evans CP, Kung HJ and deVere White RW. An androgen-regulated miRNA suppresses Bak1 expression and induces androgen-independent growth of prostate cancer cells. Proc Natl Acad Sci U S A 2007; 104: 19983-19988.
- [11] Liu LH, Li H, Li JP, Zhong H, Zhang HC, Chen J and Xiao T. miR-125b suppresses the proliferation and migration of osteosarcoma cells through down-regulation of STAT3. Biochem Biophys Res Commun 2011; 416: 31-38.
- [12] Iorio MV and Croce CM. MicroRNA dysregulation in cancer: diagnostics, monitoring and therapeutics. A comprehensive review. EMBO Mol Med 2012; 4: 143-159.
- [13] Ebrahimi F, Gopalan V, Smith RA and Lam AK. miR-126 in human cancers: clinical roles and current perspectives. Exp Mol Pathol 2014; 96: 98-107.
- [14] Stroup DF, Berlin JA, Morton SC, Olkin I, Williamson GD, Rennie D, Moher D, Becker BJ, Sipe TA and Thacker SB. Meta-analysis of observational studies in epidemiology: a proposal for reporting. Meta-analysis Of Observational Studies in Epidemiology (MOOSE) group. JAMA 2000; 283: 2008-2012.
- [15] Tierney JF, Stewart LA, Ghersi D, Burdett S and Sydes MR. Practical methods for incorporating summary time-to-event data into meta-analysis. Trials 2007; 8: 16.

- [16] DerSimonian R and Laird N. Meta-analysis in clinical trials. Control Clin Trials 1986; 7: 177-188.
- [17] Li W, Xie L, He X, Li J, Tu K, Wei L, Wu J, Guo Y, Ma X, Zhang P, Pan Z, Hu X, Zhao Y, Xie H, Jiang G, Chen T, Wang J, Zheng S, Cheng J, Wan D, Yang S, Li Y and Gu J. Diagnostic and prognostic implications of microRNAs in human hepatocellular carcinoma. Int J Cancer 2008; 123: 1616-1622.
- [18] Nishida N, Yokobori T, Mimori K, Sudo T, Tanaka F, Shibata K, Ishii H, Doki Y, Kuwano H and Mori M. MicroRNA miR-125b is a prognostic marker in human colorectal cancer. Int J Oncol 2011; 38: 1437-1443.
- [19] Zhang Y, Yan LX, Wu QN, Du ZM, Chen J, Liao DZ, Huang MY, Hou JH, Wu QL, Zeng MS, Huang WL, Zeng YX and Shao JY. miR-125b is methylated and functions as a tumor suppressor by regulating the ETS1 proto-oncogene in human invasive breast cancer. Cancer Res 2011; 71: 3552-3562.
- [20] Feng J, Kim ST, Liu W, Kim JW, Zhang Z, Zhu Y, Berens M, Sun J and Xu J. An integrated analysis of germline and somatic, genetic and epigenetic alterations at 9p21.3 in glioblastoma. Cancer 2012; 118: 232-240.
- [21] Yuxia M, Zhennan T and Wei Z. Circulating miR-125b is a novel biomarker for screening nonsmall-cell lung cancer and predicts poor prognosis. J Cancer Res Clin Oncol 2012; 138: 2045-2050.
- [22] Song F, Yang D, Liu B, Guo Y, Zheng H, Li L, Wang T, Yu J, Zhao Y, Niu R, Liang H, Winkler H, Zhang W, Hao X and Chen K. Integrated microRNA network analyses identify a poor-prognosis subtype of gastric cancer characterized by the miR-200 family. Clin Cancer Res 2014; 20: 878-889.
- [23] Shiiba M, Shinozuka K, Saito K, Fushimi K, Kasamatsu A, Ogawara K, Uzawa K, Ito H, Takiguchi Y and Tanzawa H. MicroRNA-125b regulates proliferation and radioresistance of oral squamous cell carcinoma. Br J Cancer 2013; 108: 1817-1821.
- [24] Cappuzzo F, Sacconi A, Landi L, Ludovini V, Biagioni F, D'Incecco A, Capodanno A, Salvini J, Corgna E, Cupini S, Barbara C, Fontanini G, Crino L and Blandino G. MicroRNA signature in metastatic colorectal cancer patients treated with anti-EGFR monoclonal antibodies. Clin Colorectal Cancer 2014; 13: 37-45 e34.
- [25] Li X, Zheng J, Chen L, Diao H and Liu Y. Predictive and Prognostic Roles of Abnormal Expression of Tissue miR-125b, miR-221, and miR-222 in Glioma. Mol Neurobiol 2016; 53:577-83.
- [26] Tsang FH, Au V, Lu WJ, Shek FH, Liu AM, Luk JM, Fan ST, Poon RT and Lee NP. Prognostic marker microRNA-125b inhibits tumorigenic

properties of hepatocellular carcinoma cells via suppressing tumorigenic molecule eIF5A2. Dig Dis Sci 2014; 59: 2477-2487.

- [27] Wu JG, Wang JJ, Jiang X, Lan JP, He XJ, Wang HJ, Ma YY, Xia YJ, Ru GQ, Ma J, Zhao ZS and Zhou R. MiR-125b promotes cell migration and invasion by targeting PPP1CA-Rb signal pathways in gastric cancer, resulting in a poor prognosis. Gastric Cancer 2015; 18: 729-739.
- [28] Fu Q, Liu Z, Pan D, Zhang W, Xu L, Zhu Y, Liu H and Xu J. Tumor miR-125b predicts recurrence and survival of patients with clear-cell renal cell carcinoma after surgical resection. Cancer Sci 2014; 105: 1427-1434.
- [29] Yu X, Wei F, Yu J, Zhao H, Jia L, Ye Y, Du R, Ren X and Li H. Matrix metalloproteinase 13: a potential intermediate between low expression of microRNA-125b and increasing metastatic potential of non-small cell lung cancer. Cancer Genet 2015; 208: 76-84.
- [30] Vilquin P, Donini CF, Villedieu M, Grisard E, Corbo L, Bachelot T, Vendrell JA and Cohen PA. MicroRNA-125b upregulation confers aromatase inhibitor resistance and is a novel marker of poor prognosis in breast cancer. Breast Cancer Res 2015; 17: 13.
- [31] Paul D, Kumar A, Gajbhiye A, Santra MK and Srikanth R. Mass spectrometry-based proteomics in molecular diagnostics: discovery of cancer biomarkers using tissue culture. Biomed Res Int 2013; 2013: 783131.
- [32] Kappelmann M, Kuphal S, Meister G, Vardimon L and Bosserhoff AK. MicroRNA miR-125b controls melanoma progression by direct regulation of c-Jun protein expression. Oncogene 2013; 32: 2984-2991.
- [33] Guo X, Wu Y and Hartley RS. MicroRNA-125a represses cell growth by targeting HuR in breast cancer. RNA Biol 2009; 6: 575-583.
- [34] Nam EJ, Yoon H, Kim SW, Kim H, Kim YT, Kim JH, Kim JW and Kim S. MicroRNA expression profiles in serous ovarian carcinoma. Clin Cancer Res 2008; 14: 2690-2695.
- [35] Wang G, Mao W, Zheng S and Ye J. Epidermal growth factor receptor-regulated miR-125a-5p--a metastatic inhibitor of lung cancer. FEBS J 2009; 276: 5571-5578.
- [36] Yagishita S, Fujita Y, Kitazono S, Ko R, Nakadate Y, Sawada T, Kitamura Y, Shimoyama T, Maeda Y, Takahashi F, Takahashi K, Tamura T and Koizumi F. Chemotherapy-regulated microrna-125-her2 pathway as a novel therapeutic target for trastuzumab-mediated cellular cytotoxicity in small cell lung cancer. Mol Cancer Ther 2015; 14: 1414-1423.
- [37] Jiang L, Huang Q, Zhang S, Zhang Q, Chang J, Qiu X and Wang E. Hsa-miR-125a-3p and hsamiR-125a-5p are downregulated in non-small cell lung cancer and have inverse effects on invasion and migration of lung cancer cells. BMC Cancer 2010; 10: 318.

- [38] Wang M, Zhu X, Sha Z, Li N, Li D and Chen L. High expression of kinesin light chain-2, a novel target of miR-125b, is associated with poor clinical outcome of elderly non-small-cell lung cancer patients. Br J Cancer 2015; 112: 874-882.
- [39] Liang L, Wong CM, Ying Q, Fan DN, Huang S, Ding J, Yao J, Yan M, Li J, Yao M, Ng IO and He X. MicroRNA-125b suppressesed human liver cancer cell proliferation and metastasis by directly targeting oncogene LIN28B2. Hepatology 2010; 52: 1731-1740.
- [40] Shi XB, Xue L, Ma AH, Tepper CG, Kung HJ and White RW. miR-125b promotes growth of prostate cancer xenograft tumor through targeting pro-apoptotic genes. Prostate 2011; 71: 538-549.
- [41] Xu N, Zhang L, Meisgen F, Harada M, Heilborn J, Homey B, Grander D, Stahle M, Sonkoly E and Pivarcsi A. MicroRNA-125b down-regulates matrix metallopeptidase 13 and inhibits cutaneous squamous cell carcinoma cell proliferation, migration, and invasion. J Biol Chem 2012; 287: 29899-29908.
- [42] Nakanishi H, Taccioli C, Palatini J, Fernandez-Cymering C, Cui R, Kim T, Volinia S and Croce CM. Loss of miR-125b-1 contributes to head and neck cancer development by dysregulating TACSTD2 and MAPK pathway. Oncogene 2014; 33: 702-712.
- [43] Han Y, Liu Y, Zhang H, Wang T, Diao R, Jiang Z, Gui Y and Cai Z. Hsa-miR-125b suppresses bladder cancer development by down-regulating oncogene SIRT7 and oncogenic long noncoding RNA MALAT1. FEBS Lett 2013; 587: 3875-3882.
- [44] Hisaoka M, Matsuyama A, Nagao Y, Luan L, Kuroda T, Akiyama H, Kondo S and Hashimoto H. Identification of altered MicroRNA expression patterns in synovial sarcoma. Genes Chromosomes Cancer 2011; 50: 137-145.
- [45] Li J, You T and Jing J. MiR-125b inhibits cell biological progression of Ewing's sarcoma by suppressing the PI3K/Akt signalling pathway. Cell Prolif 2014; 47: 152-160.
- [46] Morelli E, Leone E, Cantafio ME, Di Martino MT, Amodio N, Biamonte L, Gulla A, Foresta U, Pitari MR, Botta C, Rossi M, Neri A, Munshi NC, Anderson KC, Tagliaferri P and Tassone P. Selective targeting of IRF4 by synthetic microR-NA-125b-5p mimics induces anti-multiple myeloma activity in vitro and in vivo. Leukemia 2015; 29:2173-83.
- [47] Haemmig S, Baumgartner U, Gluck A, Zbinden S, Tschan MP, Kappeler A, Mariani L, Vajtai I and Vassella E. miR-125b controls apoptosis and temozolomide resistance by targeting TNFAIP3 and NKIRAS2 in glioblastomas. Cell Death Dis 2014; 5: e1279.

- [48] Yin F, Zhang JN, Wang SW, Zhou CH, Zhao MM, Fan WH, Fan M and Liu S. MiR-125a-3p regulates glioma apoptosis and invasion by regulating Nrg1. PLoS One 2015; 10: e0116759.
- [49] Xu X, Jia R, Zhou Y, Song X, Wang J, Qian G, Ge S and Fan X. Microarray-based analysis: identification of hypoxia-regulated microRNAs in retinoblastoma cells. Int J Oncol 2011; 38: 1385-1393.
- [50] Ferretti E, De Smaele E, Po A, Di Marcotullio L, Tosi E, Espinola MS, Di Rocco C, Riccardi R, Giangaspero F, Farcomeni A, Nofroni I, Laneve P, Gioia U, Caffarelli E, Bozzoni I, Screpanti I and Gulino A. MicroRNA profiling in human medulloblastoma. Int J Cancer 2009; 124: 568-577.
- [51] Bousquet M, Quelen C, Rosati R, Mansat-De Mas V, La Starza R, Bastard C, Lippert E, Talmant P, Lafage-Pochitaloff M, Leroux D, Gervais C, Viguie F, Lai JL, Terre C, Beverlo B, Sambani C, Hagemeijer A, Marynen P, Delsol G, Dastugue N, Mecucci C and Brousset P. Myeloid cell differentiation arrest by miR-125b-1 in myelodysplastic syndrome and acute myeloid leukemia with the t(2;11)(p21;q23) translocation. J Exp Med 2008; 205: 2499-2506.
- [52] So AY, Sookram R, Chaudhuri AA, Minisandram A, Cheng D, Xie C, Lim EL, Flores YG, Jiang S, Kim JT, Keown C, Ramakrishnan P and Baltimore D. Dual mechanisms by which miR-125b represses IRF4 to induce myeloid and B-cell leukemias. Blood 2014; 124: 1502-1512.
- [53] Tili E, Michaille JJ, Luo Z, Volinia S, Rassenti LZ, Kipps TJ and Croce CM. The down-regulation of miR-125b in chronic lymphocytic leukemias leads to metabolic adaptation of cells to a transformed state. Blood 2012; 120: 2631-2638.
- [54] Ansell SM, Arendt BK, Grote DM, Jelinek DF, Novak AJ, Wellik LE, Remstein ED, Bennett CF and Fielding A. Inhibition of survivin expression suppresses the growth of aggressive non-Hodgkin's lymphoma. Leukemia 2004; 18: 616-623.