# Original Article Chlorogenic acid slows down proteinuria and renal fibrosis in 5/6-nephrectomized rats by anti-oxidation and inhibiting accumulation of extracellular matrix

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**Abstract:** Object: Chlorogenic acid (CA), a phenolic compound, was reported to have beneficial effect on cisplatin induced acute kidney injury. The present study aimed to introduce 5/6 nephrectomized rat model to further evaluate its renal protective effect. Methods: In this study, adult Wistar rats were induced to develop chronic renal failure through 5/6 nephrectomy (5/6 Nx). After that, animals were treated orally with saline, CA at 20 and 60 mg/kg daily for 20 weeks; sham-operated animals were also involved as control. After treatment for 10 and 20 weeks, blood and urine samples were collected for biochemical examination; all the kidney remnants were collected for histological examination. The protein levels of TGF- $\beta$ 1, smad2 and phosphorylated-smad2 (p-smad2) in kidney were measured. Immunohistochemistry was used to analyze the expression of TGF- $\beta$ 1, fibronectin and collagen IV in kidney tissues. Results: Results suggested that CA could reduce the proteinurea, blood urine nitrogen and blood creatinine in 5/6 Nx animals significantly, as well as oxidation stress in the kidney. By histological examination, CA administration alleviated glomerular sclerosis scores and tubulointerstitial injuries in a dose-dependent manner (P<0.01). Immunohistochemistry also suggested CA could reduce the expression of TGF- $\beta$ 1, fibronectin and collagen IV in kidney tissues of rats. Conclusions: This study suggests that the CA can improve renal function in 5/6 Nx rats effectively. Its effect may be due to its anti-oxidation and inhibiting accumulation of extracellular matrix.

Keywords: CA, 5/6 nephrectomy, TGF-β1, chronic renal failure

#### Introduction

Chronic kidney disease (CKD) is emerging as a major global health threat [1, 2], however, there is lack of long-term effective medicine against CKD currently; Searching for new potency compounds against CKD is still a major challenge.

Chlorogenic acid (CA) is a phenolic compound widely distributed in fruits and vegetables, including apples, pears, carrots, tomatoes and sweet potatoes, with the highest concentration in coffee and tea [3]. Previous studies demonstrated free radical scavenging and antioxidant activity of CA in vitro [4]. In biological studies, CA prevented chemically induced damage in liver and primary cortical neurons by reducing oxidative damage and apoptosis [5, 6]. The anti-inflammatory activity of CA has been related to the inhibition of nuclear factor-kappaB (NF- $\kappa$ B) activation and the release of pro-inflammatory cytokines in both cell cultures and mice liver [7, 8]. Previously we showed that CA was, at least in part, responsible for the hepatoprotective effects of dandelion (Taraxacum officinale L.) root extract in mice [9].

Besides hepatoprotective effects, several studies also demonstrated that CA has significant beneficial kidney protection effect in druginduced nephropathy [10] and diabetic nephropathy [11, 12]. Some studies demonstrate that CA has good anti-hypertension effect [13], and anti-hypertension is a valuable strategy to control progress of chronic kidney dysfunction [14].

To further study its renal protective effect on chronic kidney dysfunction besides diabetic nephropathy, the remnant kidney model of 5/6 nephrectomy (5/6 Nx) in rats was introduced in the current investigation. This model is often used to study the mechanisms of and potential therapeutic approaches to progression of chronic kidney disease (CKD) with renal mass reduction [15, 16]. In this model, systemic hypertension and proteinuria contribute to kidney injury and to the expression of pro-inflammatory and pro-fibrotic molecules by kidney cells. The nephrectomy is widely used as an animal model for glomerular hyperfiltration followed by glomerulosclerosis [17], it can mimic the advanced-stage nephropathy which is caused by various factors [18]. TGFβ-smad signal pathway was considered to play important roles in renal end-stage fibrosis in this model. In the current study, besides analysis of renal function and histopathological changes, we also measure the blood pressure and expression of TGF<sub>β1</sub>, smad2 and p-samd2 in the kidneys of 5/6 Nx rats, aimed to understand the underlying mechanisms of CA on renal beneficial effect.

### Materials and methods

### Chemicals

CA was purchased from sigma Aldrich (Germany), enzymic antioxidants (superoxide dismutase [SOD], catalase, glutathione peroxidase [GPx] and glutathione reductase [GRx]) and nonenzymic antioxidants (reduced glutathione [GSH] were measured by commercial kits (Nanjing Jiancheng Ltd, Nanjing, China).

### Animals

Male Wistar rats with an average weight of 200-240 g (institute of laboratory animal science, Chinese Academy of Medical Sciences, Beijing, China) were used in this study. Animals were housed in a climate-controlled vivarium with 12-h day and night cycles and were fed a standard laboratory diet and water ad libitum. The animals were randomly assigned to the chronic renal failure (CRF) and sham-operated control groups. The CRF group underwent 5/6 nephrectomy by surgical resection of the upper and lower thirds of left kidney, followed by tight nephrectomy 7 days later. The control group underwent sham operation. The procedures

were carried out under general anesthesia (sodium pentobarbital, 50 mg/kg, i.p.) using strict hemostasis and aseptic techniques. The 5/6 nephrectomized animals were randomly divided into untreated and CA-treated (20 and 60 mg/kg, dissolved in 0.5% carboxymethycellulose-Na buffer, p.o.) subgroups. The whole administration lasted for 20 weeks. The untreated group received regular water instead. Ten animals were included in each group. At the time of 10 weeks and 20 weeks, blood pressure, including the systolic blood pressure (SAP) and diastolic blood pressure [19], were measured by tail-cuff plethysmography (BP-98A; softron, Tokyo, Japan) with prior training to minimize variability in the blood pressure measurement.

At the end of the experiment, animals were anesthetized (sodium pentobarbital, 50 mg/kg i.p.) and euthanized by exsanguinations using cardiac puncture. Kidneys were removed. A piece of the kidney was separated and fixed in 10% formalin for histological examination.

All the animal experiments were approved by the Ethics Committee of Laboratory Animals of Yuhuangding Hospital, the protocol was approved on 15th, June 2014.

# Blood and urine chemistry

At the times when CA was administered for 10 and 20 weeks, the blood of rats was sampled through the eyes after rats were anesthetized with diethyl ether. Twenty-four-hour urine collections were obtained in each animal after placement in metabolic cage the day before collecting blood samples. Urine protein concentration, blood urea nitrogen (BUN) and plasma creatinine (Scr) were measured by the standard biochemical kits (Nanjing Jiancheng LTD, Nanjing, China) respectively.

### Assessment of antioxidant profile

Renal tissue was homogenized in 10 volume of 100 mmol  $KH_2PO_4$  buffer containing 1 mmol EDTA (pH 7.4) and centrifuged at 12,000× g for 30 min at 4°C. The supernatant was collected and used for enzymatic studies. Catalase assay (CAT), superoxide dismutase assay (SOD), Estimation of lipid peroxidation assay (TBARS), Glutathione-S-transferase assay (GST), Glutathione peroxidase assay (GSH-px), Reduced glutathione



Figure 1. CA decreased systolic blood pressure and diastolic blood pressure on 5/6 nephrectomized rats: Data are means  $\pm$  S.D. (n=10). \*\*P<0.01, versus sham control; #P<0.05, versus 5/6 Nx model control.

assay (GSH) was conducted respectively according to the instruction of kit manual.

#### Histological examination

The fixed renal tissue blocks were embedded in paraffin. Sections of 2  $\mu$ m thicknesses were cut and stained with hematoxylin and eosin (H&E) and Periodic acid-Schiff (PAS). The stained sections were examined under a light microscope at a magnification 200× and the severity of glomerulosclerosis was graded in a blind manner on a scale of 0-4 as previously [20]: Grade 0, normal; Grade 1, sclerotic area up to 25% (minimal); Grade 2, sclerotic area 25-50% (moderate); Grade 3, sclerotic area 50-75% (moderate to severe) and Grade 4, sclerotic area 75-100% (severe). The scores from each individual glomerulus examined (100 glomeruli from each rat) were averaged for each rat.

As for tubulointerstitial damage, a scoring system was applied (from 0 to 4), in which tubular atrophy, dilation, casts, interstitial inflammation, and fibrosis were assessed in 10 kidney fields at a magnification of ×200: 0, normal; 1, lesions in <25% of the area; 2, lesions in 25% to 50% of the area; 3, lesions in >50% of the area; and 4, lesions involving the entire area [21, 22].

#### Immunohistochemistry

Expression of TGF<sub>β</sub>1, fibronectin and Collagen IV in kidney was detected by the streptavidinperoxidase-biotin (SP) immunohistochemical method. The kidneys were cut into 4 µm thick sections, which were placed on immunohistochemical slides. After dehydration with xylene and alcohol, the antigen retrieval was accomplished. The sections were incubated in a blocking solution of 10% normal goat serum diluted in PBS, and incubated with polyclonal rabbit anti-TGF<sub>B1</sub>, fibronectin and Collagen IV antibodies (1:200, Abcam, USA) for 2 h, then incubated for 10 min at room temperature with biotinylated goat anti-rabbit secondary antibody (1:500). After that, sections were exposed to streptavidin-biotin complex conjugated to HRP. Sections were then incubated with diaminobenzene (DAB) for 5-10 min and observed under the microscope. Each sample was examined at 200× magnification under a light microscope (Leica, Germany). For each slide, the labeled surface area for three different fields, which came from three random, non-overlapping areas of one slice from each mouse, was evaluated by quantitative image analysis using Image-Pro Plus software (Bethesda, MD, USA), and the mean of the three different fields was calculated. Positive areas were expressed as percentage of positive area per glomerulus.

# Western blot analysis of TGF $\beta$ 1 and samd2/p-smad2

To detest the expression of TGF- $\beta$ 1, samd2 and p-smad2 in renal tissues, we performed the western blotting. Renal tissue lysate was prepared using a lysis buffer containing 25 mmol/L Tris-HCl, 150 mmol/l NaCl, 5 mmol/L ethylene glycol bis (2-aminoethyl ether) tetraacetic acid

Index	Sham	5/6 Nx	5/6Nx+CA (20 mg/kg)	5/6Nx+CA (60 mg/kg)
Ν	10	10	10	10
Body weight (g)	445.0±23.9	409.0±12.3	411.8±21.2	408.0±27.5
24-h urine protein (mg/day)	13.61±3.91	98.22±22.1ª	54.77±21.02°	31.01±3.62°
Blood urea nitrogen (mg/dL)	19.7±1.6	48.9±12.2ª	34.9±6.9	25.2±3.6
Plasma creatinine (mg/dL)	1.64±0.34	3.72±0.56ª	2.81±0.40°	2.16±0.43°
Creatinine clearance (ml/min)	4.1±1.32	2.1±0.56ª	3.2±1.21 <sup>b</sup>	3.5±1.40 <sup>b</sup>

Table 1. General data in the 5/6 nephrectomized rats after treatment with the CA for 10 weeks

°P<0.01, versus sham; °P<0.05, °P<0.01, versus 5/6 Nx group.

 Table 2. General data in the 5/6 nephrectomized rats after treatment with the CA for 20 weeks

Index	Sham	5/6 Nx	5/6 Nx+CA (20 mg/kg)	5/6Nx+CA (60 mg/kg)
Ν	10	10	10	10
Body weight (g)	502.7±32.1	446.7±22.1	447.0±34.7	445.3±23.8
24-h urine protein (mg/day)	19.39±5.27	198.8±52.3ª	75.10±17.75°	54.21±14.46°
Blood urea nitrogen (mg/l)	18.9.0±3.4	62.8±22.4ª	41.5±12.4	32.7±7.7 <sup>b</sup>
Plasma creatinine (mg/l)	1.12±0.17	2.53±1.06ª	1.59±0.19	1.25±0.15 <sup>b</sup>
Creatinine clearance (ml/mim)	3.7±0.92	1.2±0.44ª	2.4±0.58 <sup>b</sup>	3.3±0.87°

<sup>a</sup>P<0.01, versus sham; <sup>b</sup>P<0.05, <sup>c</sup>P<0.01, versus 5/6 Nx.

(EGTA), 5 mmol/L ethylene diamine tetraacetic acid (EDTA), 10 mmol/L NaF, 1 mmol/L phenylmethyl sulfonylfluoride (PMSF), 1% TritonX-100, 0.5% Nonidet P40, 10 mg/L aprotinin, 10 mg/L leupeptin and quantified by Bradford dye-binding procedure. Equal amounts of protein were separated by sodium dodecyl sulphate (SDS)polyacrylamide gel electrophoresis (PAGE) (5% stacking gel and 10% separating gel for β-actin (mouse monoclonal, Santa Cruz, USA), smad2 (rabbit monoclonal, CST, USA) and p-smad2 (rabbit monoclonal, CST, USA) or 12% separating gel for TGF-B1 (rabbit monoclonal, CST, USA) and electroblotted onto nitrocellulose. After blocking with 3% bovine serum albumin [23], the membranes were incubated with appropriate antibodies overnight. Membranes were then incubated with a horseradish peroxidaseconjugated secondary anti-body against rabbit or mouse immunoglobulin G (IgG) at a 1:2000 dilution for 1 h at room temperature after being washed. Reactive proteins were viewed by ECL. The signals were detected with Fuji Film Las-3000 (Tokyo, Japan). The intensity of the detected bands was analyzed using Image J program.

# Statistical analysis

All the values were represented as the means  $\pm$  standard error (S.EM.) and were analyzed by ANOVA and post hoc Bonferroni test. Difference was considered significant when P<0.05.

# Results

### CA decreased the SAP and DAP in 5/6 Nx rats

As shown in **Figure 1**, 20 weeks later, the 5/6 Nx rats established significant higher blood pressure compared to sham control after operation. After treatment for 20 weeks, CA treatment reduced the SAP by almost 28% at 60 mg/kg compared to model control (P<0.05); meanwhile, 20 mg/kg CA treatment also showed a certain effect. CA also could reduce DAP at dose dependent manner.

# CA reduced the Scr, BUN and proteinurea in 5/6 Nx rats during the long-term treatment

As expected, all the 5/6 nephrectomized animals had significant elevation of Scr, BUN and proteinuria (**Tables 1** and **2**). Ten-week administration of CA already significantly improved these biochemical parameters (**Table 1**, P<0.05). As shown in **Table 2**, after administration for 20 weeks, CA showed significant beneficial effect on reducing Scr, BUN and proteinuria, increase the creatinine clearance (P<0.01).

# Effects of CA on antioxidant profile

The results regarding the protective effects of CA against the oxidative stress in rat on kidney protein and activities of antioxidant enzymes such as CAT, SOD, GSH-Px, GSR and GST are

Treatment	CAT (U/min)	SOD (U/mg protein)	GSH-Px (nmol/ mg protein)	GSR (nmol/min/ mg protein)	GST (nmol/min/ mg protein)
Sham	18.7±2.4	20.45±2.04	46.45±1.67	211.5±9.5	130.9±14.2
5/6 Nx	6.5±0.58ª	7.13±1.87ª	19.14±2.16ª	106.4±11.8ª	57.34±15.6ª
5/6 Nx+CA (20 mg/kg)	10.4±1.12 <sup>b</sup>	11.55±1.62⁵	30.15±2.47⁵	147.4±13.8 <sup>b</sup>	105.4±12.3 <sup>b</sup>
5/6 Nx+CA (60 mg/kg)	16.34±2.83°	17.49±1.53°	42.26±2.12°	194.5±12.9°	118.3±10.4°

 Table 3. Effects of CA on renal antioxidant profile

°P<0.01, versus sham control; °P<0.05, °P<0.01, versus 5/6 Nx group.



Figure 2. Representations of PAS-stained and HE-stained kidneys of 5/6 (Nx) sham-operated rats, model control and CA-treated animals; quantitative analysis of glomerulosclerosis scores and tubulointerstitial damage index were also shown. Data are means  $\pm$  S.D. (n=10). \*\*P<0.01, versus sham control; #P<0.05, versus 5/6 Nx model control.

shown in **Table 3.** Activities of antioxidant enzymes such as CAT, SOD, GSH-Px, GSR and GST were reduced (P<0.01) in the 5/6 Nx as compared to control group. This reduction in enzymes activity was reversed significantly (P<0.01), in a concentration dependent way, by the treatment of CA as compared to the 5/6 Nx group.

# Benefits of CA on renal histopathological injuries

The 5/6 Nx group displayed glomerular hypertrophy, mesangial cell proliferation, mesangial matrix accumulation, telangiectasia or occlusions of the apillaries, thickening of the glomerular capsule wall, and focal or global sclerosis of some glomeruli. Furthermore, the renal tubules in this group showed dilation or atrophy, a large number of protein casts, interstitial widening, substantial infiltration of inflammatory cells, and focal distribution of renal interstitial microangiopathy, with narrowing and distortion of capillary cavities.

Compared with the 5/6 Nx group, the pathological changes in both CA-treated groups were alleviated to different extents: lower glomerular sclerosis scores and tubulointerstitial scores were observed in these groups in a dose dependent manner (**Figure 2**, P<0.05).

# Chlorogenic acid improves renal function in 5/6 Nx rats



**Figure 3.** Expression of  $\alpha$ -SMA, fibronectin and Collagen I in the cortex of kidney (200×) with quantitative analysis. \*\*P<0.01, versus sham control; #P<0.05, versus 5/6 Nx model control.



Figure 4. Western blot of TGFβ1, smad2 and p-smad2 in the kidney tissues with quantitative analysis. \*\*P<0.01, versus sham control; #P<0.05, ##P<0.01 versus 5/6 Nx model control.

Expression of  $\alpha$ -SMA, fibronectin and collagen I in kidneys by immunohistochemistry

As shown in **Figure 3**, after raising for 20 weeks, animals with 5/6 Nx showed higher expression of  $\alpha$ -SMA, fibronectin and collagen I by immunohistochemistry in glomeruli compared with that of sham controls (P<0.05). Treatment with 20 or 60 mg/kg CA dose-dependently decreased expression of these fibrosis-related biomolecules in glomeruli and renal interstitium (P<0.01).

# Expression of TGF $\beta$ 1 and samd2 in kidney tissues

Western blotting analyses showed that the expression of TGF- $\beta$ 1 in kidney tissues was reduced by the CA treatment, in a dose dependent manner (**Figure 4**), same trend as well as p-smad2, while for the total smad2, CA did not show significant influence on its protein level.

These findings suggested that CA might inhibit the renal fibrosis via modulation of TGF- $\beta$ 1 signaling pathway in the end stage of renal dysfunction.

### Discussions

The 5/6 nephrectomy rat is a classic model of progressive renal scarring characterized by both glomerulosclerosis and interstitial fibrosis, in which both glomerular and peritubular capillary endothelial injuries have been reported [24-26]. In this study, the levels of creatinine, BUN and proteinuria in the 5/6 Nx group were progressively elevated, which were also accompanied by typical pathological changes. Oral administration of CA was shown to ameliorate glomerulosclerosis and renal interstitial fibrosis in this study, which indicated that CA has the same renoprotective effects in the 5/6 Nx model as it was reported to ameliorate diabetic nephropathy and other drug-induced nephropathy.

Previous studies suggest that anti-oxidation characteristic of CA plays an important role in its renal beneficial effect [27-30]. In the current study, we introduce 5/6 Nx animal model to mimic the progression of renal damage resulting from reduced nephron mass. This model has been used to investigate the effects of drugs on pathophysiological events in the progression of glomerulosclerosis and chronic renal failure, which is characteristic of advanced-stage or end-stage renal dysfunction [31].

Currently, we successfully established 5/6 Nx rat model. The urinary excretion of protein in nephrectomised rats increased with time more markedly than in the sham-operated rats, as well as BUN and Scr; the blood pressure in the 5/6 Nx rats gradually rose during the experiment; in the present study, CA significantly reduced the blood pressure both for DAP and SAP, which is consistent previous studies [13, 14]. Mechanistically, the metabolites of CA attenuate oxidative stress (reactive oxygen species), which leads to the benefit of blood-pressure reduction through improved endothelial function and nitric oxide bioavailability in the arterial vasculature [13].

After treatment for 20 weeks, CA improved the blood urea nitrogen and creatinine clearance significantly, also it reduced the albumin excretion and urine protein level in the urine of 5/6 Nx rats (**Tables 1** and **2**), compared to model control. Urine protein level increases progressively with CRF progression, and urinary protein level was a key determinant for renal dysfunction via worsening tubulointerstitial fibrosis [32, 33]. By reducing the urine protein, CA treatment improved the kidney filtration and slowed down the progress of chronic renal failure.

By histopathological analysis, we further confirmed that CA has satisfactory beneficial effect on 5/6 Nx nephropathy in Wistar rats, CA significantly suppressed the glomerulosclerosis and tubulointerstitial damage in a dose-dependent manner (P<0.05 or P<0.01).

Cytokines-driven glomerular mesangial cell proliferation and overproduction of extracellular matrix (ECM) play important roles in the response of the kidneys to injury and in the development of glomerulosclerosis. Fibronectin,  $\alpha$ -SMA and collagen I are important fac-

tors for overproduction of ECM. By immunohistochemistry, we could prove that CA could inhibit the content of fibronectin,  $\alpha$ -SMA and collagen I in the renal interstitium.

TGF<sub>β1</sub> plays a key role in the renal fibrosis at the end stage of renal dysfunction. In the current study, we found lower TGF<sub>β1</sub> level in the CA-treated groups in all three doses. TGF<sup>β1</sup> overexpression contributes to progressive renal fibrosis [34], TGF-B1 is widely expressed in all cells of the kidney, where it exerts proinflammatory and profibrotic effects, mediating extracellular matrix deposition, increasing the synthesis of matrix components and reducing their degradation [35]. Putative factors inducing TGF-B1 expression include overload of renal cells with excessive filtered plasma proteins, renin-angiotensin system activation and hyperglycemia [36, 37]. Experimental studies have suggested inhibiting TGF-B1 can prevent renal insufficiency [38, 39]. By present study, we further confirm one of underlying mechanisms of CA's renal protective effect is due to down-regulation of TGF-β1.

Increased oxidation also enhanced the progression of CRF in 5/6 Nx rats. In the present study, the mean activity of the antioxidant enzymes CAT, SOD, GSH-Px, GST and GSR were found to be significantly lowered in the model group compared with that of the sham control group. However, the treatment of CA modified the biochemical changes caused by nephron loss in rat. In the present study, the mean activities of antioxidant enzymes were significantly higher compared with those of the model group and thus had a potential protective effect.

In a conclusion, long-term CA administration improves hypertension and proteinuria and attenuates glomerulosclerosis and tubularintertinum injuries in the remnant kidneys of animals with CRF induced by subtotal nephrectomy; it may be due to the anti-oxidation effect and blocking of TGF-smad signal pathway.

# Disclosure of conflict of interest

### None.

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#### References

- [1] Satirapoj B, Wang Y, Chamberlin MP, Dai T, LaPage J, Phillips L, Nast CC and Adler SG. Periostin: novel tissue and urinary biomarker of progressive renal injury induces a coordinated mesenchymal phenotype in tubular cells. Nephrol Dialys Transplant 2012; 27: 2702-2711.
- [2] Levey AS, Atkins R, Coresh J, Cohen EP, Collins AJ, Eckardt KU, Nahas ME, Jaber BL, Jadoul M, Levin A, Powe NR, Rossert J, Wheeler DC, Lameire N and Eknoyan G. Chronic kidney disease as a global public health problem: approaches and initiatives-a position statement from Kidney Disease Improving Global Outcomes. Kidney Int 2007; 72: 247-259.
- [3] Upadhyay R and Mohan Rao LJ. An outlook on chlorogenic acids-occurrence, chemistry, technology, and biological activities. Crit Rev Food Sci Nutr 2013; 53: 968-984.
- [4] Xiang Z and Ning Z. Scavenging and antioxidant properties of compound derived from chlorogenic acid in South-China honeysuckle. LWT-Food Sci Technol 2008; 41: 1189-1203.
- [5] Ji L, Jiang P, Lu B, Sheng Y, Wang X and Wang Z. Chlorogenic acid, a dietary polyphenol, protects acetaminophen-induced liver injury and its mechanism. J Nutr Biochem 2013; 24: 1911-1919.
- [6] Kim J, Lee S, Shim J, Kim HW, Kim J, Jang YJ, Yang H, Park J, Choi SH, Yoon JH, Lee KW and Lee HJ. Caffeinated coffee, decaffeinated coffee, and the phenolic phytochemical chlorogenic acid up-regulate NQO1 expression and prevent H(2)O(2)-induced apoptosis in primary cortical neurons. Neurochem Int 2012; 60: 466-474.
- [7] Hwang SJ, Kim YW, Park Y, Lee HJ and Kim KW. Anti-inflammatory effects of chlorogenic acid in lipopolysaccharide-stimulated RAW 264.7 cells. Inflamm Res 2014; 63: 81-90.
- [8] Shi H, Dong L, Jiang J, Zhao J, Zhao G, Dang X, Lu X and Jia M. Chlorogenic acid reduces liver inflammation and fibrosis through inhibition of toll-like receptor 4 signaling pathway. Toxicology 2013; 303: 107-114.
- [9] Domitrovic R, Jakovac H, Romic Z, Rahelic D and Tadic Z. Antifibrotic activity of Taraxacum officinale root in carbon tetrachloride-induced liver damage in mice. J Ethnopharmacol 2010; 130: 569-577.
- [10] Domitrovic R, Cvijanovic O, Susnic V and Katalinic N. Renoprotective mechanisms of chlorogenic acid in cisplatin-induced kidney injury. Toxicology 2014; 324: 98-107.

- [11] Jin S, Chang C, Zhang L, Liu Y, Huang X and Chen Z. Chlorogenic acid improves late diabetes through adiponectin receptor signaling pathways in db/db mice. PLoS One 2015; 10: e0120842.
- [12] Karthikesan K, Pari L and Menon VP. Antihyperlipidemic effect of chlorogenic acid and tetrahydrocurcumin in rats subjected to diabetogenic agents. Chem Biol Interact 2010; 188: 643-650.
- [13] Zhao Y, Wang J, Ballevre O, Luo H and Zhang W. Antihypertensive effects and mechanisms of chlorogenic acids. Hypertens Res 2012; 35: 370-374.
- [14] Suzuki A, Yamamoto N, Jokura H, Yamamoto M, Fujii A, Tokimitsu I and Saito I. Chlorogenic acid attenuates hypertension and improves endothelial function in spontaneously hypertensive rats. J Hypertens 2006; 24: 1065-1073.
- [15] Alvarez-Prats A, Hernandez-Perera O, Diaz-Herrera P, Ucero AC, Anabitarte-Prieto A, Losada-Cabrera A, Ortiz A and Rodriguez-Perez JC. Combination therapy with an angiotensin II receptor blocker and an HMG-CoA reductase inhibitor in experimental subtotal nephrectomy. Nephrol Dialys Transplant 2012; 27: 2720-2733.
- [16] Remuzzi G. Nephropathic nature of proteinuria. Curr Opin Nephrol Hypertens 1999; 8: 655-663.
- [17] Sasaki M, Shikata K, Okada S, Miyamoto S, Nishishita S, Kataoka HU, Sato C, Wada J, Ogawa D and Makino H. The macrophage is a key factor in renal injuries caused by glomerular hyperfiltration. Acta Med Okayama 2011; 65: 81-89.
- [18] Ozcan A, Ware K, Calomeni E, Nadasdy T, Forbes R, Satoskar AA, Nadasdy G, Rovin BH, Hebert LA and Brodsky SV. 5/6 nephrectomy as a validated rat model mimicking human warfarin-related nephropathy. Am J Nephrol 2012; 35: 356-364.
- [19] Stearns V, Chapman JA, Ma CX, Ellis MJ, Ingle JN, Pritchard KI, Budd GT, Rabaglio M, Sledge GW, Le Maitre A, Kundapur J, Liedke PE, Shepherd LE and Goss PE. Treatment-associated musculoskeletal and vasomotor symptoms and relapse-free survival in the NCIC CTG MA.27 adjuvant breast cancer aromatase inhibitor trial. J Clin Oncol 2015; 33: 265-271.
- [20] Tapia E, Sanchez-Lozada LG, Soto V, Manrique AM, Ortiz-Vega KM, Santamaria J, Medina-Campos ON, Cristobal M, Avila-Casado C, Pedraza-Chaverri J, Rodriguez-Iturbe B and Franco M. Sildenafil treatment prevents glomerular hypertension and hyperfiltration in rats with renal ablation. Kidney Blood Press Res 2012; 35: 273-280.

- [21] Porsti I, Fan M, Koobi P, Jolma P, Kalliovalkama J, Vehmas TI, Helin H, Holthofer H, Mervaala E, Nyman T and Tikkanen I. High calcium diet down-regulates kidney angiotensin-converting enzyme in experimental renal failure. Kidney Int 2004; 66: 2155-2166.
- [22] Li P, Ma LL, Xie RJ, Xie YS, Wei RB, Yin M, Wang JZ and Chen XM. Treatment of 5/6 nephrectomy rats with sulodexide: a novel therapy for chronic renal failure. Acta pharmacol Sin 2012; 33: 644-651.
- [23] Reigstad MM, Larsen IK, Myklebust TA, Robsahm TE, Oldereid NB, Omland AK, Vangen S, Brinton LA and Storeng R. Risk of breast cancer following fertility treatment-A registry based cohort study of parous women in Norway. Int J Cancer 2015; 136: 1140-1148.
- [24] Zhong F, Liu X, Zhou Q, Hao X, Lu Y, Guo S, Wang W, Lin D and Chen N. 1H NMR spectroscopy analysis of metabolites in the kidneys provides new insight into pathophysiological mechanisms: applications for treatment with Cordyceps sinensis. Nephrol Dial Transplant 2012; 27: 556-565.
- [25] Floege J, Alpers CE, Burns MW, Pritzl P, Gordon K, Couser WG and Johnson RJ. Glomerular cells, extracellular matrix accumulation, and the development of glomerulosclerosis in the remnant kidney model. Lab Invest 1992; 66: 485-497.
- [26] Kren S and Hostetter TH. The course of the remnant kidney model in mice. Kidney Int 1999; 56: 333-337.
- [27] Sadeghnia HR, Yousefsani BS, Rashidfar M, Boroushaki MT, Asadpour E and Ghorbani A. Protective effect of rutin on hexachlorobutadiene-induced nephrotoxicity. Ren Fail 2013; 35: 1151-1155.
- [28] Korkmaz A and Kolankaya D. Protective effect of rutin on the ischemia/reperfusion induced damage in rat kidney. J Surg Res 2010; 164: 309-315.
- [29] Kamalakkannan N and Stanely Mainzen Prince P. Rutin improves the antioxidant status in streptozotocin-induced diabetic rat tissues. Mol Cell Biochem 2006; 293: 211-219.
- [30] Shimoi K, Shen B, Toyokuni S, Mochizuki R, Furugori M and Kinae N. Protection by alpha Grutin, a water-soluble antioxidant flavonoid, against renal damage in mice treated with ferric nitrilotriacetate. Jpn J Cancer Res 1997; 88: 453-460.

- [31] Tsunenari I, Ohmura T, Seidler R, Chachin M, Hayashi T, Konomi A, Matsumaru T, Sumida T, Hayashi N and Horie Y. Renoprotective effects of telmisartan in the 5/6 nephrectomised rats. J Renin Angiotensin Aldosterone Syst 2007; 8: 93-100.
- [32] Breigeiron MK, Lucion AB and Sanvitto GL. Effects of renovascular hypertension on reproductive function in male rats. Life Sci 2007; 80: 1627-1634.
- [33] Ohno T, Takemura G, Murata I, Kagawa T, Akao S, Minatoguchi S, Fujiwara T and Fujiwara H. Water extract of the root of Lindera strychnifolia slows down the progression of diabetic nephropathy in db/db mice. Life Sci 2005; 77: 1391-1403.
- [34] Reeves WB and Andreoli TE. Transforming growth factor beta contributes to progressive diabetic nephropathy. Proc Natl Acad Sci U S A 2000; 97: 7667-7669.
- [35] Li H, Zheng X, Wang H, Zhang Y, Xin H and Chen X. XLF-III-43, a novel coumarin-aspirin compound, prevents diabetic nephropathy in rats via inhibiting advanced glycation end products. Eur J Pharmacol 2010; 627: 340-347.
- [36] Gagliardini E and Benigni A. Therapeutic potential of TGF-beta inhibition in chronic renal failure. Expert Opin Biolog Ther 2007; 7: 293-304.
- [37] Gagliardini E and Benigni A. Role of anti-TGFbeta antibodies in the treatment of renal injury. Cytokine Growth Factor Rev 2006; 17: 89-96.
- [38] Ziyadeh FN, Hoffman BB, Han DC, Iglesias-De La Cruz MC, Hong SW, Isono M, Chen S, Mc-Gowan TA and Sharma K. Long-term prevention of renal insufficiency, excess matrix gene expression, and glomerular mesangial matrix expansion by treatment with monoclonal antitransforming growth factor-beta antibody in db/db diabetic mice. Proc Natl Acad Sci U S A 2000; 97: 8015-8020.
- [39] Zhang HJ, Jin J, Zhou WQ, Zhang Y, Li Y, Liu JY, Zhang S and Chen XG. Nicousamide, a potent inhibitor of phosphorylation by TGF-β receptor II. Acta Pharmaceutica Sinica B 2011; 1: 160-165.