Original Article Circulating miR-21 and miR-423-5p as biomarkers for heart failure in heart valve disease patients

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Abstract: Objective: The potential diagnostic value of circulating miRNAs in heart failure (HF) due to heart valve disease (VHD) remains elusive. The purpose of this study was to investigate the potential value of serum miR-21 and miR-423-5p for HF diagnosis. Methods: Serum samples from 60 VHD patients with HF were collected for investigation, and 60 healthy subjects' samples were used as the control. The expression level of serum miR-21 and miR-423-5p was measured by QRT-PCR. ROC curve analysis and Pearson correlation analysis were performed to determine the diagnostic value. Results: The expression of miR-21 and miR-423-5p in sera was markedly increased in VHD patients than normal control (both P < 0.05). Besides, serum miR-21 and miR-423-5p showed independent and effective diagnostic utilities for HF due to VHD. The AUC of serum miR-423-5p was 0.805 for distinguishing the VHD patients with pulmonary hypertension (PH). Conclusions: Serum miR-21 and miR-423-5p showed potential diagnostic values on HF due to VHD. Moreover, serum miR-423-5p could function as an effective biomarker for diagnostic values on HF due to VHD.

Keywords: Circulating microRNA, miR-21, miR-423-5p, valvular heart disease, heart failure, ROC curve

Introduction

As a debilitating chronic disease, heart failure (HF) has become one of the most tough health problems worldwide. The prevalence of HF is expected to accelerate in the next couples of years mainly due to the unhealthy habits established in modern society [1, 2]. HF is often characterized with ventricular filling and decreased ejection fraction [3, 4]. At the late stage or severe state of multiple vascular cardiac diseases, patients were often suffered from HF [5]. In order to diagnose HF more accurately and precisely, significant efforts have been made to indentify a series of biomarkers for HF diagnosis in clinic. The B-type natriuretic peptide (BNP) and N-terminal pro-brain natriuretic peptide (NT-proBNP) were considered as promising biomarkers for HF diagnosis [6]. Also, BNP and NT-proBNP can serve as adjunctive biomarkers for evaluating the severity and defining the progression of HF in congenital cardiac diseases [7, 8]. Although great values were implied for BNP and NT-proBNP in the prediction of HF, there are still some limitations and the results

from several studies remains controversial. It was reported that single BNP measurement was not enough to determine the degree of congestion in acute systolic HF [9]. Recent study showed an inverse association between BNP and NT-proBNP levels and body mass index, indicating the limitation in diagnosis of obesity patients with HF [10]. Thus, novel non-invasive biomarkers were required for noninvasive diagnosis on HF.

Valvular heart disease (VHD) has become an increasingly common type in patients who suffered from age-related cardiac disorders [11]. Although the prevalence of VHD gradually decreased, the incidence of VHD was still estimated at approximately 2.5% in industrialized countries [12]. With the progression of VHD, the endocardiosis and cardiac dysfunction would eventually lead to HF [13]. We discovered that after interval treatments on patients with VHD, the accuracy and specificity of NT-proBNP on evaluating the severity of VHD reduced significantly. Moreover, to date, the diagnostic values of biomarkers for VHD patients with pulmonary

hypertension (PH) still remain limited [14, 15]. Hence, we aimed to find alternative biomarkers for VHD diagnosis in the present study.

MicroRNAs (miRNAs), as a group of non-coding RNAs with 21-23 nucleotides in length, have been proven to serve critical roles in the process of various human cardiac diseases, such as myocardial infarction, chronic myocarditis, as well as HF [16-18]. Many reports revealed that aberrant expression patterns of miRNAs were associated with cardiac dysfunction, indicating that miRNAs might be involved in the mechanisms of heart diseases and qualified to be promosing biomarkers for HF [19-22]. Multiple miRNAs in biofluids were recognized as circulating biomarkers for HF [23, 24]. Wong et al. discovered that the specific miRNA panels enjoyed great discrimination power than NT-proBNP in HF with reduced and preserved left ventricular ejection fraction [25]. Moreover, the plasma miR-30d level was identified to be closely associated with response to cardiac resynchronization therapy (CRT) in HF patients with dyssynchrony [26]. The article of Lai et al. also demonstrated circulating miRNAs could be biomarkers in the early stage of HF in Chinese population [27]. The miRNAs in VHD patients were also identified as potential biomarkers for disease progression, while the diagnostic value remained unclear [28].

A study concerning Dachshunds suggested that miR-21 showed a trend of down-regulation in the mild to moderate VHD-HF group [29]. Moreover, miR-423-5p expression was increased in HF patients than control despite the body mass index, indicating it might be a convincing predictor for HF [30]. Meanwhile, through our clinical validations, we predicted miR-423-5p expression might be associated with VHD failure patients with HF. Therefore, this study intended to confirm the diagnostic value of miR-21 and miR-423-5p in HF due to VHD.

Materials and methods

Subjects

Sixty VHD patients with HF and sixty healthy individuals without significant cardiovascular disease from Sichuan Provincial People's Hospital (Chengdu, China) between August 2014 and April 2016 were recruited in the present study. The disease history of VHD patients was documented in hospitalized medical records.

The assistance examinations such as general 12-leads ECG (Electrocardiograph), TTE (transthoracic echocardiography), and detection of expression levels of specific indicators including NT-proBNP, hsCRP (hyper-sensitive C-reactive protein) as well as physical examinations were conducted on participants. The patients who was over 50 years old and suspected to be suffered from coronary diseases were underwent coronary angiography. The state of HF was clarified according to NYHA (New York Heart Association) class [31, 32]. The patients included in this research should meet the following criterion: (1) age over 18 years old; (2) confirmed to be suffered from VHD through the above gold examinations; (3) owned the definite symptoms of HF, which were in accordance with the NYHA II-IV class diagnosis criterion. The exclusion criterion were listed below: (1) patients who had the history of myocardial infarction or underwent acute HF, or coronary artery stenosis area over 75% by coronary angiography, or treated stent implantation before: (2) patients with primary pulmonary diseases; (3) patients with hyperthyroidism, hypothyroidism, and chronic anemia; (4) who suffered from primary or secondary hypertension. The serum samples from 60 healthy subjects were also enrolled as control. Furthermore, patients were diagnosed with PA according to the updated WHO clinical classification [33], inclusion criteria including mean pulmonary artery pressure $(mPAP) \ge 25 mmHg$, pulmonary wedge pressure $\leq 15 \text{ mmHg}$.

For each participant, 5 ml fasting blood was withdrawn in ethylenediaminetetraacetic acidcontaining tubes. The whole blood samples were processed for serum extraction immediately, and the serum samples were stored at -80°C until further processing.

The study was approved by the Ethic Committee of Sichuan Provincial People's Hospital and performed in compliance with the Helsinki Declaration. Written informed consent documents were obtained from all the participants prior to their inclusion.

Cardiac ultrasound examination

LVEDd (left ventricular end-diastolic diameter) and LVEF (left ventricular ejection fraction) were measured based on Simpson biplane method [34]. The max reversal flow velocity of tricuspid valve was detected by Doppler ultrasound at apical four chamber view.

Gene name	Primer sequence (5' to 3')
miR-21	ACACTCCAGCTGGGTAGCTTATCAGACTGAT
	ACTGGTGTCGTGGAGTCG
miR-423-5p	ATGGTTCGTGGGTGAGGGGCAGAGA
	GCGAGAGCAGGGTCCGAGGTATTC
U6	CTCGCTTCGGCAGCACA
	AACGCTTCACGAATTTGCGT

Table 1. The sequences of QRT-PCR primers

 Table 2. Clinical characteristics of participants enrolled in this study

Characteristics	VHD	Control	P value
Case number	60	60	
Age (year)	60.25 ± 12.03	55.92 ± 9.18	0.497
Gender (male/female)	25/35	28/32	0.581
Log (NT-proBNP)	3.89 ± 0.46	1.18 ± 0.51	< 0.001
HsCRP (mg/L)	6.38 ± 2.78	3.19 ± 1.80	< 0.001
LVEDd (mm)	52.14 ± 7.69	50.05 ± 5.21	0.084
LVEF (%)	56.23 ± 11.75	N.A	

NT-proBNP, N-terminal pro-brain natriuretic peptide; HsCRP, High-sensitive C-reactive protein; LVEDd, Left ventricular end-diastolic diameter; LVEF, Left ventricular ejection fraction. N.A = Non-available.

RNA extraction and quantitative reverse transcription-PCR (QRT-PCR)

The RNA from serum samples was isolated using mirVana[™] PARIS miRNA isolation kit (Ambion Inc, Austin TX, USA) according to the protocol. RNA sample concentration and purity were assessed via NanoDrop ND-1000 spectrophotometer (Nanodrop, USA). 40 ng of total RNA was reverse transcribed to cDNA using a TaqMan MicroRNA Reverse Transcription Kit (Applied Biosystems, Foster City, CA, USA). QRT-PCR reactions for miRNAs were performed using the TagMan MicroRNA PCR Kit (Applied Biosystems, Foster City, CA, USA) on an ABI PRISM 7900HT sequence detection system (Applied Biosystems). The sequences of the primers were listed in Table 1. The 2-AACt cycle threshold (Ct) method was used to investigate the relative expression levels of target miRNAs [35, 36], which were normalized to that of U6. Each experiment was repeated triplicate.

Statistical analysis

All statistical analyses were performed with SP-SS 21.0 software (Chicago, Illinois, USA). Data were presented in the form of mean \pm standard deviation (SD). Receiver-operating characteristic (ROC) curves were plotted and the area under the ROC curve (AUC) was calculated to evaluate the diagnostic value of using serum levels of miRNAs. Student's *t*-test or chi-square test was conducted for the comparison between different groups when appropriate. Pearson correlation analysis was used to estimate the association between two parameters. All statistical tests were 2-tailed and *P* values < 0.05 were considered statistically significant.

Results

Clinical characteristics of participants

1 60 VHD patients with HF were included in this study, whose average age was 60.25 \pm 12.03 years, with 25 male and 35 female. Meanwhile, 60 healthy subjects underwent routine physical examinations were set as control, whose average age of 55.92 \pm 9.18 years, with 28 male and 32 female. There was no significant difference of age, gender, and several other factors between VHD group and control group. However, log (NTproBNP) in VHD group (3.89 \pm 0.46) was markedly higher than that in control group (1.18 \pm 0.51) as well as the lovel of HsCPP (6.28 \pm

0.51), as well as the level of HsCRP (6.38 \pm 2.78 mg/L versus 3.19 \pm 1.80 mg/L). More details about the clinical characteristics of participants were documented in **Table 2**.

Overexpression of serum miR-21 and miR-423 in VHD patients

As shown in **Figure 1A**, serum miR-21 was significantly up-regulated in VHD group rather than that in control group (P < 0.05). Similarly, serum miR-423-5p of VHD group expressed significantly higher than that of control group (P < 0.05). According to Pearson correlation analysis, we found that the expression levels of serum miR-21 and miR-423-5p was separately positive correlated with HsCRP level in VHD patients, as demonstrated in **Figure 1B**. Besides, serum miR-21 level was also closely correlated to serum miR-423-5p level in VHD group.

The diagnostic value of serum miR-21 and miR-423-5p for HF due to VHD

The ROC curves of serum miR-21, serum miR-423-5p and NT-proBNP were plotted and pre-



Figure 1. Relative expression of serum miR-21 and miR-423-5p detected by QRT-PCR. A. The significant higher expression level of miR-21 and miR-423-5p in VHD group rather than control group (all P < 0.05). B. The positive correlation among miR-21, miR-423-5p, and HsCRP in VHD group.



Figure 2. ROC curves of miR-21, miR-423-5p and NT-proBNP.

sented in **Figure 2**. With the AUC of 0.772, it showed that serum miR-21 had the relative promising diagnosis utility for HF due to VHD, which might function as a supplementary biomarker for HF diagnosis in clinic. Moreover, the

AUC of serum miR-423-5p was 0.965, even higher than NT-proBNP (with the AUC of 0.900), indicating it could serve as an accurate diagnosis biomarker for HF in VHD patients.

Serum miR-423-5p could determine VHD with PH

Interestingly, as shown in **Figure 3A**, we observed that serum miR-423-5p had probable higher levels in VHD patients with PH (pulmonary hypertension). Hence, the VHD patients were allocated into two groups based on whether suffered from PH. The clinical characteristics of VHD with PH or non-PH patients were presented in **Table 3**. It

turned out that serum miR-423-5p level in VHD with PH group was significantly higher than that in VHD with non-PH group (P < 0.05), while no significant difference was found on serum miR-21 and NT-proBNP (which did not show in



Figure 3. Relationship between miR-423-5p and VHD patients with PH. A. The level of miR-423-5p in VHD patients with PH was significantly higher than that in VHD with non-PH group, P < 0.05; B. The ROC curve of miR-423-5p diagnosing VHD with PH.

the article). Next, the ROC curve of serum miR-423-5p on determining VHD with PH was performed. The AUC of serum miR-423-5p on VHD with PH was 0.805 (**Figure 3B**); indicating serum miR-423-5p could determine VHD with or without PH.

Discussion

MiRNAs, as a set of conserved non-coding small RNAs, could regulate targets genes posttranscription involved in the process of cell proliferation, differentiation, apoptosis, and angiogenesis by directly targeting specific mRNAs. After miRNA expression in serum was discov-

ered, multiple studies clarified miRNA could be transferred through cell in various ways, and manage to avoid the degradation of RNase by interacting with other medium, ending with stably exiting in extracellular fluid, including serum, plasma, urine, salvia, sweat and even tear [37, 38]. These findings provided sufficient evidences for circulating miRNAs detection in clinical application. Compared with miRNAs from tissue samples, the circulating miRNAs are easily to acquire, convenient to be measured repeatedly, as well as have the advantages of great stability and excellent sensitivity [39]. Recently, the circulating miRNAs have been reported as important biomarkers for various cardiac diseases [40]. The potential diagnostic utility of circulating miRNAs (miR-125a-5p, -190a, -550a-5p, and -638) was identified for discriminating HF with reduced left ventricular ejection fraction [25]. Besides, the diagnostic and prognostic value of circulating miRNAs in HF was detected through different ways [41, 42]. However, the diagnostic value of miRNAs in HF with VHD was poorly investigated.

MiR-21 could promote cardiac fibrosis by regulating Bcl-2, resulting in the development of HF [43]. On one hand, studies showed miR-21 was overexpressed in cardiac myofibroblasts and deficient myocardial cells [44, 45]. On the other hand, Sayed et al. discovered miR-21 as a downstream effector of Akt mediating the suppression of myocaidial cells apoptosis, which means the overexpression of miR-21 may also alleviate HF [46]. The role of miR-21 in HF seems contradictory, which might be explained that the expression of miR-21 was associated with the progression of human cardiac diseases. Other study suggested that miR-21 repressed PDCD4 to enhance valve cell migra-

Characteristics	PH	non-PH	P value
Case number	25	35	
Age (year)	60.17 ± 12.17	60.33 ± 12.11	0.984
Gender (male/female)	11/14	14/21	0.757
Log (NT-proBNP)	3.83 ± 0.45	3.95 ± 0.47	0.339
HsCRP (mg/L)	6.09 ± 2.75	6.67 ± 2.77	0.441
LVEDd (mm)	52.14 ± 7.69	50.05 ± 5.21	0.237
LVEF (%)	55.60 ± 12.15	56.85 ± 11.51	0.697
Serum miR-21	1.66 ± 0.58	1.72 ± 0.84	0.766
Serum miR-423-5p	1.26 ± 0.57	0.64 ± 0.33	< 0.001

Table 3. Clinical characteristics of VHD patients with PH and with non-PH $% \left({{\mathbf{T}_{\mathrm{A}}}^{\mathrm{T}}} \right)$

NT-proBNP, N-terminal pro-brain natriuretic peptide; HsCRP, High-sensitive C-reactive protein; LVEDd, Left ventricular end-diastolic diameter; LVEF, Left ventricular ejection fraction.

tion during cardiac valvulogenesis, indicating the potential association between miR-21 and VHD [47]. In the present study, the expression level of serum miR-21 in VHD group was significantly higher than that of control group. It revealed that serum miR-21 might have a potential diagnostic value for HF in VHD patients. Although the predicting efficacy of serum miR-21 was not very sufficient, it could serve as an effective combined indicator with serum miR-423-5p.

The role of miR-423-5p was also investigated in multiple studies. Although Bauter et al. found circulating level of miR-423-5p was not a useful biomarker for indicating left ventricular remodeling after myocardial infarction, other article showed miR-423-5p in plasma owned great prognostic value of acute HF [48, 49]. While miR-423-5p was enriched in pericardial fluid, whose expression pattern was obviously different and independent compared to others circulating microRNAs such as miR-133a, miR-126, and miR-92a, indicating miR-423-5p may serve as a good biomarker for HF due to VHD [50]. Our study revealed that serum miR-423 in VHD group expressed markedly higher than that in control group. Interestingly, we discovered that miR-423-5p was significantly associated with pulmonary hypertension in VHD patients. As the primary lung diseases patients were excluded from this study, the high pressure in pulmonary artery of participants was caused by VHD, which characterized as PH-LHD (pulmonary hypertension due to left heart disease). According to recent reports, about 2/3 of chronic HF patients were suffered from LHD-PH [51]. It was believed that the severe chronic HF was the common reason leading to LHD-PH, and the incidence of diastolic HF with LHD-PH was higher than systolic HF with LHD-PH [52, 53]. In our research, the VHD patients were divided into two groups based on whether suffered from pulmonary hypertension (PH). The data showed that miR-423-5p in serum had greater sensitivity than NT-proBNP in diagnosis of VHD with PH. Accordingly, serum miR-423-5p could be considered as a promising prognostic biomarker for evaluating VHD with PH.

Some intriguing relationships were also observed in this study. The expression levels of serum miR-21 and miR-423-5p in VHD patients were positively associated with HsCRP, indicating these two miRNAs might be involved in the mechanism of inflammation on HF. Studies manifested that miR-21 may activate NF-kB or target PDCD4, one of the stimuli inflammation proteins, and then regulate the inflammation process [47, 54]. However, the role of miR-423-5p in inflammation process of cardiac diseases needed to be further investigated in future. The positive association between circulating levels of miR-21 and miR-423-5p in VHD patients may also need to be explained. It is speculated that miR-21 and miR-423-5p could probably regulate the cardiac function in cooperation, while it needed further functional study for validation.

In conclusion, we discovered that serum miR-21 and miR-423-5p were up-regulated in VHD patients and could serve as potential and independent biomarkers for HF due to VHD diagnosis. Moreover, miR-423-5p showed great sensitivity for evaluating severity of VHD patients with PH and owned excellent diagnostic utility for VHD with PH prognosis. It is expected more studies would be performed to confirm the results of our research, and the mechanisms of miR-423-5p and miR-21 remained to be clarified in the near future.

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Disclosure of conflict of interest

None.

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References

- Rogers C and Bush N. Heart failure: pathophysiology, diagnosis, medical treatment guidelines, and nursing management. Nurs Clin North Am 2015; 50: 787-799.
- [2] Greenberg B. Gene therapy for heart failure. J Cardiol 2015; 66: 195-200.
- [3] Marangou J and Paul V. Current attitudes on cardiac devices in heart failure: a review. Clin Ther 2015; 37: 2206-2214.
- [4] Ferrari R, Bohm M, Cleland JG, Paulus WJ, Pieske B, Rapezzi C and Tavazzi L. Heart failure with preserved ejection fraction: uncertainties and dilemmas. Eur J Heart Fail 2015; 17: 665-671.
- [5] Rossano JW. Clinical management of patients with acute heart failure. Cardiol Young 2015; 25 Suppl 2: 67-73.
- [6] Don-Wauchope AC and McKelvie RS. Evidence based application of BNP/NT-proBNP testing in heart failure. Clin Biochem 2015; 48: 236-246.
- [7] Cantinotti M, Law Y, Vittorini S, Crocetti M, Marco M, Murzi B and Clerico A. The potential and limitations of plasma BNP measurement in the diagnosis, prognosis, and management of children with heart failure due to congenital cardiac disease: an update. Heart Fail Rev 2014; 19: 727-742.
- [8] Karlstrom P, Dahlstrom U, Boman K and Alehagen U. Responder to BNP-guided treatment in heart failure. The process of defining a responder. Scand Cardiovasc J 2015; 49: 316-324.
- [9] Omar HR and Guglin M. A single BNP measurement in acute heart failure does not reflect the degree of congestion. J Crit Care 2016; 33: 262-265.
- [10] Madamanchi C, Alhosaini H, Sumida A and Runge MS. Obesity and natriuretic peptides, BNP and NT-proBNP: mechanisms and diagnostic implications for heart failure. Int J Cardiol 2014; 176: 611-617.
- [11] Halbach M, Wahlers T, Baldus S and Rudolph V. [Indications for surgery for valvular heart disease]. Dtsch Med Wochenschr 2015; 140: 1733-1740.

- [12] lung B and Vahanian A. Epidemiology of acquired valvular heart disease. Can J Cardiol 2014; 30: 962-970.
- [13] Miller S and Flynn BC. Valvular heart disease and postoperative considerations. Semin Cardiothorac Vasc Anesth 2015; 19: 130-142.
- [14] Magne J, Pibarot P, Sengupta PP, Donal E, Rosenhek R and Lancellotti P. Pulmonary hypertension in valvular disease: a comprehensive review on pathophysiology to therapy from the HAVEC Group. JACC Cardiovasc Imaging 2015; 8: 83-99.
- [15] Guha A, Amione-Guerra J and Park MH. Epidemiology of pulmonary hypertension in left heart disease. Prog Cardiovasc Dis 2016; 59: 3-10.
- [16] Murach KA and McCarthy JJ. MicroRNAs, heart failure, and aging: potential interactions with skeletal muscle. Heart Fail Rev 2017; 22: 209-218.
- [17] Liang J, Bai S, Su L, Li C, Wu J, Xia Z and Xu D. A subset of circulating microRNAs is expressed differently in patients with myocardial infarction. Mol Med Rep 2015; 12: 243-247.
- [18] van den Hoogen P, van den Akker F, Deddens JC and Sluijter JP. Heart failure in chronic myocarditis: a role for microRNAs? Curr Genomics 2015; 16: 88-94.
- [19] Katz MG, Fargnoli AS, Williams RD, Kendle AP, Steuerwald NM and Bridges CR. MiRNAs as potential molecular targets in heart failure. Future Cardiol 2014; 10: 789-800.
- [20] Melman YF, Shah R and Das S. MicroRNAs in heart failure: is the picture becoming less miRky? Circ Heart Fail 2014; 7: 203-214.
- [21] Gidlof O and Erlinge D. MicroRNAs in the failing heart-novel therapeutic targets? Scand Cardiovasc J 2014; 48: 328-334.
- [22] Souza RW, Fernandez GJ, Cunha JP, Piedade WP, Soares LC, Souza PA, de Campos DH, Okoshi K, Cicogna AC, Dal-Pai-Silva M and Carvalho RF. Regulation of cardiac microRNAs induced by aerobic exercise training during heart failure. Am J Physiol Heart Circ Physiol 2015; 309: H1629-1641.
- [23] Vegter EL, van der Meer P, de Windt LJ, Pinto YM and Voors AA. MicroRNAs in heart failure: from biomarker to target for therapy. Eur J Heart Fail 2016; 18: 457-468.
- [24] Ellis KL, Cameron VA, Troughton RW, Frampton CM, Ellmers LJ and Richards AM. Circulating microRNAs as candidate markers to distinguish heart failure in breathless patients. Eur J Heart Fail 2013; 15: 1138-1147.
- [25] Wong LL, Armugam A, Sepramaniam S, Karolina DS, Lim KY, Lim JY, Chong JP, Ng JY, Chen YT, Chan MM, Chen Z, Yeo PS, Ng TP, Ling LH, Sim D, Leong KT, Ong HY, Jaufeerally F, Wong R, Chai P, Low AF, Lam CS, Jeyaseelan K

and Richards AM. Circulating microRNAs in heart failure with reduced and preserved left ventricular ejection fraction. Eur J Heart Fail 2015; 17: 393-404.

- [26] Melman YF, Shah R, Danielson K, Xiao J, Simonson B, Barth A, Chakir K, Lewis GD, Lavender Z, Truong QA, Kleber A, Das R, Rosenzweig A, Wang Y, Kass DA, Singh JP and Das S. Circulating MicroRNA-30d is associated with response to cardiac resynchronization therapy in heart failure and regulates cardiomyocyte apoptosis: a translational pilot study. Circulation 2015; 131: 2202-2216.
- [27] Lai KB, Sanderson JE, Izzat MB and Yu CM. Micro-RNA and mRNA myocardial tissue expression in biopsy specimen from patients with heart failure. Int J Cardiol 2015; 199: 79-83.
- [28] Oury C, Servais L, Bouznad N, Hego A, Nchimi A and Lancellotti P. MicroRNAs in valvular heart diseases: potential role as markers and actors of valvular and cardiac remodeling. Int J Mol Sci 2016; 17.
- [29] Hulanicka M, Garncarz M, Parzeniecka-Jaworska M and Jank M. Plasma miRNAs as potential biomarkers of chronic degenerative valvular disease in Dachshunds. BMC Vet Res 2014; 10: 205.
- [30] Thome JG, Mendoza MR, Cheuiche AV, La Porta VL, Silvello D, Dos Santos KG, Andrades ME, Clausell N, Rohde LE and Biolo A. Circulating microRNAs in obese and lean heart failure patients: a case-control study with computational target prediction analysis. Gene 2015; 574: 1-10.
- [31] Bjork JB, Alton KK, Georgiopoulou VV, Butler J and Kalogeropoulos AP. Defining advanced heart failure: a systematic review of criteria used in clinical trials. J Card Fail 2016; 22: 569-577.
- [32] Miller-Davis C, Marden S and Leidy NK. The New York heart association classes and functional status: what are we really measuring? Heart Lung 2006; 35: 217-224.
- [33] Simonneau G, Gatzoulis MA, Adatia I, Celermajer D, Denton C, Ghofrani A, Gomez Sanchez MA, Krishna Kumar R, Landzberg M, Machado RF, Olschewski H, Robbins IM and Souza R. Updated clinical classification of pulmonary hypertension. J Am Coll Cardiol 2013; 62: D34-41.
- [34] Kammoun I, Zakhama L, Boussaidi H, Mimouni M, Marrakchi S, Slama I, Naccache S, Herbegue B, Ibn El Hadj Z, Boussabah E, Jebri F, Thameur M, Addad F, Ben Youssef S and Kachboura S. [Evaluation of left ventricular function by systolic time intervals]. Tunis Med 2014; 92: 752-755.
- [35] Schmittgen TD and Livak KJ. Analyzing realtime PCR data by the comparative C(T) method. Nat Protoc 2008; 3: 1101-1108.

- [36] Livak KJ and Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. Methods 2001; 25: 402-408.
- [37] Lv Y, Qi R, Xu J, Di Z, Zheng H, Huo W, Zhang L, Chen H and Gao X. Profiling of serum and urinary microRNAs in children with atopic dermatitis. PLoS One 2014; 9: e115448.
- [38] Brown JN, Brewer HM, Nicora CD, Weitz KK, Morris MJ, Skabelund AJ, Adkins JN, Smith RD, Cho JH and Gelinas R. Protein and microRNA biomarkers from lavage, urine, and serum in military personnel evaluated for dyspnea. BMC Med Genomics 2014; 7: 58.
- [39] Oliveira-Carvalho V, da Silva MM, Guimaraes GV, Bacal F and Bocchi EA. MicroRNAs: new players in heart failure. Mol Biol Rep 2013; 40: 2663-2670.
- [40] Min PK and Chan SY. The biology of circulating microRNAs in cardiovascular disease. Eur J Clin Invest 2015; 45: 860-874.
- [41] Marfella R, Di Filippo C, Potenza N, Sardu C, Rizzo MR, Siniscalchi M, Musacchio E, Barbieri M, Mauro C, Mosca N, Solimene F, Mottola MT, Russo A, Rossi F, Paolisso G and D'Amico M. Circulating microRNA changes in heart failure patients treated with cardiac resynchronization therapy: responders vs. non-responders. Eur J Heart Fail 2013; 15: 1277-1288.
- [42] Akat KM, Moore-McGriff D, Morozov P, Brown M, Gogakos T, Correa Da Rosa J, Mihailovic A, Sauer M, Ji R, Ramarathnam A, Totary-Jain H, Williams Z, Tuschl T and Schulze PC. Comparative RNA-sequencing analysis of myocardial and circulating small RNAs in human heart failure and their utility as biomarkers. Proc Natl Acad Sci U S A 2014; 111: 11151-11156.
- [43] Dong S, Ma W, Hao B, Hu F, Yan L, Yan X, Wang Y, Chen Z and Wang Z. microRNA-21 promotes cardiac fibrosis and development of heart failure with preserved left ventricular ejection fraction by up-regulating Bcl-2. Int J Clin Exp Pathol 2014; 7: 565-574.
- [44] Gupta SK, Itagaki R, Zheng X, Batkai S, Thum S, Ahmad F, Van Aelst LN, Sharma A, Piccoli MT, Weinberger F, Fiedler J, Heuser M, Heymans S, Falk CS, Forster R, Schrepfer S and Thum T. miR-21 promotes fibrosis in an acute cardiac allograft transplantation model. Cardiovasc Res 2016; 110: 215-226.
- [45] Lorenzen JM, Schauerte C, Hubner A, Kolling M, Martino F, Scherf K, Batkai S, Zimmer K, Foinquinos A, Kaucsar T, Fiedler J, Kumarswamy R, Bang C, Hartmann D, Gupta SK, Kielstein J, Jungmann A, Katus HA, Weidemann F, Muller OJ, Haller H and Thum T. Osteopontin is indispensible for AP1-mediated angiotensin II-related miR-21 transcription during cardiac fibrosis. Eur Heart J 2015; 36: 2184-2196.
- [46] Sayed D, He M, Hong C, Gao S, Rane S, Yang Z and Abdellatif M. MicroRNA-21 is a down-

stream effector of AKT that mediates its antiapoptotic effects via suppression of Fas ligand. J Biol Chem 2010; 285: 20281-20290.

- [47] Kolpa HJ, Peal DS, Lynch SN, Giokas AC, Ghatak S, Misra S, Norris RA, Macrae CA, Markwald RR, Ellinor P, Bischoff J and Milan DJ. miR-21 represses Pdcd4 during cardiac valvulogenesis. Development 2013; 140: 2172-2180.
- [48] Bauters C, Kumarswamy R, Holzmann A, Bretthauer J, Anker SD, Pinet F and Thum T. Circulating miR-133a and miR-423-5p fail as biomarkers for left ventricular remodeling after myocardial infarction. Int J Cardiol 2013; 168: 1837-1840.
- [49] Seronde MF, Vausort M, Gayat E, Goretti E, Ng LL, Squire IB, Vodovar N, Sadoune M, Samuel JL, Thum T, Solal AC, Laribi S, Plaisance P, Wagner DR, Mebazaa A and Devaux Y; GREAT Network. Circulating microRNAs and outcome in patients with acute heart failure. PLoS One 2015; 10: e0142237.
- [50] Gundara JS, Zhao J, Gill AJ, Lee JC, Delbridge L, Robinson BG, McLean C, Serpell J and Sidhu SB. Noncoding RNA blockade of autophagy is therapeutic in medullary thyroid cancer. Cancer Med 2015; 4: 174-182.

- [51] Gerges M, Gerges C, Pistritto AM, Lang MB, Trip P, Jakowitsch J, Binder T and Lang IM. Pulmonary hypertension in heart failure. Epidemiology, right ventricular function, and survival. Am J Respir Crit Care Med 2015; 192: 1234-1246.
- [52] Breitling S, Ravindran K, Goldenberg NM and Kuebler WM. The pathophysiology of pulmonary hypertension in left heart disease. Am J Physiol Lung Cell Mol Physiol 2015; 309: L924-941.
- [53] Malhotra R, Dhakal BP, Eisman AS, Pappagianopoulos PP, Dress A, Weiner RB, Baggish AL, Semigran MJ and Lewis GD. Pulmonary vascular distensibility predicts pulmonary hypertension severity, exercise capacity, and survival in heart failure. Circ Heart Fail 2016; 9.
- [54] Wei C, Li L, Kim IK, Sun P and Gupta S. NFkappaB mediated miR-21 regulation in cardiomyocytes apoptosis under oxidative stress. Free Radic Res 2014; 48: 282-291.