Original Article Relationship between FcyRIIB gene polymorphisms, periodontitis and adverse pregnancy outcomes in pregnant women

Yanming Wang¹, Noriko Sugita², Xiaoqing Wang¹, Peisong Meng¹, Lili Gao¹, Jiang Lin¹, Hiromasa Yoshie², Liangjia Bi¹

¹Department of Stomatology, The Fourth Hospital of Harbin Medical University, Harbin, Heilongjiang Province, China; ²Division of Periodontology, Department of Oral Biological Science, Niigata University Graduate School of Medical and Dental Sciences, Niigata, Japan

Received January 19, 2017; Accepted February 22, 2017; Epub April 1, 2017; Published April 15, 2017

Abstract: Background: FcyRllb acts as a negative feedback regulator by inhibiting B-cell antigen receptor-elicited activation signals through tyrosine phosphorylation of immunoreceptor tyrosine-based inhibitory motif (ITIM). FcyRllb gene polymorphisms might be related to the lower level of IgG antibody response to periodontal bacteria and lead to the development of periodontitis. Our previous studies showed that there was significant association between FcyRllb gene polymorphisms in pregnant women and adverse pregnancy outcomes, such as preeclampsia or preterm birth with low birth weight. Periodontitis as a risk factor for adverse pregnancy outcomes has been widely and generally reported, and many studies found inflammation might lead to the development of adverse pregnancy outcomes. Review: We assume the development of adverse pregnancy outcomes may be attributed in part to inflammation such as periodontitis enhanced by FcyRllb gene polymorphisms. Although significant associations between clinical periodontal parameters and adverse pregnancy outcomes in our study. Therefore, we will summary and discuss previous studies about associations between FcyRllb gene polymorphisms, periodontitis and adverse pregnancy outcomes in our study. Therefore, we will summary and discuss previous studies about associations between FcyRllb gene polymorphisms, periodontitis and adverse pregnancy outcomes, such as the biological mechanism between them in this review.

Keywords: FcyRIIb gene polymorphisms, periodontitis, adverse pregnancy outcomes, inflammation

Introduction

Adverse pregnancy outcome represents a significant problem for modern obstetrics because of their increasing frequency and resulting socioeconomic impact. Many studies have implicated inflammation caused by maternal periodontal infection in adverse pregnancy outcomes development, [1-4] however, the mechanism of underlying this relationship is unclear. The role of genetic polymorphisms in systemic diseases development has been generally and widely accepted. In previous studies, we have identified significant association between FcyRIIb gene polymorphisms and periodontitis, between FcyRIIb gene polymorphisms and adverse pregnancy outcomes [5-9]. Therefore, we hypothesized the development of adverse pregnancy outcomes might be associated with FcγRIIb gene polymorphisms-associated inflammation caused by periodontal infection. Although we did not find direct significant associations between the prevalence of periodontitis and adverse pregnancy outcomes in pregnant women, a significant association between subgingival periodontal bacteria and adverse pregnancy outcomes was identified [10]. In this review, we will summarize and discuss previous studies about the association among FcγRIIb gene polymorphisms, periodontitis and adverse pregnancy outcomes, and analyze the possible biological mechanism

FcyRIIb

FcγRII is encoded by three homologous genes on chromosome 1q23: FcγRIIa, b and c [11]. FcγRIIa and FcγRIIc elicit activatory signals via



Figure 1. Signaling pathway for inhibitory FcγRIIB. FcγRIIB acts as a negative feedback regulator by inhibiting B-cell antigen receptor (BCR)-elicited activation signals through tyrosine phosphorylation of immunoreceptor tyrosine-based inhibition motif (ITIM).

an immunoreceptor tyrosine-based activation motif (IAIM). In contrast, FcyRIIb contains an immunoreceptor tyrosine-based inhibition motif (ITIM) on the cytoplasmic tail [12]. FcyRIIb encodes three transcripts (IIb, IIb, and IIb) due to alternative mRNA splicing. FcyRIIb, is exclusively expressed on B cells and has complete domains from all exons. Upon-co-crosslinking with the B-cell antigen receptor (BCR) by IgG immune complexes (ICs), FcyRIIb, acts as a negative feedback regulator by inhibiting B-cell antigen receptor-elicited activation signals through tyrosine phosphorylation of immunoreceptor tyrosine-based inhibition motif (ITIM) [13, 14] (Figure 1). The paired expression of activating and inhibitory molecules on the same cell is important for a balanced immune response. The FcyRIIb is one of receptors of the immunoglobulin G (IgG) that is the main type of antibody found in blood and extracellular fluid. By binding many kinds of pathogens such as viruses, bacteria, and fungi, IgG protects the body from infection. Previous studies have demonstrated FcyRIIb triggered inflammation using FcyRIIb deficient mice [15]. The studies of FcyRIIb expression regulation showed that the level of FcyRIIb expression might be reduced by complement component 5a, interferon-g, tumor necrosis factor-a and interleukin-1b. It was reported that interleukin-4 could up-regulate FcyRIIb expression on myeloid cells and down-regulated that on activated B cells. Interleukin-5, 10, 13 and transforming growth factor-B up-regulated FcyRIIb expression on innate effector cells [16-19].

FcγRIIb gene polymophisms and related diseases

Studies showed that a lower level of IgG production against periodontal bacteria caused by FcyRIIb gene polymorphisms may lead to periodontitis [7, 20]. IgG is the only isotype that has receptors to facilitate passage through the human placenta, thereby providing protection to the fetus in utero. Along with IgA secreted in the breast milk, residual IgG absorbed through the placenta provides the neonate with humoral immunity before its own immune system develops [21, 22]. Therefore, reduced maternal IgG levels caused by FcyRIIb gene polymorphisms might lead to the development of adverse pregnancy outcomes. These studies suggested that FcyRIIB gene polymorphisms were greatly related to the diseases.

Eleven single-nucleotide polymorphisms (SNPs) in the FcyRIIb gene were previously identified and confirmed to be FcyRIIb-specific. Of these SNPs, three SNPs and one SNP resulted in amino acid substitutions in exon4 (Thr203-Met, Tyr205-Phe and Ser207-Ala) and exon5 (Ile232-Thr), respectively. The other five SNPs were detected in introns 4 and 5, leading to no amino acid substitution [5] (Table 1). FcyRIIb-Ile232 polymorphism significantly increased in systemic lupus erythematosus and lupus nephritis patients [23, 24]. Studies also demonstrated that there was an association between FcyRIIb-Ile232 and susceptibility to anti-GBM disease [25]. FcyRIIb promoter variant-386C-120A downregulated the expression of FcyRIIb and greatly related to the chronic inflammatory demyelinating polyneuropathy. Helicobacter pylori infection also downregulated the expression of FcyRIIb and induced idiopathic thrombocytopenic purpura [26]. These studies showed that the FcyRIIb gene polymorphisms were associated with several diseases, especially autoimmune diseases [27, 28].

FcyRIIb gene polymorphisms and periodontitis

Periodontopathic bacteria such as *Porphyromonas gingivalis* are known to affect the local host immunity [29-31]. Indeed, patients with periodontitis displayed significantly higher serum IgG responses to the *P. gingivalis* 40-KDa outer membrane protein (OMP) than those of the healthy group. The serum IgG subclass distribution for patients with periodontitis

Table 1. FcyRIIB gene polymorphisms

	<u> </u>			
Nucleotide	Amino acid	Position	Nucleotide	Position
nt 608 (C \rightarrow T)	Thr 203 \rightarrow Met	Exon4	nt 645+7 (A \rightarrow C)	Intron4
nt 609 (G \rightarrow A)	Thr 203 \rightarrow Thr	Exon4	nt 645+25 (G \rightarrow A)	Intron4
nt 612 (G \rightarrow A)	Leu 204 \rightarrow Leu	Exon4	nt 646-184 (A \rightarrow G)	Intron4
nt 614 (A \rightarrow T)	Tyr 205 \rightarrow Phe	Exon4	nt 646-86 (C \rightarrow T)	Intron4
nt 619 (T \rightarrow G)	Ser 207 \rightarrow Ala	Exon4	nt 759+27 (T \rightarrow G)	Intron5
nt 695 (T \rightarrow C)	Ile 203 \rightarrow Thr	Exon5		

Eleven single-nucleotide polymorphisms (SNPs) were confirmed to be Fc γ RIIB-specific, of these SNPs, three and one SNPs resulted in amino-acid substitutions in exon4 (Thr203-Met, Tyr205-Phe and Ser207-Ala) and exon5 (Ile232-Thr). The other five SNPs were detected in intron4 and 5, leading to no amino-acid substitution.

and healthy individuals was $IgG_1>IgG_2>IgG_3$ for the anti-*P. gingivalis*-40-KDa OMP response [32]. Furthermore, the ability of *Aggregatibacter actinomycetemcomitans*, *Prevotella intermedia* and *Fusobacterium nucleatum* to bind IgG Fc fragments has been demonstrated, on the other hand, the ability of *P. gingivalis* possessed IgG Fc-binding activity has not been observed [33, 34]. Periodontitis is a complex chronic subgingival plaque-induced inflammatory diseases influenced by multiple factors, including genetics, behavior and the environment. Many genetic association studies have been conducted in periodontology [35].

According to the above reports, FcyRIIb gene polymorphisms may play a primary role in periodontitis development, because there are large numbers of FcyRII-bearing B lymphocytes in periodontal lesions. Additionally, to date FcyRIIb is the only known inhibitory receptor in the FcgR family, which is pivotal in the regulation of B cell activation. Yasuda et al. observed a significant difference in the FcyRIIb-232I/T allele (exon5) distribution between the aggressive periodontitis and healthy control groups, with enrichment of 232T in the aggressive periodontitis group. The same report revealed that FcyRIIb-nt646-184A/G allele (intron4) distribution was significantly different between the chronic periodontitis and healthy control groups, with enrichment of nt646-184A in the chronic periodontitis group [5]. These results support the association of FcyRIIb gene polymorphism with periodontitis susceptibility. Additionally, the FcyRIIb-232T allele might be related to the reduced IgG antibody response to P. gingivalis in chronic periodontitis patients [20]. FcyRIIbnt645+25AA carriers with chronic periodontitis

displayed significantly higher mean clinical attachment (CAL) levels and a significantly lower IgG response to *P. gingivalis* sonicate and to the 40-KDa OMP (outer membrane protein) compared with patients with those of Fcγ-RIIb-nt645+GG carriers [7]. These results suggest that the association of the FcγRIIb gene polymorphisms with periodontitis might be related to the lower levels of antibody response to periodontal bac-

teria. Human FcyRIIb suppresses B lymphocytes activation through cross-linking with the B cell receptor via immune complexes. This function of FcyRIIb is essential for the negative regulation of antibody complexes [20]. Higher FcyRIIb expression in subjects caused by FcyRIIb gene polymorphisms might induce a lower IgG level response to periodontal bacteria. The association of FcyRIIb gene polymorphisms with periodontitis susceptibility may be related to inflammation caused by a lower level of production of IgG against periodontal bacteria. The FcyRIIB genetic polymorphism in mouse strains was associated with down-regulation of its expression, possibly contributing to autoimmune diseases susceptibility caused by highaffinity IgG autoantibodies [36]. Unlike other genetic polymorphisms reported in periodontology, most Fcy receptor polymorphisms reported not only have established biological functions but also associate with other autoimmune diseases such as rheumatoid arthritis and systemic lupus erythematosus [35].

Adverse pregnancy outcomes and periodontitis

Adverse pregnancy outcomes are caused by miscarriage, threatened premature labor, preterm birth (with low birth weight), preeclampsia and pregnancy-induced hypertension (PIH), gestational diabetes mellitus, intrauterine growth retardation (IUGR), stillbirth. Associations between preterm birth (with low birth weight) and preeclampsia with periodontitis have been mainly and widely reported [37-42] (**Table 2**). Preterm birth is defined as delivery after week 22 but before week 37 of gestation, while low birth weight is defined as fetal weight <2500 g.

Adverse pregnancy outcomes	Population	Periodontitis	References
Preterm birth (PTB)	UK	+	Siqueira et al., 2007
		+	Piscoya et al., 2012
		+	Lopez et al., 2002
		+	Guimaraes et al., 2010
		+	Lunardelli & Peres, 2005
	Italian	-	Vettore et al., 2008
	French	-	Nabet et al., 2010
	Canadian	-	Wood et al., 2006
	UK	-	Moore et al., 2004
	American	+	Rakoto-Alson et al., 2010
		+	Offenbancher et al., 2006
	Spain	+	Agueda et al., 2008
	Jordanian	+	Habashneh et al., 2012
	American	-	Srinivas et al., 2009
Low birth weight (LBW)	American	+	Offenbacher et al., 1996
	US	+	Gomes-Fillho et al., 2007
	Italian	+	Vettore et al., 2008
	Jordanian	+	Khader et al., 2009
	Spain	-	Agueda et al., 2008
	US	-	Davenport et al., 2002
	German	-	Noack et al., 2005
Preeclampsia (PE)	Turkish	+	Canakci et al., 2004
	Brazil	+	Cota et al., 2006
	French	+	Nabet et al., 2010
	Canadian	-	Taghzouti et al., 2011
	Jordan	-	Khader et al., 2006

Table 2. Associations between adverse pregnancy outcomes and periodontitis in different population

Abbreviation: +, positive association reported; -, negative associationreported.

Preeclampsia is a pregnancy condition characterized by hypertension and proteinuria after 20 weeks of gestation and is one of the leading causes of maternal, fetal and neonatal mortality worldwide. Infections play a significant role in spontaneous preterm labor and birth as well as in related neonatal complications [43]. Furthermore, birth canal infections play an important role in the etiopathogenesis of preterm birth [44]. Systemic maternal infections are hypothesized to raise the risk of placental infection, premature rupture of membranes, premature labor and preterm birth as a result of inflammatory cytokines release and increased prostaglandin production [45].

Periodontitis is an infectious disease caused by the direct effect of periodontopathic bacteria and the accompanied host immune response [46-48], which is associated with an increase in systemic inflammatory cytokine levels. Periodontal organisms have been isolated from the amniotic fluid, suggesting hematogenous spread [49]. Therefore, it is possible that periodontal disease could potentially affect pregnancv outcomes through indirect mechanisms involving inflammatory cytokines or direct translocation of bacteria and its products to the fetoplacental unit. Moreover, periodontal disease is associated with adverse pregnancy outcomes, such as preterm delivery, preeclampsia abortion and stillbirth, low birth weight (LBW) infants and preterm LBW infants [2, 3, 37, 50-53].

Bacteria cause local immune response in periodontal pockets. Proinflammatory cytokines and IgG against periodontal bacteria released from immune-related host cells can enter the bloodstre-

am and increase prostaglandin and cytokine levels, which may induce adverse pregnancy outcomes [54-57]. Previous studies showed that maternal subgingival A. actinomycetemcomitans DNA levels were associated with preterm birth and preeclampsia [6, 10]. A prospective cohort study of 13 circulation cytokines mid-pregnancy revealed significant associations between IL-1B, IL-2, IL-12, interferon-y (IFNG), IL-4, IL-6 and transforming growth factor- β levels and preterm delivery at <35 weeks with histological chorioamnionitis [58]. This study showed that both periodontitis and preterm birth were caused by infection and aggravatedbyhost-inducedinflammation.Additionally, a lower level of serum IgG against periodontopathic bacteria was more closely associated with preterm birth compared with term birth [59].



Figure 2. The mechanism of associations between $Fc\gamma RIIB$ gene polymorphisms, periodontitis, and adverse pregnancy outcomes. The development of adverse pregnancy outcomes might be attributable to $Fc\gamma RIIB$ gene polymorphisms-associated inflammation caused by periodontal infection through up-regulated proinflammatory cytokines.

A meta-analysis by Khader and Ta'ani indicated that periodontal diseases in pregnant women significantly increased the risk of subsequent preterm birth or low birth weight [55]. In contrast, Michalowicz et al. reported that treatment of periodontitis in pregnant women did not significantly alter rates of preterm birth and low birth weight [60]. Vettore et al. compared periodontal clinical measurements and the levels and proportions of 39 bacterial species in subgingival biofilm samples in women with preterm and non-preterm births and concluded that maternal periodontal microbiota and periodontal disease clinical characteristics were not associated with preterm birth [61]. In our previous studies, a significant association between clinical periodontal parameters and adverse pregnancy outcomes was not observed. It suggested that subgingival periodontal bacteria were associated with adverse pregnancy outcomes susceptibility [8, 10].

Association of FcγRIIB gene polymorphisms and periodontitis with adverse pregnancy outcomes

As infection caused by periodontal bacteria in periodontal pockets can alter serum proinflammatory cytokine levels in pregnant women, infection may induce chorioamnionitis and result in adverse pregnancy outcomes. Significant associations between levels of proinflammatory cytokines, such as IL-1 β , IL-2, IL-12, interferon- γ (IFNG), IL-4, IL-6 and transforming growth factor- β and preterm delivery at <35 weeks with histological chorioamnionitis were identified [58]. Low IgG production against the periodontal bacterium *P. gingivalis* in early pregnancy was associated with intrauterine growth retardation and some instances of preterm birth. Furthermore, lower serum IgG1 levels against anti-*P. gingivalis* OMP and higher C-reactive protein (CRP) levels were associated with preterm birth with chorioamnionitis [59]. Maternal IgG against bacterial antigens are transported into fetal blood using endosomes and Fc receptors.

In previous reports, FcyRIIb-232I/T and FcyR-IIb-nt645+25A/G polymorphisms were associated with periodontitis. The association of the 232T allele and nt645+25AA genotype carriers with periodontitis might be related to the lower levels of IgG antibody response to P. gingivalis [7, 20]. Moreover, the FcyRIIb-nt645+ 25A/G polymorphism has been suggested to be a susceptibility factor for adverse pregnancy outcomes, such as preterm birth or preeclampsia [6, 8]. FcyRIIb protein expression on the cell surface in peripheral B lymphocytes was higher in healthy donors with the FcyRIIb-nt645+25AA genotype than that of FcyRIIb-nt645+25GG genotype carriers [7]. Therefore, we assume that adverse pregnancy outcomes might be attributed to FcyRIIb gene polymorphism-associated inflammation caused by periodontal infection through up-regulated proinflammatory cytokines levels (Figure 2). However, no significant association between clinical periodontal parameters and adverse pregnancy outcomes has been found. Rather only subgingival periodontal bacteria DNA levels were associated with adverse pregnancy outcomes in our previous studies (Table 3) [6, 8, 10].

Summary of previous studies

FcyRIIb gene polymorphisms were significantly associated with preterm birth, preeclampsia, pregnancy-induced hypertension, periodontitis and antibacterial IgG levels in previous reports. Among 22 immunnoregulatory polymorphisms, only IL-6 and Fc α R polymorphisms were significantly associated with preterm birth after adjustment for confounders.

We did not identify significant associations between adverse pregnancy outcomes and periodontitis or any clinical periodontal parameters, inconsistent with the results from most **Table 3.** Associations between gene polymorphisms, peri-
odontitis and adverse pregnancy outcomes in our previous
studies

	Preterm Birth	Preeclampsia
Periodontitis (+)	Ν	Ν
Mean Clinical Attachment Loss	Ν	Ν
Subgingival bacterial level (log per 10 µL)		
Actinobacillus actinomycetemcomitans	Υ	Y
Porphyromonas gingivalis	Ν	Ν
Serum IgG antibody level		
A. actinomycetemcomitans	Ν	Ν
P. gingivalis	Υ	Ν
FcyRIIA polymorphism	Ν	Ν
FcyRIIB polymorphism	Υ	Y
FcyRIIIB polymorphism	Ν	Y
IL-6 gene polymorphism	Υ	NT
FcαR polymorphism	Y	NT

N-no significant association; Y-Significant association; NT-Not tested.

previous case-control studies. These discrepancies may be explained by differing parities and periodontitis severity. Mean CAL (2.42 mm) in the preterm birth group in our study was lower than that in previous studies (3.00 mm) [62]. The periodontitis definition criterion in our study was 60% of sites with a clinical attachment level of \geq 3 mm. In contrast, in most other studies, they adopted one or more sites of pocket depth \geq 4 mm and attachment loss \geq 3 mm in the same site, or one or more sites of attachment loss \geq 4 mm [63, 64].

However, subgingival *A. actinomycetemcomitans* DNA level was associated with preterm birth and preeclampsia [6, 10]. Moreover, significantly lower serum anti-*P. gingivalis* OMP IgG1 and higher CRP were observed in sera obtained during the first trimester from women who delivered preterm with chorioamnionitis [58].

Previously, we demonstrated the association of $Fc\gamma RIIb$ gene polymorphisms with periodontitis susceptibility in patients with chronic periodontitis [7, 20]. However, periodontal conditions and subject age differed between the previous studies with chronic periodontitis patients and the present studies with pregnant women.

Adverse pregnancy outcomes associated with periodontitis may be caused by translocation of

periodontal bacteria and/or its products to the fetoplacental unit which induced proinflammatory cytokines up-regulation [49]. The association of $Fc\gamma$ RIIb gene polymorphisms with preterm birth with low birth weight and preeclampsia might result from lower immune protection against bacterial infection and subsequent up-regulation of proinflammatory cytokines and CRP. Further studies should be undertaken to confirm this hypothesis.

Acknowledgements

This work was supported by Grants-in-Aid for scientific Research (19592385) and for Challenging Exploratory Research (21659480) from the Japan Society for Promo-

tion of Science (JSPS), Tokyo, Japan. This study was approved by the committee of the Fourth Hospital of Harbin Medical University.

Disclosure of conflict of interest

None.

Address correspondence to: Yanming Wang, Department of Stomatology, The Fourth Hospital of Harbin Medical University, 37 Yiyuan Street, Nangang District, Harbin 150001, Heilongjiang Province, China. Tel: +86-451-82576565; Fax: +86-451-82576566; E-mail: y_f_081@sina.com

References

- [1] Offenbacher S, Lieff S, Boggess KA, Murtha AP, Madianos PN, Champagne CM, McKaig RG, Jared HL, Mauriello SM, Auten RL Jr, Herbert WN, Beck JD. Maternal periodontitis and prematurity. Part I: obstetric outcome of prematurity and growth restriction. Ann Periodontol 2001; 6: 164-74.
- [2] Govindaraju P, Venugopal S, Shivakumar MA, Sethuraman S, Ramaiah SK, Mukundan S. Maternal periodontal disease and preterm birth: a case-control study. J Indian Soc Periodontol 2015; 19: 512-5.
- [3] Reza Karimi M, Hamissi JH, Naeini SR, Karimi M. The relationship between maternal periodontal status of and preterm and low birth weight infants in Iran: a case control study. Glob J Health Sci 2016; 8: 184-8.

- [4] Ide M, Papapanou PN. Epidemiology of association between maternal periodontal disease and adverse pregnancy outcomes-systematic review. J Periodontol 2013; 84: S181-94.
- [5] Yasuda K, Sugita N, Kobayashi T, Yamamoto K, Yoshie H. FcyRIIB gene polymorphisms in Japanese periodontitis patients. Genes Immun 2003; 4: 541-6.
- [6] Iwanaga R, Sugita N, Hirano E, Sasahara J, Kikuchi A, Tanaka K, Yoshie H. FcγRIIB polymorphisms, periodontitis and preterm birth in Japanese pregnant women. J Periodontal Res 2011; 46: 292-302.
- [7] Sugita N, Iwanaga R, Kobayashi T, Yoshie H. Association of the FcγRIIB-nt645+25A/G polymorphism with the expression level of the FcγRIIB receptor, the antibody response to Porphyromonas gingivalis and the severity of periodontitis. J Periodontal Res 2012; 47: 105-13.
- [8] Wang Y, Sugita N, Kikuchi A, Iwanaga R, Hirano E, Shimada Y, Sasahara J, Tanaka K, Yoshie H. FcyRIIB-nt645+25A/G gene polymorphism and periodontitis in Japanese women with preeclampsia. Int J Immunogenet 2012; 39: 492-500.
- [9] Sugita N, Kobayashi T, Kikuchi A, Shimada Y, Hirano E, Sasahara J, Tanaka K, Yoshie H. Immunoregulatory gene polymorphisms in Japanese women with preterm births and periodontitis. J Reprod Immunol 2012; 93: 94-101.
- [10] Hirano E, Sugita N, Kikuchi A, Shimada Y, Sasahara J, Iwanaga R, Tanaka K, Yoshie H. The association of aggregatibacter actinomycetemcomitans with preeclampsia in a subset of Japanese pregnant women. J Clin Periodontol 2012; 39: 229-38.
- [11] Qiu WQ, de Bruin D, Brownstein BH, Pearse R, Ravetch JV. Organization of the human and mouse low-affinity FcgammaR genes: duplication and recombination. Science 1990; 248: 732-5.
- [12] Muta T, Kurosaki T, Misulovin Z, Sanchez M, Nussenzweig MC, Ravetch JV. A 13-amino-acid motif in the cytoplasmic domain of FcγRIIB modulates B-cell receptor signalling. Nature 1994; 368: 70-3.
- [13] Budde P, Bewarder N, Weinrich V, Schulzeck O, Frey J. Tyrosine-containing sequence motifs of the human immunoglobulin G receptors FcyRIIB1 and FcyRIIB2 essential for endocytosis and regulation of calcium flux in B cells. J Biol Chem 1994; 269: 30636-44.
- [14] D'Ambrosio D, Hippen KL, Minskoff SA, Mellman I, Pani G, Siminovitch KA, Cambier JC. Recruitment and activation of PTP1C in negative regulation of antigen receptor signaling by FcγRIIB1. Science 1995; 268: 293-7.

- [15] Bolland S, Ravetch JV. Spontaneous autoimmune disease in FcγRIIB-deficient mice results from strain-specific epistasis. Immunity 2000; 13: 277-85.
- [16] Rudge EU, Cutler AJ, Pritchard NR, Smith KG. Interleukin 4 reduces expression of inhibitory receptors on B cells and abolishes CD22 and Fc gamma RII-mediated B cell suppression. J Exp Med 2002; 195: 1079-85.
- [17] Nimmerjahn F, Ravetch JV. Divergent immunoglobulin g subclass activity through selective Fc receptor binding. Science 2005; 310: 1510-2.
- [18] Tridandapani S, Wardrop R, Baran CP, Wang Y, Opalek JM, Caligiuri MA, Marsh CB. TGF-beta 1 suppresses [correction of supresses] myeloid Fc gamma receptor function by regulating the expression and function of the common gamma-subunit. J Immunol 2003; 170: 4572-7.
- [19] Guriec N, Daniel C, Le Ster K, Hardy E, Berthou C. Cytokine-regulated expression and inhibitory function of FcγRIIB1 and -B2 receptors in human dendritic cells. J Leukoc Biol 2006; 79: 59-70.
- [20] Honma Y, Sugita N, Kobayashi T, Abiko Y, Yoshie H. Lower antibody response to Porphyromonas gingivalis associated with immunoglobulin G Fcgamma receptor IIB polymorphism. J Periodontal Res 2008; 43: 706-11.
- [21] Mallery DL, McEwan WA, Bidgood SR, Towers GJ, Johnson CM, James LC. Antibodies mediate intracellular immunity through tripartite motif-containing 21 (TRIM21). Proc Natl Acad Sci U S A 2010; 107: 19985-90.
- [22] Anthony RM, Ravetch JV. A novel role for the IgG Fc glycan: the anti-inflammatory activity of sialylated IgG Fcs. J Clin Immunol 2010; 30 Suppl 1: S9-14.
- [23] Zidan HE, Sabbah NA, Hagrass HA, Tantawy EA, El-Shahawy EE, Nageeb GS, Abdul-Sattar AB. Association of FcyRIIB and FcyRIIA R131H gene polymorphisms with renal involvement in Egyptian systemic lupus erythematosus patients. Mol Biol Rep 2014; 41: 733-9.
- [24] Pradhan V, Patwardhan M, Nadkarni A, Ghosh K. FcγRIIB gene polymorphisms in Indian systemic lupus erythematosus (SLE) patients. Indian J Med Res 2011; 134: 181-5.
- [25] Zhou XJ, Lv JC, Bu DF, Yu L, Yang YR, Zhao J, Cui Z, Yang R, Zhao MH, Zhang H. Copy number variation of FCGR3A rather than FCGR3B and FCGR2B is associated with susceptibility to anti-GBM disease. Int Immunol 2010; 22: 45-51.
- [26] Satoh T, Miyazaki K, Shimohira A, Amano N, Okazaki Y, Nishimoto T, Akahoshi T, Munekata S, Kanoh Y, Ikeda Y, Higashihara M, Takahashi S, Kuwana M. Fcgamma receptor IIB gene polymorphism in adult Japanese patients with

primary immune thrombocytopenia. Blood 2013; 122: 1991-2.

- [27] Wu J, Lin R, Huang J, Guan W, Oetting WS, Sriramarao P, Blumenthal MN. Functional Fcgamma receptor polymorphisms are associated with human allergy. PLoS One 2014; 9: e89196.
- [28] Espeli M, Smith KG, Clatworthy MR. FcγRIIB and autoimmunity. Immunol Rev 2016; 269: 194-211.
- [29] Olsen I, Yilmaz O. Modulation of inflammasome activity by Porphyromonas gingivalis in periodontitis and associated systemic diseases. J Oral Microbiol 2016; 8: 30385.
- [30] Silva N, Abusleme L, Bravo D, Dutzan N, Garcia-Sesnich J, Vernal R, Hernandez M, Gamonal J. Host response mechanisms in periodontal diseases. J Appl Oral Sci 2015; 23: 329-55.
- [31] Mysak J, Podzimek S, Sommerova P, Lyuya-Mi Y, Bartova J, Janatova T, Prochazkova J, Duskova J. Porphyromonas gingivalis: major periodontopathic pathogen overview. J Immunol Res 2014; 2014: 476068.
- [32] Kobayashi T, Kaneko S, Tahara T, Hayakawa M, Abiko Y, Yoshie H. Antibody responses to Porphyromonas gingivalis hemagglutinin A and outer membrane protein in chronic periodontitis. J Periodontol 2006; 77: 364-9.
- [33] Guo M, Han YW, Sharma A, De Nardin E. Identification and characterization of human immunoglobulin G Fc receptors of fusobacterium nucleatum. Oral Microbiol Immunol 2000; 15: 119-23.
- [34] Grenier D, Michaud J. Demonstration of human immunoglobulin G Fc-binding activity in oral bacteria. Clin Diagn Lab Immunol 1994; 1: 247-9.
- [35] Chai L, Song YQ, Leung WK. Genetic polymorphism studies in periodontitis and Fcgamma receptors. J Periodontal Res 2012; 47: 273-85.
- [36] Jiang Y, Hirose S, Abe M, Sanokawa-Akakura R, Ohtsuji M, Mi X, Li N, Xiu Y, Zhang D, Shirai J, Hamano Y, Fujii H, Shirai T. Polymorphisms in IgG Fc receptor IIB regulatory regions associated with autoimmune susceptibility. Immunogenetics 2000; 51: 429-35.
- [37] Basha S, Shivalinga Swamy H, Noor Mohamed R. Maternal periodontitis as a possible risk factor for preterm birth and low birth weight--a prospective study. Oral Health Prev Dent 2015; 13: 537-44.
- [38] Srinivas SK, Sammel MD, Stamilio DM, Clothier B, Jeffcoat MK, Parry S, Macones GA, Elovitz MA, Metlay J. Periodontal disease and adverse pregnancy outcomes: is there an association? Am J Obstet Gynecol 2009; 200: 497, e1-8.
- [39] Agueda A, Ramon JM, Manau C, Guerrero A, Echeverria JJ. Periodontal disease as a risk

factor for adverse pregnancy outcomes: a prospective cohort study. J Clin Periodontol 2008; 35: 16-22.

- [40] Papapanou PN, Behle JH, Kebschull M, Celenti R, Wolf DL, Handfield M, Pavlidis P, Demmer RT. Subgingival bacterial colonization profiles correlate with gingival tissue gene expression. BMC Microbiol 2009; 9: 221.
- [41] Radnai M, Gorzo I, Urban E, Eller J, Novak T, Pal A. Possible association between mother's periodontal status and preterm delivery. J Clin Periodontol 2006; 33: 791-6.
- [42] Rylev M, Kilian M. Prevalence and distribution of principal periodontal pathogens worldwide. J Clin Periodontol 2008; 35: 346-61.
- [43] Pararas MV, Skevaki CL, Kafetzis DA. Preterm birth due to maternal infection: Causative pathogens and modes of prevention. Eur J Clin Microbiol Infect Dis 2006; 25: 562-9.
- [44] Koucky M, Germanova A, Hajek Z, Parizek A, Kalousova M, Kopecky P. Pathophysiology of preterm labour. Prague Med Rep 2009; 110: 13-24.
- [45] Goldenberg RL, Hauth JC, Andrews WW. Intrauterine infection and preterm delivery. N Engl J Med 2000; 342: 1500-7.
- [46] Slots J, Ting M. Actinobacillus actinomycetemcomitans and porphyromonas gingivalis in human periodontal disease: occurrence and treatment. Periodontol 2000 1999; 20: 82-121.
- [47] Gustafsson A, Ito H, Asman B, Bergstrom K. Hyper-reactive mononuclear cells and neutrophils in chronic periodontitis. J Clin Periodontol 2006; 33: 126-9.
- [48] Surna A, Kubilius R, Sakalauskiene J, Vitkauskiene A, Jonaitis J, Saferis V, Gleiznys A. Lysozyme and microbiota in relation to gingivitis and periodontitis. Med Sci Monit 2009; 15: CR66-73.
- [49] Hillier SL, Krohn MA, Cassen E, Easterling TR, Rabe LK, Eschenbach DA. The role of bacterial vaginosis and vaginal bacteria in amniotic fluid infection in women in preterm labor with intact fetal membranes. Clin Infect Dis 1995; 20 Suppl 2: S276-8.
- [50] Parihar AS, Katoch V, Rajguru SA, Rajpoot N, Singh P, Wakhle S. Periodontal disease: a possible risk-factor for adverse pregnancy outcome. J Int Oral Health 2015; 7: 137-42.
- [51] Harjunmaa U, Jarnstedt J, Alho L, Dewey KG, Cheung YB, Deitchler M, Ashorn U, Maleta K, Klein NJ, Ashorn P. Association between maternal dental periapical infections and pregnancy outcomes: results from a cross-sectional study in Malawi. Trop Med Int Health 2015; 20: 1549-1558.
- [52] Desai K, Desai P, Duseja S, Kumar S, Mahendra J, Duseja S. Significance of maternal peri-

odontal health in preeclampsia. J Int Soc Prev Community Dent 2015; 5: 103-7.

- [53] Sitholimela CS, Shangase LS. The association between periodontitis and pre-term birth and/ or low birth weight: a literature review. SADJ 2013; 68: 162-6.
- [54] Scannapieco FA, Wang B, Shiau HJ. Oral bacteria and respiratory infection: effects on respiratory pathogen adhesion and epithelial cell proinflammatory cytokine production. Ann Periodontol 2001; 6: 78-86.
- [55] Khader YS, Ta'ani Q. Periodontal diseases and the risk of preterm birth and low birth weight: a meta-analysis. J Periodontol 2005; 76: 161-5.
- [56] Rosa MI, Pires PD, Medeiros LR, Edelweiss MI, Martinez-Mesa J. Periodontal disease treatment and risk of preterm birth: a systematic review and meta-analysis. Cad Saude Publica 2012; 28: 1823-33.
- [57] Schwendicke F, Karimbux N, Allareddy V, Gluud C. Periodontal treatment for preventing adverse pregnancy outcomes: a meta- and trial sequential analysis. PLoS One 2015; 10: e0129060.
- [58] Gargano JW, Holzman C, Senagore P, Thorsen P, Skogstrand K, Hougaard DM, Rahbar MH, Chung H. Mid-pregnancy circulating cytokine levels, histologic chorioamnionitis and spontaneous preterm birth. J Reprod Immunol 2008; 79: 100-10.

- [59] Sasahara J, Kikuchi A, Takakuwa K, Sugita N, Abiko Y, Yoshie H, Tanaka K. Antibody responses to porphyromonas gingivalis outer membrane protein in the first trimester. Aust N Z J Obstet Gynaecol 2009; 49: 137-41.
- [60] Michalowicz BS, Hodges JS, DiAngelis AJ, Lupo VR, Novak MJ, Ferguson JE, Buchanan W, Bofill J, Papapanou PN, Mitchell DA, Matseoane S, Tschida PA; Study OPT. Treatment of periodontal disease and the risk of preterm birth. N Engl J Med 2006; 355: 1885-94.
- [61] Vettore MV, Leao AT, Leal Mdo C, Feres M, Sheiham A. The relationship between periodontal disease and preterm low birthweight: clinical and microbiological results. J Periodontal Res 2008; 43: 615-26.
- [62] Offenbacher S, Katz V, Fertik G, Collins J, Boyd D, Maynor G, McKaig R, Beck J. Periodontal infection as a possible risk factor for preterm low birth weight. J Periodontol 1996; 67: 1103-13.
- [63] Giannella L, Giulini S, Cerami LB, La Marca A, Forabosco A, Volpe A. Periodontal disease and nitric oxide levels in low risk women with preterm labor. Eur J Obstet Gynecol Reprod Biol 2011; 158: 47-51.
- [64] Nabet C, Lelong N, Colombier ML, Sixou M, Musset AM, Goffinet F, Kaminski M; Epipap Group. Maternal periodontitis and the causes of preterm birth: the case-control Epipap study. J Clin Periodontol 2010; 37: 37-45.