Original Article

Lycium barbarum polysaccharide inhibits gastric cancer cell proliferation, migration and invasion by down-regulation of MMPs and suppressing epithelial-mesenchymal transition

Qian Chen¹, Rongliang Shi¹, Daowen Jiang¹, Weiyan Liu¹, Zhenyi Jia²

¹Department of General Surgery, Minhang Hospital, Fudan university, China; ²Department of General Surgery, Sixth People's Hospital Affiliated to Shanghai Jiao Tong University, China

Received November 10, 2016; Accepted January 13, 2017; Epub July 1, 2017; Published July 15, 2017

Abstract: Objective: To evaluate the effect of *Lycium barbarum* polysaccharide (LBP) on gastric cancer (GC) cells and to explore the associated mechanism. Methods: Human GC SGC-7901 cells were divided into control, 10 μ M LBP, 20 μ M LBP and 50 μ M LBP groups. CCK8 assay and Transwell assay were performed to evaluate the proliferation, migration and invasion of SGC-7901 cells. Western blotting was used to determine protein expressions. Results: The proliferation, migration and invasion decreased significantly in 20 μ M LBP and 50 μ M LBP groups. As the result of Western blotting, protein levels of MMP2, MMP9, Snail and vimentin decreased in 20 μ M LBP and 50 μ M LBP groups with different degrees. The expression E-cadherin significantly increased in all three experimental groups. The phosphorylation levels of AKT and PI3K in 20 μ M LBP and 50 μ M LBP groups were much lower than control group. Conclusion: LBP could inhibit the proliferation, migration and invasion of human GC cells bydown-regulation of MMPs and suppression of epithelial-mesenchymal transition (EMT).

Keywords: Lycium barbarum polysaccharide, gastric cancer, MMPs, epithelial-mesenchymal transition

Introduction

Gastric cancer (GC) is the second most frequent lethal cancer worldwide with approximately 50% cases in China [1, 2]. Most patients who are diagnosed with GC are in advanced stage of disease and predict extremely poor prognosis. The 5-year survival rate for GC patients varies between 5% in Stage IV and 90% in Stage I [3]. Although, in recent years, tremendous progress has been made in the development and advancement of GC research, the underlying molecular mechanisms involving migration and invasion are still poorly understood.

Lycium barbarum L. is a Solanaceous defoliated shrubbery that widely grows in arid and semi-arid regions of Northwestern China, Southeastern Europe and Mediterranean areas. Lycium barbarum L., also called Goji berry or wolfberry, is 1-2 cm long, bright orange-red

ellipsoid berries [4]. Lycium barbarum L. has been used in East Asia as a traditional herbal medicine and functional food [5, 6], with a large variety of beneficial effects on reducing blood glucose and serum lipids, nourishing eyes, kidneys and liver, anti-radiation, immunity improvement, anti-aging, anticancer, anti-fatigue, enhancing hemopoiesis and male infertility and so on [7-10]. Lycium barbarum polysaccharides (LBP) are a group of water-soluble glycoconjugates isolated from the aqueous extracts of Lycium barbarum L., containing six monosaccharides (arabinose, rhamnose, xylose, mannose, galactose and glucose) [11]. As one of the major active ingredients responsible for above biological activities, LBP are estimated to comprise 5-8% of the dried fruits [5, 12]. LBP exerted beneficial effects on the animal models of ocular diseases, such as protecting retinal ganglion cells and retinal vasculature from acute ocular hypertension, preserving retinal function after partial optic nerve transaction, reducing

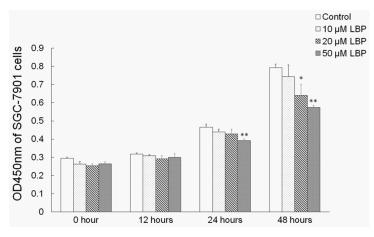


Figure 1. CCK8 assay was performed 0, 12, 24 and 48 h after adding LBP. To determine the amount of cells alive, absorbance was measured on 450 nm wavelength. *, *P*<0.05, vs control group; **, *P*<0.01, vs control group.

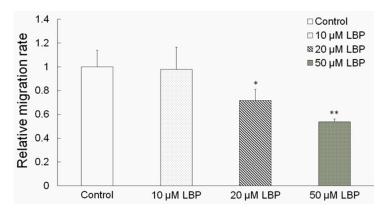


Figure 2. The relative migration rate of each group. *, *P*<0.05, vs control group; **, *P*<0.01, vs control group.

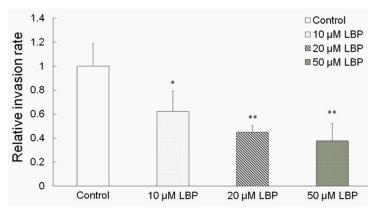


Figure 3. The relative invasion rate of each group. *, *P*<0.05, vs control group; **, *P*<0.01, vs control group.

neuronal damage, blood-retinal barrier disruption and oxidative stress in retinal ischemia/reperfusion injury [13-16].

Matrix metalloproteinases (MMPs) are a family of zinc-dependent endopeptidases which play an important role in the proteolytic destruction of extracellular matrix and basement membranes, thereby, they are essential for tumor invasion and metastasis [17]. MMPs, particularly MMP-2 and MMP-9 have been implicated in cancer invasion and metastasis [18].

Epithelial-mesenchymal transition (EMT) is a process in which epithelial cells lose their cell-cell contacts and undergo remodeling of the cytoskeleton to form a migratory phenotype [19]. The expression of epithelial marker proteins. such as E-cadherin and Keratin, are downregulated, while the expression of mesenchymal markers, such as vimentin and N-cadherin, are upregulated with EMT [20]. Therefore, it is widely accepted that EMT plays an important role in cancer metastasis of several human malignancies including GC [21]. The inhibition of EMT may improve GC, so EMT could be a very promising therapeutic target.

In this study, SGC-7901 cells were used to investigate the effects of LBP on human GC cells and the related mechanisms.

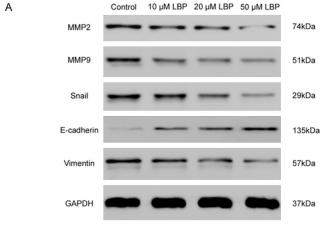
Methods

Cell line

A human GC SGC-7901 cell line (Beijing Dingguo Changsheng Biotech Co., Ltd, China) was cultured in RPMI1640 (Gibco, USA) containing 100 U/mL penicillin, 100 mg/L streptomycin and 10% heat-inactivated fetal bovine serum (Gibco, USA) at 37°C in humidified 5% CO₂ incubator.

Cell proliferation assay

SGC-7901 cells proliferation was measured by Cell Counting Kit-8 detection kit (Dojindo, Japan). Cells were seeded at a concentration of



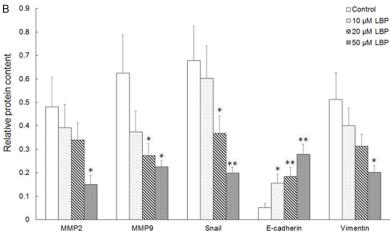


Figure 4. Western blotting. A. Western blotting of MMP2, MMP9, Snail, E-cadherin, Vimentin and their corresponding internal reference (GAPDH); B. The histogram for the content of proteins in each group. *, *P*<0.05, vs control group; **, *P*<0.01, vs control group.

 5×10^3 cells per well in 96-well plates. All experiments were conducted in triplicate. After grown for 24 h resulted in about 70-80% confluence, cells were performed with different treatments as described above. At 0, 12, 24 and 48 h after transfection, CCK-8 solution was applied at 10 μL per well and followed by 2-h incubation at 37°C. Absorbance values of all wells were then determined at 450 nm in Microplate Reader (Bio-Rad, USA).

Migration assay

Migration assays were performed by seeding 1×10^4 cells in 100 µL of RPMI1640 on top of Transwell cell culture inserts consisting of a non-coated polyethylene terephthalate membrane (24-well inserts, 8.0 µm pore size; Corning, USA). The lower chamber was filled with 0.6 mL of RPMI1640. After incubation for 24 h, the non-migrating cells were scraped off,

and the membranes were fixed and stained using the Diff-Quik™ stain kit (Sysmex, Japan). Cells that had migrated through the membranes were quantified by determination of the cell number in three randomly chosen visual fields at × 200 magnification.

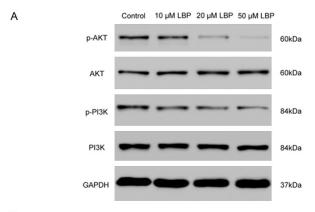
Matrigel invasion assay

Tumor cells in serum-free RPMI1640 were seeded on top of transwell inserts with 8 µm pore-size polyethylene terephthalate membrane coated with Matrigel™ basement membrane matrix (BD Biosciences, USA), whereas the lower chamber was filled with RPMI1640 with 1% FBS as chemoattractants. Cells were cultured for 24 h before the non-migrating cells in the inserts were scraped off; membranes were fixed and stained using Diff-Quik™ stain kit (Sysmex, Japan). The cells that had migrated through the membrane were quantified by determination of the cell number in three

randomly chosen visual fields at × 200 magnification.

Western blotting

Whole cell lysates were harvested and samples (50 µg protein/lane) were fractionated by SDS-PAGE and transferred to PVDF membranes. The membranes were incubated in 5% skimmed milk for 1 h at room temperature, and overnight at 4°C with primary antibodies against MMP2 (1:1000), MMP9 (1:500), Snail (1:1000), Ecadherin (1:1000), vimentin (1:1000) p-AKT (1:1000), AKT (1:1000), p-PI3K (1:1000), PI3K (1:1000) or GAPDH (1:2000) (Santa Cruz, USA). GAPDH was used as a loading control. And then secondary antibody (1:1000) was added and incubated for 2 h at room temperature. Bands were visualized using an ECL chemiluminescence kit (Genview, USA) and quantitated by Quantity One (Bio-Rad, USA).



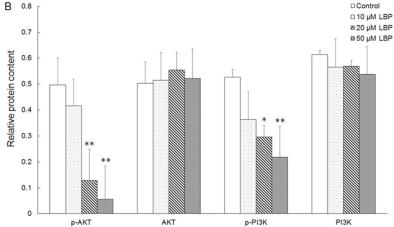


Figure 5. Western blotting. A. Western blotting of p-AKT, AKT, p-Pl3K, Pl3K and their corresponding internal reference (GAPDH); B. The histogram for the content of proteins in each group. *, P<0.05, vs control group; **, P<0.01, vs control group.

Statistical analysis

All data were expressed as mean \pm SD (\pm s), and the statistical differences between two different groups were assessed by Student's t-test. P<0.05 indicated a significant difference, P<0.01 indicated that there was a very significant difference. Prism 5.0 (Graphpad Software, Inc., Canada) was used to calculate statistical differences between groups.

Results

LBP inhibited the proliferation of SGC-7901 cells

As the result of CCK8 assay, the cell proliferation decreased very significantly in 50 μ M LBP group after 24 and 48 h (P<0.01, P<0.01). The cell proliferation also decreased in 20 μ M LBP group after 48 h (P<0.05). There was no difference between 10 μ M LBP group and control

group at any time point (*P*>0.05) (**Figure 1**). It demonstrated that LBP with high concentration could inhibit the proliferation of human GC cells.

LBP inhibited the migration of SGC-7901 cells

Migrated cells through the membrane in each group were counted in three random fields and then the relative migration rate was calculated (relative migration rate = the number of migrated cells/the number of migrated cells in control group). Figure 2 showed that the relative migration rate of 20 μM LBP and 50 μM LBP groups decreased significantly (P < 0.05, P < 0.01). It indicated that LBP could inhibit GC cell migration.

LBP inhibited the invasion of SGC-7901 cells

The relative invasion rate of each group was shown in Figure 3. The invasion of

SGC-7901 cells could be suppressed by LBP even with a low concentration.

LBP affected the protein expression in SGC-7901 cells

SGC-7091 cells in 50 μ M LBP group showed a significant decrease protein expression of MMP2 (P<0.05), MMP9 (P<0.05), Snail (P<0.01) and vimentin (P<0.05) (**Figure 4**). While 20 μ M LBP could significantly reduce the expression of MMP9 (P<0.05) and Snail (P<0.05). An increase expression of E-cadherin was observed in all three experimental groups (P<0.05; P<0.01; P<0.01).

As shown in **Figure 5**, AKT and PI3K were expressed at a similar level in control group and experimental groups, however LBP reduced the phosphorylation of these proteins in a concentration-dependent manner.

Discussion

LBP was a kind of polysaccharide-protein complex, which has anticancer and immunologic enhancement activities [22, 23]. This study showed that LBP inhibited the proliferation, migration and invasion of human GC cell line SGC-7901.

One potential mechanism by which LBP could inhibit the invasion and migration of GC cells is the down-regulation of MMPs levels [24]. It is well established that secretion of MMPs with the capacity for extracellular matrix (ECM) degradation is a feature of metastatic cancer cells [25]. MMP2 and MMP9 are two of the most well-characterized MMPs and are closely associated with cancer invasion and metastasis due to their strong proteolytic activity of ECM [26].

EMT plays key roles in the pathogenesis of cancer and other human diseases [27, 28]. During EMT, expression levels of the adhesion molecule E-cadherin are decreased, whereas vimentin levels are increased. These molecular alterations possibly cause dysfunctional cell-cell adhesion and loss of cell-cell junctions, thereby allowing dissemination of tumor cells from the primary sites. According to our experimental results, LBP had certain effects on inhibiting the migration and invasion of GC cells by the suppression of EMT.

PI3K signaling plays a key role in inducing and maintaining EMT. Cells expressing a constitutively active form of PKB/AKT, the most important downstream effector of PI3K signaling, induces the expression of Snail, which in turn represses E-cadherin gene transcription and induces EMT [29]. In the present study, it was shown that LBP reduced the expression of Snail and the phosphorylation of PI3K and AKT in cells. These results suggested that LBP could inhibit the PI3K/AKT/Snail signaling pathway which was involved in EMT of human GC cells.

Conclusion

In conclusion, LBP with certain concentration could inhibit the proliferation, migration and invasion of human GC cells by down-regulation of MMP2 and MMP9 and suppression EMT in GC cells.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Zhenyi Jia, Department of General Surgery, Sixth People's Hospital Affiliated to Shanghai Jiao Tong University, 600 Yishan Road, Xuhui District, Shanghai 200030, China. Tel: +86-21-64361349; Fax: +86-21-643-61349; E-mail: zhenyijia_1@126.com

References

- [1] Ferlay J, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. Int J Cancer 2010; 127: 2893-917.
- [2] Jing JJ, Liu HY, Hao JK, Wang LN, Wang YP, Sun LH, Yuan Y. Gastric cancer incidence and mortality in Zhuanghe, China, between 2005 and 2010. World J Gastroenterol 2012; 18: 1262-9.
- [3] Wang J, Yu JC, Kang WM, Ma ZQ. Treatment strategy for early gastric cancer. Surg Oncol 2012; 21: 119-23.
- [4] Amagase H, Farnsworth NR. A review of botanical characteristics, phytochemistry, clinical relevance in efficacy and safety of Lycium barbarum fruit (Goji). Food Research International 2011; 44: 1702-17.
- [5] Jin M, Huang Q, Zhao K, Shang P. Biological activities and potential health benefit effects of polysaccharides isolated from Lycium barbarum L. Int J Biol Macromol 2013; 54: 16-23.
- [6] Wang Z, Liu Y, Sun Y, Mou Q, Wang B, Zhang Y, Huang L. Structural characterization of LbGp1 from the fruits of Lycium barbarum L. Food Chem 2014; 159: 137-42.
- [7] Luo Q, Cai Y, Yan J, Sun M, Corke H. Hypoglycemic and hypolipidemic effects and antioxidant activity of fruit extracts from Lycium barbarum. Life Sci 2004; 76: 137-49.
- [8] Tang WM, Chan E, Kwok CY, Lee YK, Wu JH, Wan CW, Chan RY, Yu PH, Chan SW. A review of the anticancer and immunomodulatory effects of Lycium barbarum fruit. Inflammopharmacology 2012; 20: 307-14.
- [9] Tian XM, Wang R, Zhang BK, Wang CL, Guo H, Zhang SJ. Impact of Lycium barbarum polysaccharide and Danshensu on vascular endothelial growth factor in the process of retinal neovascularization of rabbit. Int J Ophthalmol 2013; 6: 59-61.
- [10] Luo Q, Li J, Cui X, Yan J, Zhao Q, Xiang C. The effect of Lycium barbarum polysaccharides on the male rats reproductive system and spermatogenic cell apoptosis exposed to low-dose

Effects of LBP on human GC cells

- ionizing irradiation. J Ethnopharmacol 2014; 154: 249-58.
- [11] Wang CC, Chang SC, Chen BH. Chromatographic determination of polysaccharides in Lycium barbarum Linnaeus. Food Chemistry 2009; 116: 595-603.
- [12] Tang HL, Chen C, Wang SK, Sun GJ. Biochemical analysis and hypoglycemic activity of a polysaccharide isolated from the fruit of Lycium barbarum L. Int J Biol Macromol 2015; 77: 235-42.
- [13] Chan HC, Chang RC, Koon-Ching Ip A, Chiu K, Yuen WH, Zee SY, So KF. Neuroprotective effects of Lycium barbarum Lynn on protecting retinal ganglion cells in an ocular hypertension model of glaucoma. Exp Neurol 2007; 203: 269-73.
- [14] Li SY, Yang D, Yeung CM, Yu WY, Chang RC, So KF, Wong D, Lo AC. Lycium barbarum polysaccharides reduce neuronal damage, blood-retinal barrier disruption and oxidative stress in retinal ischemia/reperfusion injury. PLoS One 2011; 6: e16380.
- [15] Mi XS, Feng Q, Lo AC, Chang RC, Lin B, Chung SK, So KF. Protection of retinal ganglion cells and retinal vasculature by Lycium barbarum polysaccharides in a mouse model of acute ocular hypertension. PLoS One 2012; 7: e45469.
- [16] Chu PH, Li HY, Chin MP, So KF, Chan HH. Effect of lycium barbarum (wolfberry) polysaccharides on preserving retinal function after partial optic nerve transection. PLoS One 2013; 8: e81339.
- [17] Nabeshima K, Inoue T, Shimao Y, Sameshima T. Matrix metalloproteinases in tumor invasion: role for cell migration. Pathol Int 2002; 52: 255-64.
- [18] Khasigov PZ, Podobed OV, Gracheva TS, Salbiev KD, Grachev SV, Berezov TT. Role of matrix metalloproteinases and their inhibitors in tumor invasion and metastasis. Biochemistry (Mosc) 2003; 68: 711-7.
- [19] Gao D, Vahdat LT, Wong S, Chang JC, Mittal V. Microenvironmental regulation of epithelialmesenchymal transitions in cancer. Cancer Res 2012; 72: 4883-9.

- [20] Tsai JH, Yang J. Epithelial-mesenchymal plasticity in carcinoma metastasis. Genes Dev 2013; 27: 2192-206.
- [21] Watanabe T, Takahashi A, Suzuki K, Kurusu-Kanno M, Yamaguchi K, Fujiki H, Suganuma M. Epithelial-mesenchymal transition in human gastric cancer cell lines induced by TNF-alphainducing protein of Helicobacter pylori. Int J Cancer 2014; 134: 2373-82.
- [22] Gan L, Zhang SH, Liu Q, Xu HB. A polysaccharide-protein complex from Lycium barbarum upregulates cytokine expression in human peripheral blood mononuclear cells. Eur J Pharmacol 2003; 471: 217-22.
- [23] Gan L, Zhang SH, Liu Q, Xu HB. Immunomodulation and antitumor activity by a polysaccharide-protein complex from Lycium barbarum. Int Immunopharmacol 2004; 4: 563-9.
- [24] Huang Q, Lan F, Wang X, Yu Y, Ouyang X, Zheng F, Han J, Lin Y, Xie Y, Xie F, Liu W, Yang X, Wang H, Dong L, Wang L, Tan J. IL-1beta-induced activation of p38 promotes metastasis in gastric adenocarcinoma via upregulation of AP-1/cfos, MMP2 and MMP9. Mol Cancer 2014; 13: 18.
- [25] Hanna SC, Krishnan B, Bailey ST, Moschos SJ, Kuan PF, Shimamura T, Osborne LD, Siegel MB, Duncan LM, O'Brien ET 3rd, Superfine R, Miller CR, Simon MC, Wong KK, Kim WY. HI-F1alpha and HIF2alpha independently activate SRC to promote melanoma metastases. J Clin Invest 2013; 123: 2078-93.
- [26] Lee SJ, Kim WJ, Moon SK. Role of the p38 MAPK signaling pathway in mediating interleukin-28A-induced migration of UMUC-3 cells. Int J Mol Med 2012; 30: 945-52.
- [27] Thiery JP, Acloque H, Huang RY, Nieto MA. Epithelial-mesenchymal transitions in development and disease. Cell 2009; 139: 871-90.
- [28] Kyprianou N. ASK-ing EMT not to spread cancer. Proc Natl Acad Sci U S A 2010; 107: 2731-2
- [29] Tiwari N, Gheldof A, Tatari M, Christofori G. EMT as the ultimate survival mechanism of cancer cells. Semin Cancer Biol 2012; 22: 194-207.