Original Article

High level of STAT4 expression is associated with the deterioration of breast cancer

Rongquan He1*, Hao Chen2*, Zhenbo Feng2, Yiwu Dang2, Tingqing Gan1, Gang Chen2, Lihua Yang1

Departments of ¹Medical Oncology, ²Pathology, The First Affiliated Hospital of Guangxi Medical University, Nanning, P. R. China. *Equal contributors.

Received December 31, 2015; Accepted March 22, 2016; Epub June 15, 2016; Published June 30, 2016

Abstract: *Purpose*: Dysregulation of signal transducers and activators of transcription 4 (STAT4) has been reported in several classes of malignancies. However, its expression and clinicopathological contribution in breast cancer remains unclarified. The purpose of the current study was to explore the clinicopathological significance of STAT4 and its role on the deterioration in breast cancer. *Methods*: A total of 81 samples from breast cancer patients and 40 cases of adjacent non-cancerous breast tissues were recruited, and the expression of STAT4 was detected with immunohistochemistry. Additionally, the association between STAT4 and clinical parameters was analyzed. *Results*: The expression level of STAT4 in breast cancer was significantly higher than that in non-cancerous breast tissues (*P*<0.001). Moreover, a notable correlation was found between STAT4 expression and the tumor size (r=0.504, *P*<0.001). There were also positive associations of STAT4 with lymph nodes with cancer spreading (r=0.419, *P*<0.001), and with clinical TNM stage of breast cancer (r=0.541, *P*<0.001). *Conclusions*: Up-regulation of STAT4 in breast cancer could indicate the deterioration of breast cancer. STAT4 should be taken into consideration in the development of novel diagnostic and therapeutic program for breast cancer.

Keywords: STAT4, breast cancer, immunohistochemistry, deterioration, TNM

Introduction

Breast cancer is one of the most common diseases in women. Incidence and mortality due to breast cancer has been growing for last 50 years [1, 2]. There were estimated 227,000 new cases of breast cancer diagnosed in women in the United States and estimated 39,500 breast cancer deaths in 2012 [3, 4]. As World Health Organization (WHO) 2012 reported, breast cancer is the prominent cause of death in women, accounting 23% of all cancer deaths. In Asian countries, one in every three females faces the possibility of breast cancer in their lifetime [5, 6]. Breast cancer is a heterogeneous disease, not only in its features and clinical course, but also in its molecular profile [7]. Current approaches for assessing disease prognosis, using clinicopathological elements such as age, tumor size, tumor grade, and extent of nodal involvement in their evaluation, are inadequate in value for estimating the risk of the prognosis of breast cancer. Thus, there is a disquieting requirement for the identification of best diagnosing and predicting marker for breast cancer.

The Janus Kinase-Signal Transducers and Activators of Transcription (JAK-STAT) pathways have been reported to play essential roles in the immune, neuronal, hematopoietic and hepatic systems. STAT family members can be divided into two groups according to their specific functions. One is made up of STAT1, STAT3, and STAT5, activated in diverse tissues by means of a sequence of ligands and involved in IFN signaling, development of the mammary gland, response to growth hormone, and embryogenesis. The other group includes STAT2, STAT4, and STAT6, which are activated by a few cytokines and play a distinctive role in the development of T-cells and in IFN-gamma signaling [8]. Among the STAT family members, STAT4 is a vital transcription factor that is critical for the differentiation of Th1 cells in promoting cellular immune reaction [9]. Furthermore, the role of STAT4 has also been investigated in a small number of malignancies, that is, abnor-

Table 1. Relationship between STAT4 and Ki-67 expression and clinicopathological features

| Clinicopathological features | | N (total) | n (STAT4 positive, %) | Z | Р |
|------------------------------|-------------------|-----------|-----------------------|--------|---------|
| Tissue | Non-cancerous | 40 | 9 (22.5%) | 4.426 | <0.001 |
| | Breast cancer | 81 | 53 (65.4%) | | |
| Age | <50 | 38 | 26 (68.42%) | -0.528 | 0.597 |
| | ≥50 | 43 | 27 (62.79%) | | |
| Histological grade* | Carcinoma in situ | 2 | 0 (0.00%) | 4.168 | 0.244 |
| | 1 | 5 | 3 (60.00%) | | |
| | II | 55 | 38 (69.09%) | | |
| | III | 19 | 12 (63.16%) | | |
| Tumor size* | Tis | 2 | 0 (0.00%) | 21.417 | < 0.001 |
| | T1 | 15 | 4 (26.67%) | | |
| | T2 | 44 | 30 (68.18%) | | |
| | T3 | 16 | 15 (93.75%) | | |
| | T4 | 4 | 4 (100%) | | |
| Lymph node metastasis* | NO | 38 | 17 (44.74%) | 14.604 | 0.002 |
| | N1 | 18 | 14 (77.78%) | | |
| | N2 | 17 | 14 (82.35%) | | |
| | N3 | 8 | 8 (100%) | | |
| TNM* | 0 | 2 | 0 (0.00%) | 26.197 | 0.000 |
| | 1 | 10 | 1 (10.00%) | | |
| | II | 37 | 23 (62.16%) | | |
| | III | 32 | 29 (90.63%) | | |
| Molecular subgroup* | Luminal A | 23 | 15 (65.22%) | 0.805 | 0.669 |
| | Her2 | 36 | 22 (61.11%) | | |
| | TN | 22 | 16 (72.73%) | | |
| ER and PR | Negative | 58 | 38 (65.52%) | -0.025 | 0.980 |
| | Positive | 23 | 8 (34.78%) | | |
| HER2 | Negative | 45 | 31 (68.89%) | -0.727 | 0.467 |
| | Positive | 36 | 22 (61.11%) | | |
| Ki-67 grade | Low | 5 | 4 (80.00%) | 0.500 | 0.479 |
| | High | 76 | 49 (64.47%) | | |
| P53 grade | Low | 42 | 30 (71.43%) | 1.387 | 0.239 |
| | High | 39 | 23 (58.97%) | | |
| P16 grade | Low | 28 | 15 (53.57%) | 1.014 | 0.314 |
| | High | 53 | 38 (71.70%) | | |
| E-cadherin | Low | 24 | 18 (75.00%) | 1.380 | 0.240 |
| | High | 57 | 35 (61.40%) | | |
| Vimentin | Negative | 56 | 41 (73.21%) | 4.858 | 0.028 |
| | Positive | 25 | 12 (48.00%) | | |

^{*}Kruskal-Wallis H test was performed. The following pairwise comparisons were performed with chi-square test: Tumor size: T1 vs T2: Z=-2.786, P=0.005; T1 vs T3: Z=-3.77, P<0.001; T1 vs T4: Z=-2.569, P=0.010; T2 vs T3: Z=-2.006, P=0.045. Lymph node metastasis: N0 vs N1: Z=-2.302, P=0.021; N0 vs N2: Z=-2.576, P=0.010; N0 vs N3: Z=-2.821, P=0.005. TNM stages: stage I vs stage II: Z=-2.896, P=0.004; stage I vs stage III: Z=-2.716, Z=-0.007.

mal expression of STAT4 was found in different neoplasia, including colorectal cancer [10, 11], gastric cancer [12], cutaneous T-cell lymphoma [13-15] and hepatocellular carcinoma (HCC) [9, 16, 17]. However, there is a lacuna in the possible function of STAT4 in breast cancer. Thus, in the current study, we detected the STAT4 protein expression in the clinical samples of 81 cases of breast cancer and investigated its clinical significance.

Methods

Tissue samples

A total of 81 cases of FFPE breast cancer and 40 of their corresponding non-cancerous adjacent breast tissues were enrolled in the current study. The age of the breast cancer patients ranged from 30 to 76 years, with a mean age of 50.69 years. All samples were from female patients. Clinicopathological information was provided from medical records and the main parameters were listed in Table 1. All cases were initial tumorectomies without treatment and randomly selected in the First Affiliated Hospital of Guangxi Medical University, China, between January 2012 and December 2013. The study protocol was approved by the Ethical Committee of the First Affiliated Hospital of Guangxi Medical University. Written informed consent was obtained from the patients and clinicians for the usage of the samples for research. All samples were reviewed and diagnosed by two independent pathologists.

Immunohistochemistry

The fixed tissue samples were embedded in paraffin. Sections were deparaffinized in xylene and hydrated through a graded series of ethanol and then rehydrated and subjected to antigen retrieval by microwaving. Endogenous peroxidase activity was quenched with 3% hydrogen peroxide for 10 min at room temperature. Antigen retrieval of the sections was achieved in a multifunctional microwave histoprocessor at 100°C by microwave heating of the samples on slides in 0.01 mol/L of pH 6.0 citrate buffer for 20 min. Biomarker expression was immunohistochemically detected by primary antibodies of mouse monoclonal antibody STAT4 (PL68, sc-101160, Santa Cruz Biotec.CO., CA, USA, 1:300 dilution), rabbit monoclonal antibody Ki-67, P53, P16, E-cadherin, Vimentin (Beijing Zhongshan Jinqiao Biotec.CO., China) for 60 min at room temperature followed by 30 min staining with biotinylated secondary anti-mouse/rabbit antibody (Cat. No. D-3004, Shanghai Long Island Biotec. CO., LTD, China). Finally, positive staining was visualized with diaminobenzidine and cell nuclei were counterstained with haematoxylin. The positive signals of STAT4, E-cadherin and Vimentin locate in the cytoplasm. Negative (-), weakly positive (+), moderately positive (++),

and strongly positive (+++) were determined according to the immunodetection of stain intensity and amounts of positive cells by two pathologists (HC and GC), who discussed each case until they reached a consensus. All of (+), (++), and (+++) were considered as positive expression [18]. The positive signal of Ki-67, P53 and P16 is distributed in the nuclei. The proliferation index (PI) of Ki-67 was calculated with the formula (number of positive cells/total number of the cells ×100%) by counting at least 10 representative visions of high magnification (40×40). P53 and P16 were scored with the same strategy as Ki-67.

Statistical analysis

SPSS20.0 was used for statistical analysis. Chi-square test and Kruskal-Wallis H test were performed to analyze the difference of STAT4 expression between distinct clinicopathological groups. Bivariate correlations between two independent variables were analyzed by calculating the Spearman's correlation coefficients. Moreover, ROC curve was performed to analyze the predictive value of STAT4 to distinguish cancer from non-cancer. Statistical significance was determined at a *P*<0.05 level.

Results

STAT4 protein was found to be expressed in 53 out of 81 cases breast cancer tissues (65.4%) and the positive rate was significantly higher than that in the non-cancerous breast tissue (22.5%, 9/40, P<0.001, Table 1; Figure 1). In addition, ROC curve was performed to explore the diagnostic value of STAT4 for breast cancer. The area under curve (AUC) of STAT4 was 0.715 (95% CI: 0.618-0.812, P<0.001). In terms of the tumor size of breast cancer, the STAT4 expression was 0%, 26.67%, 68.18%, 93.75%, and 100%, for Tis, T1, T2, T3 and T4, respectively. Spearman's correlation test showed a positive association between STAT4 expression and tumor size (r=0.504, P<0.001, Figure 2). Similar trend was also observed between STAT4 expression and lymph node metastasis. The positive rate of STAT4 expression was 44.74%, 77.78%, 82.35%, 100%, with the increasing numbers of lymph nodes with cancer spreading (r=0.419, P<0.001, Figure 2). Furthermore, STAT4 expression was closely related to the clinical TNM stage of breast cancer (r=0.541, P<0.001, Figure 2). With the deterioration of

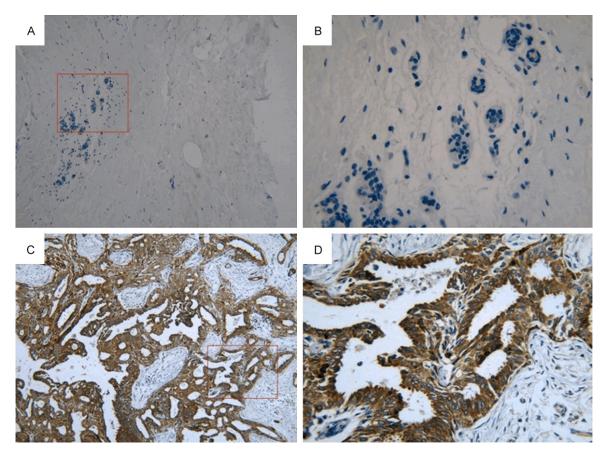


Figure 1. STAT4 expression in breast cancer and non-cancerous breast tissues. Negative STAT4 expression in non-cancerous breast tissue (A: \times 100; B: \times 400, visualized by the red boxed area from A); Positive STAT4 expression in breast cancer tissue (C: \times 100; D: \times 400, enlarged from the area within the red box in C, immunohistochemistry).

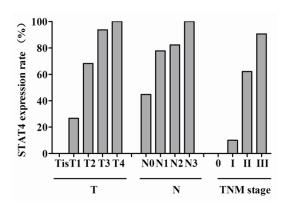


Figure 2. Different STAT4 expression rate in diverse subtypes of breast cancer. T: the size of tumor, N: lymph node with cancer spreading.

breast cancer, the positive rate of STAT4 expression rose with 0%, 10%, 62.16% and 90.63% for stage 0, stage I, stage II and stage III, respectively. However, no relative relationship was observed between STAT4 expression and other parameters, such as age, histology, mo-

lecular subtype, the status of ER, PR or HER2 (all *P*>0.05, **Table 1**).

To further understand the potential mechanism of STAT4 in breast cancer, we investigated the relationship between the expression of STAT4 and some other biomarkers, which are representative of the status of cell proliferation, invasion and metastasis. The STAT4 expression rate in the Vimentin positive group was 48% (12/25), markedly lower than that in the Vimentin negative group (73.21%, 41/56, P=0.028). STAT4 expression was negatively related to Vimentin expression (r=-0.286, r=0.01). No significant relationship was found between STAT4 expression and other markers, such as Ki-67, P53, P16 or E-cadherin (**Table 1**).

Discussion

Breast cancer is a complex and heterogeneous disease. Due to the heterogeneity, the mecha-

nism of breast tumorigenesis remains still not completely understood [19, 20]. STAT4, locating on chromosome 2q32, is a member of the STAT superfamily of transcription factors. Dysregulation of STAT4 has been reported in several cancers. In colorectal cancer [10, 11] and gastric cancer [12], over-expression of STAT4 protein was detected. In contrast, underexpression of STAT4 protein was found in cutaneous T-cell lymphoma [13-15] and HCC [9, 16, 17]. In the present study, we found STAT4 protein expression in 22.5% samples of non-cancerous breast tissues using immunohistochemistry, which is consistent with the previous report [21], showing that STAT4 was expressed in mammary tissue. However, we primarily discovered that significantly higher STAT4 expression was noted in breast cancer tissues (65.4%) than that in non-cancerous breast, which is similar as the fact that the high expression of STAT4 was observed in colorectal cancer [10, 11] and gastric cancer [12]. Furthermore, the potential mechanism of STAT4 has been studied in these two classes of cancers. Cheng et al. [10] reported that silencing of STAT4 gene could suppress cell proliferation and invasion of colorectal cancer cells. Zhou et al [12] found that STAT4 could function as a target gene of miR-141 in gastric cancer, which was downregulated. Over-expression of STAT4 in vitro could mimic miR-141 action in the growth and invasion of gastric cancer. To date, no available study has been performed to explore the mechanism of STAT4 on breast cancer. Future work is urgently needed to figure out the role of STAT4 in breast cancer. However, the immunostaining result in current study indicates that STAT4 might play an essential role in the carcinogenesis of breast cancer and could also be a potential tool for the diagnosis of breast cancer.

We are also interested in the relationship between STAT4 protein and the progression of breast cancer. In the current study, we found that the positive ratio of STAT4 expression was rising with the deterioration of breast cancer. Higher STAT4 expression was observed in the groups of larger tumor, more lymph nodes with cancer spreading and more advanced clinical stage, which suggests that STAT4 might play a vital role in the development of breast cancer. Cheng et al [10] reported an analogous phenomenon in colorectal cancer and STAT4 expression was related with the Duke's staging

and depth of invasion in colorectal cancer patients. However, the molecular mechanism involved has not yet been clarified.

Signaling by estrogen receptor (ER), progesterone receptor (PR), and/or human EGF-like receptor 2 (HER2) is a key driver in the progress of a great majority of breast cancers [22-24]. Molecular characterization of primary cancers has identified major subtypes that relate with ER/PR/HER2 status, and also subgroup divisions that indicate other molecular and cellular characteristics of the tumors [25, 26]. However, several challenges remain to improve breast cancer controlling and patient survival, for which the integration of novel markers into current practice should be advantageous [27, 28]. In the current study, we have compared the STAT4 expression to the status of ER, PR, HER2 in breast cancer. However, no significant correlation has been found between STAT4 and ER/ PR/HER2 status so far. The relationship of STAT4 with ER/PR/HER2 status needs further investigation with larger patient size and functional experiments.

Finally, we compared STAT4 expression to some biomarkers representing the cell proliferation, migration and epithelial-mesenchymal transition (EMT) in breast cancer [29-33]. No significant relationship was found between STAT4 and Ki-67, P53, P16 or E-cadherin. However, reverse correlation was observed between STAT4 and Vimentin, which is important in the transformation of a normal cell to an invasive tumor cell [34]. Vimentin is an intermediate filament expressed in tissues of normal mesenchymal origin. It is known to express aberrantly in epithelial cancers of prostate, gastrointestinal tract, breast, central nervous system, lung, and malignant melanomas [35]. In recent years. Vimentin has been recognized as a marker for EMT. However, more researches would be crucial to evaluate the link between STAT4 and EMT, between STAT4 and Vimentin in breast cancer.

In conclusion, the current finding reveals that high-level STAT4 expression might be a novel molecular alteration involved in the carcinogenesis and progression of breast cancer. STAT4 might serve as a valuable biomarker to monitor the clinical course of patients with breast cancer. Chemotherapy-induced STAT4 deficiency was reported to be due to the reduced levels of

STAT4 mRNA and the protein stability [14]. Hence, STAT4 should be taken into consideration in the development of novel diagnostic and molecular therapeutic program for breast cancer, although more consolidated studies are still required.

Acknowledgements

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Disclosure of conflict of interest

None.

Address correspondence to: Drs. Gang Chen and Lihua Yang, Departments of Pathology, The First Affiliated Hospital of Guangxi Medical University, No. 6 Shuangyong Road, Nanning 530021, Guangxi Zhuang Autonomous Region, P. R. China. Tel: 00-867715356534; Fax: 00867715356534; E-mail: chen_gang_triones@163.com (GC); Tel: 0086771-3277289; Fax: 00867713277289; E-mail: 15087-1746@qq.com (LHY)

References

- [1] Sarmento de Almeida G, Leal Almeida LA, Rodrigues Araujo GM, Weller M. Reproductive risk factors differ among breast cancer patients and controls in a public hospital of paraiba, northeast Brazil. Asian Pac J Cancer Prev 2015; 16: 2959-2965.
- [2] Kim Y, Yoo KY, Goodman MT. Differences in incidence, mortality and survival of breast cancer by regions and countries in Asia and contributing factors. Asian Pac J Cancer Prev 2015; 16: 2857-2870.
- [3] Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. CA Cancer J Clin 2015; 65: 5-29.
- [4] Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. CA Cancer J Clin 2015; 65: 87-108.
- [5] Donepudi MS, Kondapalli K, Amos SJ, Venkanteshan P. Breast cancer statistics and markers. J Cancer Res Ther 2014; 10: 506-511.
- [6] Miao H, Hartman M, Bhoo-Pathy N, Lee SC, Taib NA, Tan EY, Chan P, Moons KG, Wong HS, Goh J, Rahim SM, Yip CH, Verkooijen HM. Predicting survival of de novo metastatic breast cancer in Asian women: systematic review and validation study. PLoS One 2014; 9: e93755.
- [7] Maass N, Schütz F, Fasching PA, Fehm T, Janni W, Kümmel S, Kolberg HC, Lüftner D,

- Wallwiener M, Lux MP. Breast Cancer Update 2014 Focus on the Patient and the Tumour. Geburtshilfe Frauenheilkd 2015; 75: 170-182.
- [8] Calò V, Migliavacca M, Bazan V, Macaluso M, Buscemi M, Gebbia N, Russo A. STAT proteins: from normal control of cellular events to tumorigenesis. J Cell Physiol 2003; 197: 157-168.
- [9] Wang Y, Qu A, Wang H. Signal Transducer and Activator of Transcription 4 in Liver Diseases. Int J Biol Sci 2015; 11: 448-455.
- [10] Cheng JM, Yao MR, Zhu Q, Wu XY, Zhou J, Tan WL, Zhan SH. Silencing of stat4 gene inhibits cell proliferation and invasion of colorectal cancer cells. J Biol Regul Homeost Agents 2015; 29: 85-92.
- [11] Slattery ML, Lundgreen A, Kadlubar SA, Bondurant KL, Wolff RK. JAK/STAT/SOCSsignaling pathway and colon and rectal cancer. Mol Carcinogenes 2013; 52: 155-166.
- [12] Zhou X, Xia Y, Su J, Zhang G. Down-regulation of miR-141 induced by helicobacter pylori promotes the invasion of gastric cancer by targeting STAT4. Cell Physiol Biochem 2014; 33: 1003-1012.
- [13] Litvinov IV, Cordeiro B, Fredholm S, Odum N, Zargham H, Huang Y, Zhou Y, Pehr K, Kupper TS, Woetmann A, Sasseville D. Analysis of STAT4 expression in cutaneous T-cell lymphoma (CTCL) patients and patient-derived cell lines. Cell Cycle 2014; 13: 2975-2982.
- [14] Lupov IP, Voiles L, Han L, Schwartz A, De La Rosa M, Oza K, Pelloso D, Sahu RP, Travers JB, Robertson MJ, Chang HC. Acquired STAT4 deficiency as a consequence of cancer chemotherapy. Blood 2011; 118: 6097-6106.
- [15] Netchiporouk E, Litvinov IV, Moreau L, Gilbert M, Sasseville D, Duvic M. Deregulation in STAT signaling is important for cutaneous T-cell lymphoma (CTCL) pathogenesis and cancer progression. Cell Cycle 2014; 13: 3331-3335.
- [16] Wang G, Chen JH, Qiang Y, Wang DZ, Chen Z. Decreased STAT4 indicates poor prognosis and enhanced cell proliferation in hepatocellular carcinoma. World J Gastroenterol 2015; 21: 3983-3993.
- [17] Wubetu GY, Utsunomiya T, Ishikawa D, Yamada S, Ikemoto T, Morine Y, Iwahashi S, Saito Y, Arakawa Y, Imura S, Kanamoto M, Zhu C, Bando Y, Shimada M. High STAT4 expression is a better prognostic indicator in patients with hepatocellular carcinoma after hepatectomy. Ann Surg Oncol 2014; 21 Suppl 4: S721-728.
- [18] Huang S, Chen G, Dang Y, Chen LH. Overexpression of DcR3 and its significance on tumor cell differentiation and proliferation in glioma. ScientificWorldJournal 2014; 2014: 605236.

- [19] Li YT, Ni D, Yang L, Zhao Q, Ou JH. The prevalence of BRCA1/2 mutations of triple-negative breast cancer patients in Xinjiang multiple ethnic region of China. Eur J Med Res 2014; 19: 35
- [20] Tseng HS, Chen LS, Kuo SJ, Chen ST, Wang YF, Chen DR. Tumor characteristics of breast cancer in predicting axillary lymph node metastasis. Med Sci Monit 2014; 20: 1155-1161.
- [21] Watson CJ. Stat transcription factors in mammary gland development and tumorigenesis. J Mammary Gland Biol Neoplasia 2001; 6: 115-127.
- [22] Wang WJ, Lei YY, Mei JH, Wang CL. Recent Progress in HER2 Associated Breast Cancer. Asian Pac J Cancer Prev 2015; 16: 2591-2600
- [23] Ning SF, Li JL, Luo CP, Wei CH, Lu YK, Liu HZ, Wei WE, Zhang LT. Human epidermal growth factor receptor 2 expression in breast cancer: correlation with clinical pathological features. Int J Clin Exp Pathol 2014; 7: 8740-8747.
- [24] Zhang F, Kang H, Xu Q. Estrogen increases secretion of stromal cell derived factor-1 in human breast cancer cells. Int J Clin Exp Med 2014; 7: 5529-5534.
- [25] Verma S. Advances in treating HER2-positive breast cancer: an interview with Sunil Verma. BMC Med 2014: 12: 129.
- [26] Lavaud P, Andre F. Strategies to overcome trastuzumab resistance in HER2-overexpressing breast cancers: focus on new data from clinical trials. BMC Med 2014; 12: 132.
- [27] Lee SB, Lee JW, Yu JH, Ko BS, Kim HJ, Son BH, Gong G, Lee HJ, Kim SB, Jung KH, Ahn JH, Lee W, Sung J, Ahn SH. Preoperative serum HER2 extracellular domain levels in primary invasive breast cancer. BMC Cancer 2014; 14: 929.

- [28] Surazynski A, Miltyk W, Wolczynski S, Palka J. The effect of prolactin and estrogen cross-talk on prolidase- dependent signaling in MCF-7 cells. Neoplasma 2013; 60: 355-363.
- [29] Repetto O, De Paoli P, De Re V, Canzonieri V, Cannizzaro R. Levels of soluble E-cadherin in breast, gastric, and colorectal cancers. BioMed Res Int 2014; 2014: 408047.
- [30] Milicevic Z, Bajic V, Zivkovic L, Kasapovic J, Andjelkovic U, Spremo-Potparevic B. Identification of p53 and its isoforms in human breast carcinoma cells. ScientificWorldJournal 2014; 2014: 618698.
- [31] Li FY, Wu SG, Zhou J, Sun JY, Lin Q, Lin HX, Guan XX, He ZY. Prognostic value of Ki-67 in breast cancer patients with positive axillary lymph nodes: a retrospective cohort study. PLoS One 2014; 9: e87264.
- [32] Zhou Y, Ming J, Xu Y, Zhang Y, Jiang J. ERbeta1 inhibits the migration and invasion of breast cancer cells through upregulation of E-cadherin in a Id1-dependent manner. Biochem Biophys Res Commun 2015: 457: 141-147.
- [33] Liang Z, Sun XY, Xu LC, Fu RZ. Abnormal expression of serum soluble E-cadherin is correlated with clinicopathological features and prognosis of breast cancer. Med Sci Monit 2014; 20: 2776-2782.
- [34] Calaf GM, Balajee AS, Montalvo-Villagra MT, Leon M, Daniela NM, Alvarez RG, Roy D, Narayan G, Abarca-Quinones J. Vimentin and Notch as biomarkers for breast cancer progression. Oncol Lett 2014; 7: 721-727.
- [35] Hemalatha A, Suresh TN, Kumar ML. Expression of vimentin in breast carcinoma, its correlation with Ki67 and other histopathological parameters. Indian J Cancer 2013; 50: 189-194.