Original Article Serum metabolic changes in rats after conventional external beam radiotherapy by gas chromatography-mass spectrometry

Yan He^{1*}, Zhiyi Wang^{2*}, Ke Su³, Jing Zhang³, Zixia Lin³, Jingjing Mo³, Congcong Wen³, Lufeng Hu⁴, Qing Wu⁴

¹The Institute of Molecular Medicine, School of Optometry and Ophthalmology and Eye Hospital, Wenzhou Medical University, Wenzhou 325000, China; ²The Second Affiliated Hospital and Yuying Children's Hospital, Wenzhou Medical University, Wenzhou 325000, China; ³Laboratory Animal Centre, Wenzhou Medical University, Wenzhou 325035, China; ⁴The First Affiliated Hospital of Wenzhou Medical University, Wenzhou 325000, China. ^{*}Equal contributors.

Received December 13, 2015; Accepted March 30, 2016; Epub June 15, 2016; Published June 30, 2016

Abstract: Conventional external beam radiotherapy has been widely used in various clinical malignant and pain management applications. In this study, we developed a serum metabolomic method based on gas chromatography-mass spectrometry (GC-MS) to evaluate the effect of conventional external beam radiation on rats. Thirty rats were randomly divided to radiation group (600 lx, 800 lx) and control group. Radiation group were under radiation (600 lx, 800 lx) for 1 h. Blood samples were collected from the rats from the control group and radiation group at first, second and third days, respectively. Partial least squares-discriminate analysis (PLS-DA) revealed that radiation group increased; the level of d-Glucose of the 800 lx radiation group decreased at the first day. Compared to the control group, the level of propanoic acid of the 600 lx radiation group increased; the level of propanoic acid and ethanedioic acid of the 600 lx radiation group increased at the second day. Compared to the control group, the level of propanoic acid and ethanedioic acid of the 600 lx radiation group increased; the level of propanoic acid and ethanedioic acid of the 600 lx radiation group increased; the level of propanoic acid of the 600 lx radiation group increased at the second day. Compared to the control group, the level of propanoic acid of the 600 lx radiation group increased; the level of d-Glucose of the 800 lx radiation group increased; the level of d-Glucose of the 800 lx radiation group increased; the level of d-Glucose of the 800 lx radiation group increased; the level of d-Glucose of the 800 lx radiation group decreased at the first day. The results indicate that metabolomic methods based on GC-MS may be useful to elucidate effect of radiation on rat through the exploration of biomarkers (propanoic acid, d-Glucose, ethanedioic acid).

Keywords: Metabolomics, GC/MS, radiation, rat

Introduction

Radiotherapy is the treatment of disease, esp cancer, by means of alpha or beta particles emitted from an implanted or ingested radioisotope, or by means of a beam of high-energy radiation [1-3]. In recent years, radiotherapy has been widely used in various clinical malignant and pain management applications [4, 5]. However, its inevitable and invisible damage to our bodies cannot be ignored, especially the radiation induced liver disease (RILD), which is mainly fatal complications secondary to radiotherapy [6-8]. Therefore, more and more efficient methods are applied to assess the injury severity as well as to take proper measures.

Analytical sensitivity is increasing with the use of new noninvasive methods. One of these is

metabolomics, which appears useful for finding specific biomarkers of radiation [9-11]. Metabolomics is an emerging field with great potential for radiation biodosimetry for the fact that blood cells and serum have proven to be abundant sources of human radiation biomarkers [12-15]. To date, few studies interiorly use metabonomics method in radiation damagerelated researches. In this study, we have harnessed the gas chromatography-mass spectrometry and various multivariate data analyses to uncover metabolomic responses in irradiated mice.

Material and methods

Chemicals and animals

Trimethylchlorosilane (TMCS) and N-Methyl-N-(trimethylsilyl) trifluoroacetamide (MSTFA) we-

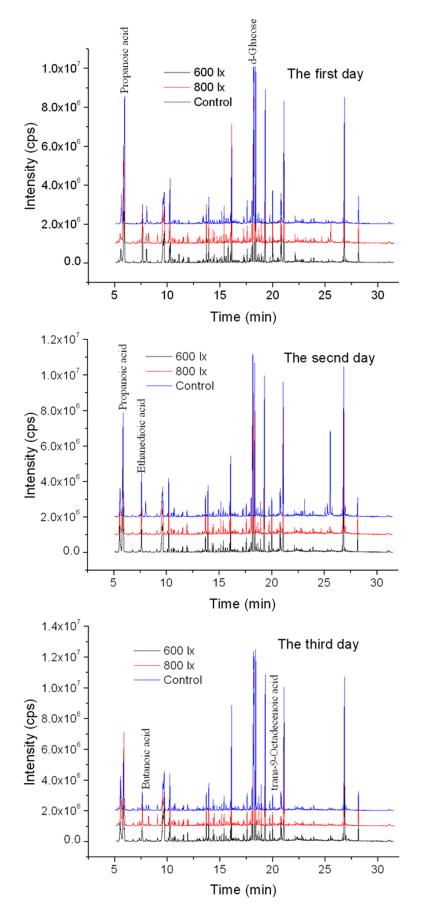


Figure 1. Typical GC-MS total ion chromatogram of rat serum after radiation at the first day, the second day and the third day.

re purchased from Sigma-Aldrich (Shanghai, China). HPLC-grade n-heptane and acetonitrile were purchased from Tedia Reagent Company (Shanghai, China). Pyridine and methylhydroxylamine hydrochloride were purchased from Aladdin Industrial, Inc. (Shanghai, China). Sprague-Dawley rats (male, 220±20 g) were purchased from Shanghai SLAC Laboratory Animal Co., Ltd.

Instrumentation and conditions

Agilent 6890N-5975B GC/ MS, HP-5MS (0.25 mm×30 m×0.25 mm), were from Agilent Company (Santa Clara, California, USA). The temperature was then gradually increased to 260°C at a rate of 10°C/min, and then kept at 260°C for 10 minutes. The GC oven was initially set at 80°C and was kept at this temperature for 5 minutes. Mass detection was conducted first in El mode with electron energy of 70 eV, then in full-scan mode with m/z 50-550, and finally, by splitless mode injection [16, 17].

Sample preparation

The 250 μ L of acetonitrile was added to 100 μ L of serum, kept in an ice-bath for 15 min, and then were centrifuged at 10000 g for 10 minutes at 4°C. The 150 μ L of the supernatant was transferred to a GC vial and evaporated to dryness under a stream of nitrogen gas. Methoximation was carried out at 70°C for 24 h after 50 μ L of methylhydroxyl-

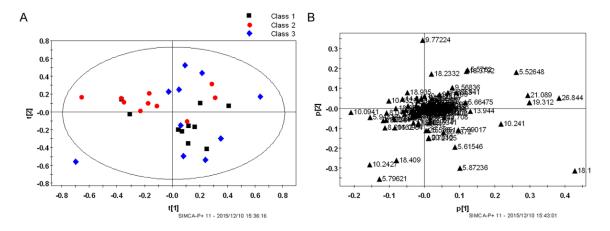


Figure 2. PCA score results of rat serum samples (A), after radiation (600 lx, Class 1; 800 lx, Class 2), Control (Class 3) at the first day; the corresponding load diagram (B).

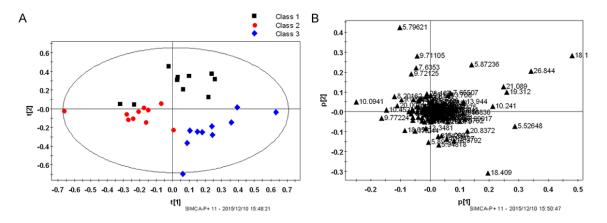


Figure 3. PLS-DA score results of rat serum samples (A), after radiation (600 lx, Class 1; 800 lx, Class 2), Control (Class 3) at the first day; the corresponding load diagram (B).

amine hydrochloride (15 mg/mL in pyridine) was added. The 50 μ L MSTFA (with 1% TMCS as the catalyst) was added and kept at 70°C for another hour, and then vortexed after adding 150 μ L n-heptane [17].

Metabolomics study

Rats were housed under a natural light-dark cycle conditions with controlled temperature (22°C). All thirty rats were housed at Laboratory Animal Research Center of Wenzhou Medical University. All experimental procedures were approved ethically by the Administration Committee of Experimental Animals of Wenzhou Medical University.

Thirty rats $(220\pm20 \text{ g})$ were randomly divided to radiation group (600 lx, 800 lx) and control group. Radiation group were under radiation (600 lx, 800 lx) for 1 h. Blood samples were collected from the rats from the control group and radiation group at first, second and third days, respectively. The blood samples were collected and then centrifuged at 8000 g for 10 min at 4°C. The serum was stored at -80°C until measurement.

Data analysis

The GC/MS data was exported into Microsoft Excel, with the peaks normalized to the total sum of spectrum prior to multivariate analyses. The resulting data was processed through principal component analysis (PCA) and partial least squares discriminate analysis (PLS-DA) using SIMCA-P 11.5 software (Umetrics, Umea, Sweden).

Statistical analysis

Statistical analysis was carried out using SPSS software (Version 18.0, SPSS). Independent samples T-test was applied in order to detect

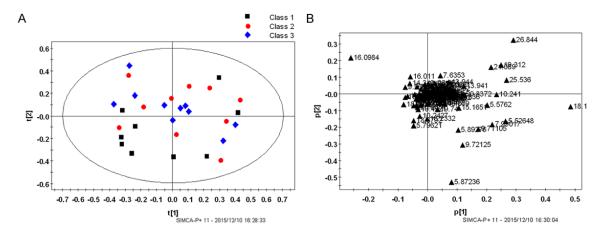


Figure 4. PCA score results of rat serum samples (A), after radiation (600 lx, Class 1; 800 lx, Class 2), Control (Class 3) at the second day; the corresponding load diagram (B).

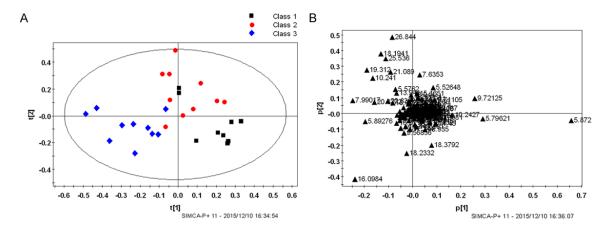


Figure 5. PLS-DA score results of rat serum samples (A), after radiation (600 lx, Class 1; 800 lx, Class 2), Control (Class 3) at the second day; the corresponding load diagram (B).

significant differences in all metabolites between two groups. A *P* value of <0.05 was considered statistically significant.

Results and discussion

Metabolomics study

Metabolomics is a newly emerging omics approach to the investigation of metabolic phenotype changes induced by environmental or endogenous factors [18-23]. It has shown promising results in healthcare fields, especially in disease diagnosis and drug-toxicity assessment, as reviewed recently [24, 25].

Figure 1 provides the typical metabolic profiles of serum at first, second and third day acquired through GC-MS technique. Metabolic profile data pretreatment resulted in a final dataset consisting of eighty metabolic features from GC-MS analyses. The endogenous metabolites in the serum were identified according to NIST 2005 mass spectrometry database.

In order to explore the metabolic profile changes of rats in radiation group (600 lx, 800 lx), we compared the GC-MS spectrum of PCA of the radiation group (600 lx, 800 lx) with the rats in the control group (**Figures 2A**, **4A** and **6A**), the corresponding load diagram was shown in **Figures 2B**, **4B** and **6B**. The PLS-DA of the radiation group (600 lx, 800 lx) with the rats in the control group (**Figures 3A**, **5A** and **7A**), the corresponding load diagram was shown in **Figures 3B**, **5B** and **7B**. **Figures 3A**, **5A** and **7A** PLS-DA score chart showed that the first principal components of the rats in the radiation group (600

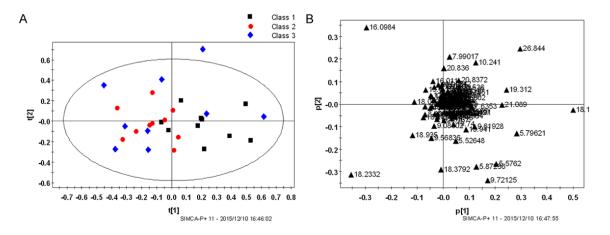


Figure 6. PCA score results of rat serum samples (A), after radiation (600 lx, Class 1; 800 lx, Class 2), Control (Class 3) at the third day; the corresponding load diagram (B).

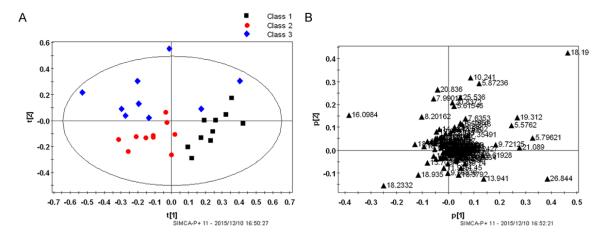


Figure 7. PLS-DA score results of rat serum samples (A), after radiation (600 lx, Class 1; 800 lx, Class 2), Control (Class 3) at the third day; the corresponding load diagram (B).

Ix, 800 Ix) were distinguished from the rats in the control group, the results of PLS-DA were better than PCA.

Changes in metabolite

Metabolomics comprises the measurement of endogenous metabolites, including amino acids, nucleic acid precursors, lipids, and degradation products of chemical intermediates in catabolism and biosynthesis. The advantage of metabolomics is that it provides the most functional measure of cellular status and can help to describe an organism's phenotype [26-28].

In this study, the changes of metabolites between radiation groups and their control group were shown in **Tables 1-3**. Compared to the control group, the level of propanoic acid of the 600 lx radiation group increased; the level of d-Glucose of the 800 lx radiation group decreased at the first day, **Table 1**. Compared to the control group, the level of propanoic acid and ethanedioic acid of the 600 lx radiation group increased at the second day, **Table 2**. Compared to the control group, the level of propanoic acid of the 600 lx radiation group increased; the level of d-Glucose of the 800 lx radiation group decreased at the first day, **Table 3**. These finding may be useful for new evidences in radiation study. Additional prospective studies will be required to better understand these observations.

Conclusion

These biomarkers (propanoic acid, d-Glucose, ethanedioic acid) could be useful for further

mes	in rat ser	um after radiation at the	inst day		
NO.	Renten time/min	Metabolite	VIP	Group	
				600 lx	800 lx
1	18.1941	d-Glucose	4.06616	-	\downarrow^*
2	18.409	L-Tyrosine	3.11377	-	-
3	5.87236	Propanoic acid	2.86904	1*	-
4	7.6353	Ethanedioic acid	2.66764	-	-
5	9.71105	Urea	2.50205	-	-
6	26.844	Benzoic acid	2.38154	-	-
7	8.20162	Glycine	2.06363	-	-
8	20.8372	trans-9-Octadecenoic acid	2.01641	-	-

Table 1. Summary of the changes in relative levels of metabolites in rat serum after radiation at the first day

Note: Variable importance in the projection (VIP) was acquired from the PLS-DA model with a threshold of 2.0. Marks indicate the direction of the change, i.e. ↓ for decrease, ↑ for increase, - for no change. Compared control group with radiation group (600, 800 lx), *P<0.05, as indicated by the statistical analysis T-test.

Table 2. Summary of the changes in relative levels of metabolites in rat serum after radiation at the second day

NO.	Renten	Matabalita	VIP	Group	
	time/min	Metabolite	VIP	600 lx	800 lx
1	5.87236	Propanoic acid	6.28194	-	1 **
2	16.0984	L-Cysteine	3.17248	-	-
3	7.6353	Ethanedioic acid	3.09429	-	1 **
4	25.536	Glycine	2.54632	-	-
5	7.99017	Butanoic acid	2.54243	-	-
6	26.844	Benzoic acid	2.21736	-	-

Note: Variable importance in the projection (VIP) was acquired from the PLS-DA model with a threshold of 2.0. Marks indicate the direction of the change, i.e. \downarrow for decrease, \uparrow for increase, - for no change. Compared control group with radiation group (600, 800 lx), **P<0.01, as indicated by the statistical analysis T-test.

Table 3. Summary of the changes in relative levels of metabolites in rat serum after radiation at the third day

NO.	Renten time/min	Metabolite	VIP	Dose group	
				Low	High
1	26.844	Benzoic acid	4.01279	-	-
2	18.1941	d-Glucose	3.80095	-	-
3	16.0984	L-Cysteine	3.54475	-	-
4	5.87236	Propanoic acid	3.20581	-	-
5	20.8372	trans-9-Octadecenoic acid	2.92908	-	\downarrow^*
6	19.312	Hexadecanoic acid	2.55216	-	-
7	21.089	Octadecanoic acid	2.45395	-	-
8	25.536	Glycine	2.44574	-	-
9	7.99017	Butanoic acid	2.16151	↓*	\downarrow^*

Note: Variable importance in the projection (VIP) was acquired from the PLS-DA model with a threshold of 2.0. Marks indicate the direction of the change, i.e. \downarrow for decrease, \uparrow for increase, - for no change. Compared control group with radiation group (600, 800 lx), *P<0.05, as indicated by the statistical analysis T-test.

radiation. We demonstrated that metabolomic methods based on GC/MS could provide a useful tool for exploring biomarkers in radiation study.

Acknowledgements

This study was supported by grants from the incubator project of the First Affiliated Hospital of Wenzhou Medical University, No. FHY2015013; the Youth Talent Program Foundation of the First Affiliated Hospital of Wenzhou Medical University, No. qnyc043.

Disclosure of conflict of interest

None.

Address correspondence to: Drs. Lufeng Hu and Qing Wu, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou 325000, Zhejiang, P. R. China. Tel: (86) 577555-79706; E-mail: hulufeng@163.com (LFH); wuqing830@163.com (QW)

References

- [1] Spatola C, Migliore M, Emanuele Liardo RL, Bevilacqua R, Luigi R, Vincenzo S, Tocco A, Pagana A, Militello C, Calvo D, Criscione A and Privitera G. Follicular dendritic cell sarcoma of mediastinum: a key role of radiotherapy in a multidisciplinary approach. Future Oncol 2015; 11: 57-61.
- [2] Song JH, Jeong BK, Choi HS, Jeong H, Kang MH, Kang JH, Kim JP, Park JJ, Woo SH, Jang HS, Choi BO and Kang KM. Comparison of Failure Patterns Between Conventional and Intensity-modulated Radiotherapy for Stage III and IV Head and Neck Squamous Cell Carcinoma. Anticancer Res 2015; 35: 6833-6840.
- [3] Alberts L, El Sharouni SY, Hofman FN, BP VANP, Tromp E, M VANV, Kastelijn EA and Schramel FM. Changes in Pulmonary Function After Stereotactic Body Radiotherapy and After Surgery for Stage I and II

Non-small Cell Lung Cancer, a Description of Two Cohorts. Anticancer Res 2015; 35: 6773-6779.

- [4] Birgani MT, Fatahiasl J, Hosseini SM, Bagheri A, Behrooz MA, Zabiehzadeh M, Meskani R and Gomari MT. Breast Radiotherapy with Mixed Energy Photons; a Model for Optimal Beam Weighting. Asian Pac J Cancer Prev 2015; 16: 7785-7788.
- [5] Yang YF, Cao XH, Bao CE and Wan X. Concurrent radiotherapy with oral fluoropyrimidine versus gemcitabine in locally advanced pancreatic cancer: a systematic review and meta-analysis. Onco Targets Ther 2015; 8: 3315-3322.
- [6] Zarva A, Mohnike K, Damm R, Ruf J, Seidensticker R, Ulrich G, Seidensticker M, Pech M, Ricke J and Amthauer H. Safety of repeated radioembolizations in patients with advanced primary and secondary liver tumors and progressive disease after first selective internal radiotherapy. J Nucl Med 2014; 55: 360-366.
- [7] Park HJ, Kim SH, Jang KM, Lim S, Kang TW, Park HC and Choi D. Added value of diffusionweighted MRI for evaluating viable tumor of hepatocellular carcinomas treated with radiotherapy in patients with chronic liver disease. AJR Am J Roentgenol 2014; 202: 92-101.
- [8] Kim M, Son SH, Won YK and Kay CS. Stereotactic ablative radiotherapy for oligometastatic disease in liver. Biomed Res Int 2014; 2014: 340478.
- [9] Cheema AK, Pathak R, Zandkarimi F, Kaur P, Alkhalil L, Singh R, Zhong X, Ghosh S, Aykin-Burns N and Hauer-Jensen M. Liver metabolomics reveals increased oxidative stress and fibrogenic potential in gfrp transgenic mice in response to ionizing radiation. J Proteome Res 2014; 13: 3065-3074.
- [10] Cheema AK, Suman S, Kaur P, Singh R, Fornace AJ Jr and Datta K. Long-term differential changes in mouse intestinal metabolomics after gamma and heavy ion radiation exposure. PLoS One 2014; 9: e87079.
- [11] Manna SK, Krausz KW, Bonzo JA, Idle JR and Gonzalez FJ. Metabolomics reveals aging-associated attenuation of noninvasive radiation biomarkers in mice: potential role of polyamine catabolism and incoherent DNA damage-repair. J Proteome Res 2013; 12: 2269-2281.
- [12] Liu H, Wang Z, Zhang X, Qiao Y, Wu S, Dong F and Chen Y. Selection of candidate radiation biomarkers in the serum of rats exposed to gamma-rays by GC/TOFMS-based metabolomics. Radiat Prot Dosimetry 2013; 154: 9-17.
- [13] Johnson CH, Patterson AD, Krausz KW, Kalinich JF, Tyburski JB, Kang DW, Luecke H, Gonzalez FJ, Blakely WF and Idle JR. Radiation metabolomics. 5. Identification of urinary biomarkers of

ionizing radiation exposure in nonhuman primates by mass spectrometry-based metabolomics. Radiat Res 2012; 178: 328-340.

- [14] Johnson CH, Patterson AD, Krausz KW, Lanz C, Kang DW, Luecke H, Gonzalez FJ and Idle JR. Radiation metabolomics. 4. UPLC-ESI-QTOFMS-Based metabolomics for urinary biomarker discovery in gamma-irradiated rats. Radiat Res 2011; 175: 473-484.
- [15] Coy SL, Cheema AK, Tyburski JB, Laiakis EC, Collins SP and Fornace A Jr. Radiation metabolomics and its potential in biodosimetry. Int J Radiat Biol 2011; 87: 802-823.
- [16] Wang X, Zhang M, Ma J, Zhang Y, Hong G, Sun F, Lin G and Hu L. Metabolic changes in paraquat poisoned patients and support vector machine model of discrimination. Biol Pharm Bull 2015; 38: 470-475.
- [17] Zhang M, Deng M, Ma J and Wang X. An evaluation of acute hydrogen sulfide poisoning in rats through serum metabolomics based on gas chromatography-mass spectrometry. Chem Pharm Bull (Tokyo) 2014; 62: 505-507.
- [18] Patti GJ, Yanes O and Siuzdak G. Innovation: Metabolomics: the apogee of the omics trilogy. Nat Rev Mol Cell Biol 2012; 13: 263-269.
- [19] Deng M, Zhang M, Huang X, Ma J, Hu L, Lin G and Wang X. A gas chromatography-mass spectrometry based study on serum metabolomics in rats chronically poisoned with hydrogen sulfide. J Forensic Leg Med 2015; 32: 59-63.
- [20] Deng M, Zhang M, Sun F, Ma J, Hu L, Yang X, Lin G and Wang X. A gas chromatography-mass spectrometry based study on urine metabolomics in rats chronically poisoned with hydrogen sulfide. Biomed Res Int 2015; 2015: 295241.
- [21] Wen C, Zhang M, Ma J, Hu L, Wang X and Lin G. Urine metabolomics in rats after administration of ketamine. Drug Des Devel Ther 2015; 9: 717-722.
- [22] Wen C, Zhang M, Zhang Y, Sun F, Ma J, Hu L, Lin G and Wang X. Brain metabolomics in rats after administration of ketamine. Biomed Chromatogr 2016; 30: 81-4.
- [23] Wang Z, Ma J, Zhang M, Wen C, Huang X, Sun F, Wang S, Hu L, Lin G and Wang X. Serum Metabolomics in Rats after Acute Paraquat Poisoning. Biol Pharm Bull 2015; 38: 1049-1053.
- [24] Monte AA, Heard KJ and Vasiliou V. Prediction of drug response and safety in clinical practice. J Med Toxicol 2012; 8: 43-51.
- [25] Mamas M, Dunn WB, Neyses L and Goodacre R. The role of metabolites and metabolomics in clinically applicable biomarkers of disease. Arch Toxicol 2011; 85: 5-17.

- [26] Hong JH, Lee WC, Hsu YM, Liang HJ, Wan CH, Chien CL and Lin CY. Characterization of the biochemical effects of naphthalene on the mouse respiratory system using NMR-based metabolomics. J Appl Toxicol 2014; 34: 1379-1388.
- [27] Zhang M, Wen C, Zhang Y, Sun F, Wang S, Ma J, Lin K, Wang X, Lin G and Hu L. Serum metabolomics in rats models of ketamine abuse by gas chromatography-mass spectrometry. J Chromatogr B Analyt Technol Biomed Life Sci 2015; 1006: 99-103.
- [28] Zhang Q, Wu H, Wen C, Sun F, Yang X and Hu L. Metabolic changes in rats after intragastric administration of MGCD0103 (Mocetinostat), a HDAC class I inhibitor. Int J Clin Exp Pathol 2015; 8: 9320-9325.