Original Article

Diagnostic value of long non-coding RNA H19, UCA1, and HOTAIR as promising biomarkers in human bladder cancer

Wei Wang¹, Zhihua Yin²

¹Department of Drug Control, Criminal Investigation Police University of China, Shenyang, P. R. China; ²Department of Public Health, China Medical University, Shenyang, P. R. China

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Abstract: Aim: Currently, there are no satisfactory markers for bladder cancer available in clinics. In this study, we selected 3 long noncoding RNAs, H19, UCA1, and HOTAIR, to assess their diagnostic value in bladder cancer screening. Method: In this research, a total of 96 cancer tissues and paired non-cancer tissues from patients with bladder cancer (BC) were collected. Total RNA was isolated from tissues using TRIzol reagent and performance of three selected IncRNAs as cancer markers were analyzed by reverse transcription-PCR. A new IncRNA-based score was conducted by Logistic regression model. Receiver operating characteristic (ROC) curves and area under the ROC curve (AUC) was generated to assess the diagnostic values of the IncRNAs. Result: Expression levels of IncRNAs H19, UCA1 and HOTAIR were remarkably increased in bladder cancer tissues compared with those in normal tissues (all P<0.001). The AUC (95% CI) of H19, UCA1, and HOTAIR were 0.717 (0.647-0.779), 0.787 (0.722-0.843), 0.713 (0.643-0.776), respectively. We conducted a new score named IncRNA-score, based on three selected LncRNAs: Lnc-Score =0.48*H19+0.49*UCA1+1.2*HOTAIR, which had the best diagnosability performance with AUC of 0.870 (0.814-0.914), sensitivity of 70.8%, specificity of 88.5%, +LR of 3.67 and -LR of 0.22. Conclusion: The 3 selected IncRNAs were remarkably increased in bladder cancer tissues and have great potential to be new sensitive, reliable biomarkers for diagnosis of bladder cancer.

Keywords: IncRNA, bladder cancer, diagnose, ROC analysis

Introduction

As one of the most common types of genitourinary tumors, human bladder cancer accounts for one half of all tumors of the urinary system [1, 2]. Despite increased early detection of bladder cancer and more frequent surgery, the incidence and mortality of bladder cancer has not changed significantly in the past several decades [3]. Therefore, an urgent need of searching new sensitive, reliable biomarkers is emphasized for bladder cancer.

Deep sequencing recently facilitated the discovery of thousands of novel transcripts, now classified as long noncoding RNAs (IncRNAs), in many vertebrate and invertebrate species [4]. Like microRNAs, IncRNA are also proving to be the key mediators of cellular differentiation, cell lineage choice, organogenesis and tissue

homeostasis [4, 5]. Dysregulation of IncRNA expression has been shown to be important in carcinogenesis and cancer metastasis [6-8]. A study that considered samples from 5,037 human tumor specimens from the cancer genome atlas project, demonstrated that, IncRNAs are highly cancer type specific compared with protein-coding genes, which provides a resource for investigating IncRNA in cancer and lays the groundwork for the development of new sensitive, reliable biomarkers for diagnosis and treatment.

Recently, most studies focus on revealing the molecular mechanism of IncRNA in cancer mechanism. In the present study, we determined the expression of selected 3 candidate IncRNAs (H19, UCA1 and HOTAIR) in human bladder tissues and assess their diagnostic values in BC screening by comparing their expres-

Table 1. Characteristics of bladder cancer patients

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Characteristics	BC patients (n=96)				
Age	61.3±6.2				
Male	69 (71.8%)				
Alcohol use					
Yes	32 (33.3%)				
No	64 (66.6%)				
Tobacco use					
Yes	28 (29.2%)				
No	68 (70.8%)				
Family history of BC					
Yes	10 (10.4%)				
No	86 (89.6%)				
TNM					
+	39 (40.6%)				
III+IV	57 (59.4%)				

sion level in cancer tissues and pair-matched adjacent normal tissues.

Materials and methods

Subjects

Total of 96 cancer tissues and paired non-cancer tissues (defined as >2 cm distance from tumor edge) from patients with urothelial carcinoma were collected. Then tissues were aliquoted into microcentrifuge tubes, marked and stored at -80°C within 2 hours of collection. Clinical data were collected, including gender, age, smoking, drinking and TNM stage. This research was conducted in strict accordance with the protocol approved by the Ethics Committee of Peking Union Medical College Hospital, and a written informed consent was obtained from each subject before their participation in the study.

Diagnosis of primary BC was clinically confirmed by histopathology or biopsy and patients who meet the following criteria were excluded 1) who has severe infection, active clinical comorbidities, or a history of any other malignancy; 2) who has received any chemotherapy, radiotherapy, or operation; 3) older than 75 years or younger than 40 years.

RNA isolation

Total RNA was isolated from tissues using TRIzol reagent according to the manufacturer's

protocol (Invitrogen). Briefly, 1 ml TRIzol was added to 100 mg of sample tissues, and then homogenated completely by a power homogenizer. 0.2 mL isopropanol was added to the mixture and incubate at room temperature for 10 minutes. Then the sample was centrifuged at 12,000×g for 10 minutes at 4°C. Wash the pellet, with 1 mL of 75% ethanol and centrifuge the tube at 7500×g for 5 minutes at 4°C. Finally, the RNA pellet was resuspended with RNase-free water. The concentration and purity of the RNA solution was measured by detecting its absorbance at 260/280 and 260/230 nm with NanoDrop 1000A spectrophotometer. (NanoDrop Technologies, Wilmington, DE). All the purified RNA samples were stored at -80°C for further processing.

Reverse transcription and quantitative realtime PCR (q-RTPCR)

To synthesize single-stranded cDNA from total RNA, we used the High Capacity cDNA Reverse Transcription Kits (Thermo). Prepare the $2\times RT$ master mix using the kit components before preparing the reaction plate. 2 μg of total RNA was added to 20 μL reaction. The reverse transcription was performed on a MJ Research PTC-200 Peltier Thermal Cycler (Global Medical Instrumentation) at $25^{\circ} C$ for 10 min, $37^{\circ} C$ for 120 min, and $85^{\circ} C$ for 5 min.

Then gRT-PCR was performed to quantify the expression level of Long non-coding RNA H19, UCA1, and HOTAIR with SYBR Green PCR Master Mix (Thermo) following the manufacturer's instructions. The amplifications reaction contained: master mix, forward primer, reverse primer, and nuclease-free water. GAPDH was used as an intrinsic control, H19 forward, 5'-ATCGGTGCCTCAGCGTTCGG-3' and reverse, 5'-CTGTCCTCGCCGTCACACCG-3'; UCA1 forward, 5'-ACGCTAA CTGGCACCTTGTT-3' and reverse, 5'-TGGGGATTACTGGGGTAGGG-3'; HO-TAIR forward, 5'-GCTGCTCCGGAATTTGAGAG-3' and reverse, 5'-TGCTGC CAGTTAGAAAAGCG-3'; GAPDH forward, 5'-AGCCA CATCGCTCAGACAC-3' and reverse, 5'-GCCCAATACGACCAAATCC-3'. The RT-PCR reaction was performed at 95°C for 2 min and in 40 cycles at 95°C for 15 s and 60°C for 1 min on an ABI 7500 thermocycler (Applied Biosystems). Relative gene expression level of each LncRNA was analyzed using the $2^{-\Delta\Delta Ct}$ method.

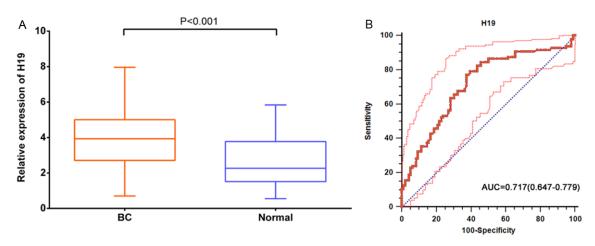


Figure 1. Diagnostic performance of IncRNA H19 in human bladder cancer (BC) tissue. A: Relative expression levels of IncRNA H19 in BC tissue and normal tissues. B: ROC curve analysis of IncRNA H19 in BC tissue and normal tissues.

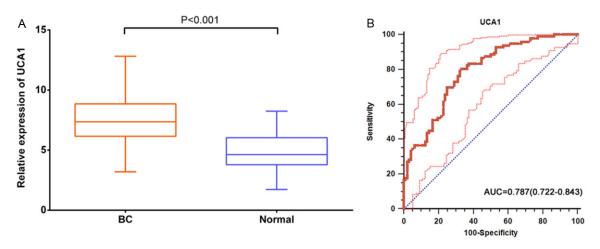


Figure 2. Diagnostic performance of IncRNA UCA1 in human bladder cancer tissue. A: Relative expression levels of IncRNA UCA1 in BC tissue and normal tissues. B: ROC curve analysis of IncRNA UCA1 in BC tissue and normal tissues.

Statistical analysis

Continuous variables are expressed as mean \pm standard deviation and categorical variables are expressed as absolute relative frequencies. We compared two groups using the t test for continuous variables and χ^2 test for categorical variables. Receiver operating characteristic (ROC) curves were constructed and area under the ROC curve (AUC) was generated to assess the diagnostic values of the candidate RNAs. We used the Logistic regression model to do the multivariable analysis, and calculate a new LncRNA-based score using the three selected LncRNAs. P value <0.05 was considered as statistically significant. All statistical analysis were

performed by R software version 3.0.1 (MathSoft Inc., USA), and the graphs were obtained from GraphPad Prism 5.0 (GraphPad Software Inc., CA).

Results

Clinical characteristics of study population

Table 1 shows detailed clinical characteristics of the 96 patients. There were 69 males and 27 females, aged from 42 from 73 (average 61.3). Among these BC patients, alcohol use accounted for 32 (33.3%), tobacco use accounted for 28 (29.2%). 10 patients had a family history of BC. Additionally, 39 patients were diagnosed as TNM I+II and 57 diagnosed as TNM III+IV.

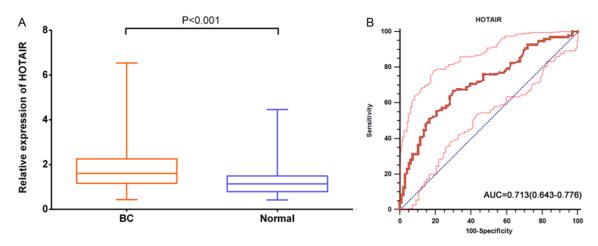


Figure 3. Diagnostic performance of IncRNA HOTAIR in human bladder cancer tissue. A: Relative expression levels of IncRNA HOTAIR in BC tissue and normal tissues. B: ROC curve analysis of IncRNA HOTAIR in BC tissue and normal tissues.

LncRNAs level in bladder cancer tissues

The expression level of three selected IncRNAs was detected by quantitative real-time PCR in BC tissues and pair-matched adjacent normal tissues form 96 BC patients. As Figures 1A, 2A. 3A shows, the relative expression of H19. UCA1, and HOTAIR in BC tissues were 3.93 (2.75, 5.00), 6.83 (5.25, 8.94), 1.61 (1.17, 2.22), respectively. While the relative expression in the normal tissues were lower, with the media (quartile) of 2.26 (1.53, 3.75), 3.99 (2.46, 5.66), 1.14 (0.80, 1.49), respectively. Statistically significant difference can be observed between BC tissues and pair-matched adjacent normal tissues (all P<0.001). Then we performed the Logistic regression model to calculate a new LncRNAbased score using the three selected LncRNAs: Lnc-Score = 0.48*H19+0.49*UCA1+ 1.2*HOTAIR.

Diagnostic performance of LncRNAs in bladder cancer detection

Receiver operating characteristic (ROC) curves were constructed and area under the ROC curve (AUC) was generated to assess the diagnostic values of the three selected LncRNAs and the Lnc-Score. As **Figures 1B**, **2B**, **3B** shows, the AUC of H19, UCA1, and HOTA-IR were 0.717 (0.647-0.779), 0.787 (0.722-0.843), 0.713 (0.643-0.776), respectively. **Table 2** shows the detail information about the ability of the three LncRNAs and Lnc-Score to diagnose the BC in patients. Among 3 selected

LncRNAs, UCA1 had the highest sensitivity of 80.2% while HOTAIR had the highest specificity of 69.9%. We also can observe that UCA1 had the highest +LR of 2.41 and the lowest -LR of 0.30. **Figure 4A** shows the distribution of risk for BC, which suggested that BC tissues had higher Lnc-Score than that of adjacent normal tissues. Compared with these 3 separate LncRNAs, Lnc-Score showed the highest AUC, with value of 0.870 (0.814-0.914). Also, the best diagnosability parameters were observed in Lnc-Score, with relative higher sensitivity of 70.8%, the highest specificity of 88.5%, highest +LR of 3.67 and lowest -LR of 0.22.

Discussion

Early diagnosis and surveillance for metastasis of bladder cancer are critical issues. Deregulation of long non-coding RNAs (IncRNAs) has been implicated in urologic malignancies and represents potential markers. In this study, we detected the relative expression levels of IncRNAs H19, UCA1 and HOTAIR in tissues from patients with bladder cancer. Higher expression of these three IncRNAs were observed in the BC tissues than those in normal tissues. Then we performed receiver operating characteristic (ROC) curves and area under the ROC curve (AUC) to assess the diagnostic values of the three IncRNAs. We can observe ideal diagnosability of the three selected IncRNAs, with AUCs of 0.717, 0.787 and 0.713, respectively. Furthermore, we constructed a new score based on these IncRNAs, named IncRNA-score, which show greater diagnosability than 3

Table 2. The receiver operating characteristic (ROC) analysis of IncRNA in human bladder cancer

miRNAs	AUC	95% CI	P value	Youden	Cut-off	Sensitivity	Specificity	+LR	-LR
H19	0.717	0.647-0.779	<0.001	0.395	2.65	77.8%	62.5%	2.06	0.37
UCA1	0.787	0.722-0.843	<0.001	0.469	4.94	80.2%	66.7%	2.41	0.30
HOTAIR	0.713	0.643-0.776	<0.001	0.365	1.34	66.7%	69.9%	2.21	0.48
Lnc-Score	0.870	0.814-0.914	<0.001	0.594	6.53	70.8%	88.5%	3.67	0.22

Note. AUC, area under the receiver operating characteristic curve; CI, confidence interval; +LR, positive likelihood ratio; -LR, negative likelihood ratio; P-value, compared with AUC of 0.5. LncRNA-Score = 0.48*H19+0.49*UCA1+1.2*HOTAIR.

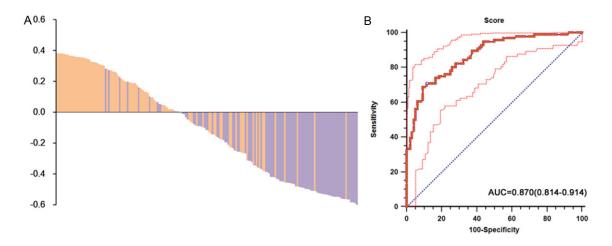


Figure 4. Diagnostic performance of LncRNA-Score in human bladder cancer tissue. A: Relative risk of LncRNA-Score in BC tissue and normal tissues. B: ROC curve analysis of LncRNA-Score in BC tissue and normal tissues.

selected miRNAs alone. Collectively, our study provides evidences that serum level of IncRNAs H19, UCA1 and HOTAIR have great clinical value as promising biomarkers in BC.

Overexpression of non-coding RNAs are implicated in metastasis of human tumors, but most are shorter non-coding like microRNAs (miR-NAs) [9]. Recently, an accumulating body of evidence has have linked specific IncRNA gene mutations with cancer, either by overlap of sequencing libraries with previously annotated GENCODE IncRNAs, or by de novo assembly of all available public datasets, raising the possibility of IncRNA-based cancer diagnostics and therapy [6, 10-13]. A study including more than 7,000 RNA-Seq libraries from 25 independent studies found that among 91,013 expressed genes, over 68% (58,648) of genes were classified as IncRNAs, of which 79% (48,952) were previously unannotated [12]. Sequencely, summary of IncRNAs with experimental data supporting the expression and functions of promoting tumor invasion and metastasis was made in some studies [6, 14].

Recently, a number of IncRNAs proved to be able to modulate Urologic neoplasms. The IncRNA RCCRT1 is upregulated remarkably in renal cell carcinoma (RCC) compared with adjacent noncancerous tissues, particularly in high-grade RCC tissues. Furthermore, siRNA-induced depletion of RCCRT1 expression suppressed migration and invasion in RCC cell lines [15]. Another study, which identified a novel IncRNA named SChLAP1, demonstrated that SChLAP1 expression increases with prostate cancer progression, and high SChLAP1 expression is associated with poor outcome after radical prostatectomy by both univariate and multivariate analysis [16].

In present study, we investigated three selected IncRNAs H19, UCA1 and HOTAIR in the bladder cancer tissues and pair-matched adjacent normal tissues from 96 patients. H19, the oldest known IncRNA, was widely studied in cancer biology even before IncRNAs had gained the attention of cancer researchers [6]. The oncogenic properties of H19 were strongly associated with antagonism of the tumor suppressor miRNA let-7 and forced EMT mediated

by the non-histone chromosomal transcriptional regulator HMGA2 [17, 18]. Also, H19 expression is directly induced by the v-myc avian myelocytomatosis viral oncogene homolog (c-MYC) and loss of the p53 tumor suppressor further supporting the importance of H19 as a potent oncogene [19, 20]. Urothelial carcinoma-associated 1 (UCA1), another upregulated IncRNA in bladder cancer, is significantly correlated with greater tumor depth and apoptosis escape [21]. Overexpression of UCA1 enhances ERK1/2 MAPK and PI3-K/AKT kinase activity, causing increased expression of the coactivator p300 and its coactivator cAMP response element-binding protein (CREB), which promotes cell cycle progression, carcinogenesis, and cancer invasion [22, 23]. HOTAIR is a noncoding 2.2-kb RNA gene located downstream, acts as a scaffold for histone modification complexes [6]. In cancer cells, HOTAIR partners with Polycomb Repressive Complex 2 (PRC2, a histone methyltransferase) to induce genomewide gene silencing and increased cancer invasiveness and metastasis in a manner dependent on PRC2 [24, 25]. Additionally, HOTAIR has been shown to regulate several genes involved in epithelial-to-mesenchyme transition (EMT) including Snail family zinc finger 1 (SNAI1), Poly r(C)-Binding Protein 1 (PCBP 1), Junctional adhesion molecule 2 (JAM2) and ABL proto-oncogene 2 (ABL2) [25-27]. These findings indicate that lincRNAs have active roles in modulating the bladder cancer and may be important targets for cancer diagnosis and therapy.

Conclusion

In conclusion, above all, our results extend the findings of previous studies about IncRNAs H19, UCA1 and HOTAIR in bladder cancer patients. Our data provide the 3 selected IncRNAs were significantly higher in bladder cancer tissues compared to normal tissues, and demonstrated the ideal diagnostic value to distinguish cancer tissues from normal tissues. However, whether this correlation is exactly proportional requires carefully scrutiny and study on a larger sample is needed to confirm this results.

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Disclosure of conflict of interest

None.

Address correspondence to: Dr. Wei Wang, Department of Drug Control, Criminal Investigation Police University of China, No. 83, The Tawan Street, Huanggu District, Shenyang 110035, P. R. China. Fax: +86-024-86982468; E-mail: zhigangji352@sina.com

References

- [1] Witjes JA, Comperat E, Cowan NC, De Santis M, Gakis G, Lebret T, Ribal MJ, Van der Heijden AG, Sherif A; European Association of Urology. EAU guidelines on muscle-invasive and metastatic bladder cancer: summary of the 2013 guidelines. Eur Urol 2014; 65: 778-792.
- [2] Yang L, Parkin DM, Li LD, Chen YD and Bray F. Estimation and projection of the national profile of cancer mortality in China: 1991-2005. Br J Cancer 2004; 90: 2157-2166.
- [3] Kim WJ and Bae SC. Molecular biomarkers in urothelial bladder cancer. Cancer Sci 2008; 99: 646-652.
- [4] Schmitz SU, Grote P and Herrmann BG. Mechanisms of long noncoding RNA function in development and disease. Cell Mol Life Sci 2016; 73: 2491-2509.
- [5] Lorenzen JM and Thum T. Long noncoding RNAs in kidney and cardiovascular diseases. Nat Rev Nephrol 2016; 12: 360-373.
- [6] Jiang C, Li X, Zhao H and Liu H. Long non-coding RNAs: potential new biomarkers for predicting tumor invasion and metastasis. Mol Cancer 2016: 15: 62.
- [7] Evans JR, Feng FY and Chinnaiyan AM. The bright side of dark matter: IncRNAs in cancer. J Clin Invest 2016; 126: 2775-2782.
- [8] Schmitt AM and Chang HY. Long noncoding RNAs in cancer pathways. Cancer Cell 2016; 29: 452-463.
- [9] Bullock MD, Sayan AE, Packham GK and Mirnezami AH. MicroRNAs: critical regulators of epithelial to mesenchymal (EMT) and mesenchymal to epithelial transition (MET) in cancer progression. Biol Cell 2012; 104: 3-12.
- [10] Beermann J, Piccoli MT, Viereck J and Thum T. Non-coding RNAs in development and disease: background, mechanisms, and therapeutic approaches. Physiol Rev 2016; 96: 1297-1325.
- [11] Silva A, Bullock M and Calin G. The clinical relevance of long non-coding RNAs in cancer. Cancers (Basel) 2015; 7: 2169-2182.
- [12] Iyer MK, Niknafs YS, Malik R, Singhal U, Sahu A, Hosono Y, Barrette TR, Prensner JR, Evans JR, Zhao S, Poliakov A, Cao X, Dhanasekaran SM, Wu YM, Robinson DR, Beer DG, Feng FY,

- lyer HK and Chinnaiyan AM. The landscape of long noncoding RNAs in the human transcriptome. Nat Genet 2015; 47: 199-208.
- [13] Yan X, Hu Z, Feng Y, Hu X, Yuan J, Zhao SD, Zhang Y, Yang L, Shan W, He Q, Fan L, Kandalaft LE, Tanyi JL, Li C, Yuan CX, Zhang D, Yuan H, Hua K, Lu Y, Katsaros D, Huang Q, Montone K, Fan Y, Coukos G, Boyd J, Sood AK, Rebbeck T, Mills GB, Dang CV and Zhang L. Comprehensive genomic characterization of long non-coding RNAs across human cancers. Cancer Cell 2015; 28: 529-540.
- [14] Bartonicek N, Maag JL and Dinger ME. Long noncoding RNAs in cancer: mechanisms of action and technological advancements. Mol Cancer 2016; 15: 43.
- [15] Song S, Wu Z, Wang C, Liu B, Ye X, Chen J, Yang Q, Ye H, Xu B and Wang L. RCCRT1 is correlated with prognosis and promotes cell migration and invasion in renal cell carcinoma. Urology 2014; 84: 730, e731-737.
- [16] Mehra R, Shi Y, Udager AM, Prensner JR, Sahu A, Iyer MK, Siddiqui J, Cao X, Wei J, Jiang H, Feng FY and Chinnaiyan AM. A novel RNA in situ hybridization assay for the long noncoding RNA SChLAP1 predicts poor clinical outcome after radical prostatectomy in clinically localized prostate cancer. Neoplasia 2014; 16: 1121-1127.
- [17] Ma C, Nong K, Zhu H, Wang W, Huang X, Yuan Z and Ai K. H19 promotes pancreatic cancer metastasis by derepressing let-7's suppression on its target HMGA2-mediated EMT. Tumour Biol 2014; 35: 9163-9169.
- [18] Yan L, Zhou J, Gao Y, Ghazal S, Lu L, Bellone S, Yang Y, Liu N, Zhao X, Santin AD, Taylor H and Huang Y. Regulation of tumor cell migration and invasion by the H19/let-7 axis is antagonized by metformin-induced DNA methylation. Oncogene 2015; 34: 3076-3084.
- [19] Barsyte-Lovejoy D, Lau SK, Boutros PC, Khosravi F, Jurisica I, Andrulis IL, Tsao MS and Penn LZ. The c-Myc oncogene directly induces the H19 noncoding RNA by allele-specific binding to potentiate tumorigenesis. Cancer Res 2006; 66: 5330-5337.

- [20] Dugimont T, Montpellier C, Adriaenssens E, Lottin S, Dumont L, lotsova V, Lagrou C, Stehelin D, Coll J and Curgy JJ. The H19 TATA-less promoter is efficiently repressed by wild-type tumor suppressor gene product p53. Oncogene 1998; 16: 2395-2401.
- [21] Xue M, Li X, Li Z and Chen W. Urothelial carcinoma associated 1 is a hypoxia-inducible factor-1alpha-targeted long noncoding RNA that enhances hypoxic bladder cancer cell proliferation, migration, and invasion. Tumour Biol 2014; 35: 6901-6912.
- [22] Wang Y, Chen W, Yang C, Wu W, Wu S, Qin X and Li X. Long non-coding RNA UCA1a(CUDR) promotes proliferation and tumorigenesis of bladder cancer. Int J Oncol 2012; 41: 276-284.
- [23] Yang C, Li X, Wang Y, Zhao L and Chen W. Long non-coding RNA UCA1 regulated cell cycle distribution via CREB through PI3-K dependent pathway in bladder carcinoma cells. Gene 2012; 496: 8-16.
- [24] Kogo R, Shimamura T, Mimori K, Kawahara K, Imoto S, Sudo T, Tanaka F, Shibata K, Suzuki A, Komune S, Miyano S and Mori M. Long noncoding RNA HOTAIR regulates polycomb-dependent chromatin modification and is associated with poor prognosis in colorectal cancers. Cancer Res 2011; 71: 6320-6326.
- [25] Gupta RA, Shah N, Wang KC, Kim J, Horlings HM, Wong DJ, Tsai MC, Hung T, Argani P, Rinn JL, Wang Y, Brzoska P, Kong B, Li R, West RB, van de Vijver MJ, Sukumar S and Chang HY. Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis. Nature 2010: 464: 1071-1076.
- [26] Wang J, Zhou Y, Lu J, Sun Y, Xiao H, Liu M and Tian L. Combined detection of serum exosomal miR-21 and HOTAIR as diagnostic and prognostic biomarkers for laryngeal squamous cell carcinoma. Med Oncol 2014; 31: 148.
- [27] Zhang ZZ, Shen ZY, Shen YY, Zhao EH, Wang M, Wang CJ, Cao H and Xu J. HOTAIR long noncoding RNA promotes gastric cancer metastasis through suppression of poly r(C)-binding protein (PCBP) 1. Mol Cancer Ther 2015; 14: 1162-1170.