# Original Article

# Clinical significance of long non-coding RNA ZEB2-AS1 in locally advanced colorectal cancer

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Abstract: Objective: The aim of this study was to investigate the clinical significance of differential expression of long non-coding RNA (IncRNA) ZEB2-AS1 in patients with colorectal cancer (CRC). Methods: mRNA expression of IncRNA ZEB2-AS1 was evaluated by real-time quantitative PCR on eighty-seven cancerous tissues and adjacent normal mucosal tissues from patients with CRC tissue. Correlation between the IncRNA ZEB2-AS1 expression and clinico-pathological characteristics of the colorectal cancer patients was evaluated, and five-year overall survival (OS) was also analyzed according to the IncRNA ZEB2-AS1 expression of the CRC patients. Moreover, Cox Regression Analysis was performed in screening prognosis factors. Results: A significantly upregulated IncRNA ZEB2-AS1 expression, with a fold change of 18.75, was found in CRC tissue compared to the normal tissue. IncRNA ZEB2-AS1 expression in CRC was correlated with death (P<0.001). The five-year OS was 43.2% and 76.7%, respectively, in patients with higher and lower IncRNA ZEB2-AS1 expression. Cox regression analysis showed that location (P=0.020), N1 staging (P=0.021) and IncRNA ZEB2-AS1 lower expression (P<0.001) were independent prognosis factors associated with a better OS. Conclusion: Expression of IncRNA ZEB2-AS1 was significantly upregulated in stage III CRC patients and affects the prognosis.

Keywords: Colorectal cancer, IncRNA ZEB2-AS1, prognosis

#### Introduction

Colorectal cancer is the fourth most common cancer and the fifth most common cause of cancer-related deaths in China, with an estimated 331,300 newly diagnosed patients and 159,300 deaths in 2012 [1]. Surgical resection, followed by adjuvant chemotherapy, is the most commonly used strategy for colorectal cancer management. Despite the overall fiveyear survival rate of colorectal cancer improving to 65%, only a 15% five-year survival rate could be found in patients presented with distant metastasis [2], which reflects poor treatment response in some of the patients with colorectal cancer. Therefore, it is necessary to search for effective biomarkers in patients with colorectal cancer to improve the therapeutic benefits of current agents and to develop individualized therapies.

Long noncoding RNAs (IncRNAs) with a length >200 nucleotides are a recently discovered

novel class of genes with regulatory function but lacking in protein-coding ability. Multiple studies have proven the critical role of IncRNA in a wide range of cellular processes, including X chromosome inactivation, splicing, imprinting, epigenetic control and gene transcription regulation [3-5]. Furthermore, studies show that dysregulated expression of IncRNAs also exists in various human diseases, especially in cancers, including breast cancer, lung cancer, gastric cancer and colorectal cancer (CRC) [6-8]. According to the most recent evidence, IncRNAs were verified to be involved in the development and progression of human CRC and may serve as novel therapeutic targets [9-11]. However, the role of IncRNAs in CRC remains largely unknown.

Here, we investigated the clinical significance of differential expression of IncRNA ZEB2-AS1 in CRC patients and performed association analyses between the expression levels and the clinicopathological characteristics, thereby

Table 1. Primers for real-time PCR

Gene name	Forward primer (5'-3')	Reverse primer (5'-3')
LncRNA ZEB2-AS1	ATGAAGAAGCCGCGAAGTGT	CACACCCTA ATACACATG CCCT
GAPDH	TCGACAGTCAGCCGCATCTT	GGCGCCCAATACGACCAAAT

**Table 2.** Relationship between IncRNA ZEB2-AS1 expression and clinical characteristics in patients with colorectal cancer

Gender  Male 41 23 18 1.381 0.24  Female 46 20 26  Age  ≤60 49 27 22 1.446 0.229  >60 38 16 22  Location  Colon 42 21 21 0.011 0.918  Rectum 45 22 23  Histology  High 5 3 2 1.527 0.466  Moderate 61 32 29  Low 21 8 13  T staging  I 1 1 0 5.746 0.125  II 8 1 7  III 39 21 18  IV 39 20 19  N staging  N1 56 27 29 0.092 0.761  N2 31 16 15  Death  Yes 35 10 25 10.88 0.001  No 52 33 19	Characteristics	No.	LncRNA Z	EB2-AS1	2	
Male       41       23       18       1.381       0.24         Female       46       20       26       26         Age       ≤60       49       27       22       1.446       0.229         >60       38       16       22       1.446       0.229         Location       20       21       21       0.011       0.918         Rectum       45       22       23         Histology       45       22       23       1.527       0.466         Moderate       61       32       29       1.527       0.466         Moderate       61       32       29       29       0.466       0.125         II       1       1       0       5.746       0.125       1<			Low	High	X <sup>2</sup>	Р
Female       46       20       26         Age       ≤60       49       27       22       1.446       0.229         >60       38       16       22       1.446       0.229         Location       20       21       21       0.011       0.918         Rectum       45       22       23       23       1.527       0.466         Moderate       61       32       29       29       0.466       0.466       0.466       0.466       0.125       0.1	Gender					
Age	Male	41	23	18	1.381	0.24
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Colon       42       21       21       0.011       0.918         Rectum       45       22       23         Histology       1       32       2       1.527       0.466         Moderate       61       32       29       29       20       29       20       20       20       20       20       20       20       25       0.125       11       1       1       1       0       5.746       0.125       11       1       1       1       0       5.746       0.125       11       1 <td>&gt;60</td> <td>38</td> <td>16</td> <td>22</td> <td></td> <td></td>	>60	38	16	22		
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Low 21 8 13 T staging I 1 1 1 0 5.746 0.125 II 8 1 7 III 39 21 18 IV 39 20 19 N staging N1 56 27 29 0.092 0.761 N2 31 16 15 Death Yes 35 10 25 10.88 0.001	High	5	3	2	1.527	0.466
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IV 39 20 19  N staging  N1 56 27 29 0.092 0.761  N2 31 16 15  Death  Yes 35 10 25 10.88 0.001	II	8	1	7		
N staging       N1       56       27       29       0.092       0.761         N2       31       16       15         Death       7es       35       10       25       10.88       0.001	III	39	21	18		
N1     56     27     29     0.092     0.761       N2     31     16     15       Death       Yes     35     10     25     10.88     0.001	IV	39	20	19		
N2 31 16 15  Death Yes 35 10 25 10.88 0.001	N staging					
Death Yes 35 10 25 10.88 0.001	N1	56	27	29	0.092	0.761
Yes 35 10 25 10.88 0.001	N2	31	16	15		
	Death					
No 52 33 19	Yes	35	10	25	10.88	0.001
	No	52	33	19		

providing information for CRC diagnosis and prognosis.

#### Materials and methods

### Patients

A total of eighty-seven cases of surgically resected CRC specimens with complete clinical data were collected from the Second Affiliated Hospital of Soochow University between January 2009 and June 2012. The inclusion criteria of the patients were: (1) diagnosis of primary CRC, (2) no history of chemotherapy and radiotherapy, and (3) histologically confirmed adenocarcinoma. All of the patients were diag-

nosed and classified by two experienced pathologists according to The Union for International Cancer Control (UICC). Moreover, eighty-seven cases of non-

cancerous mucous tissue were obtained as controls. The study protocol was approved by the Ethics Committee of the Second Affiliated Hospital of Soochow University. Written informed consent was provided by each patient that was included.

Most of the patients were followed up by the Outpatient Department. The survival period was recorded from the date of surgery to the date of death, or until the last follow up time. The last follow up was done in June 2017. The minimum follow up period was six months and the maximum period was eighty-eight months for surviving patients.

Real-time quantitative PCR for IncRNA ZEB2-AS1

Cancer tissues and normal control tissues were collected and grinded into powder before total tissue RNA extraction with TRIzol Reagent (Invitrogen). RT-PCR was carried out using a One Step SYBR® PrimeScript<sup>TM</sup> RT-PCR kit (Takara, Dalian, China) and an ABI 7500 Real-Time PCR System (Applied Biosystems, Foster City, CA, USA). Expression of the glyceraldehyde 3-phosphate dehydrogenase (GAPDH) gene was assayed simultaneously with samples as an internal control. Relative gene expression was determined by the  $2-\Delta\Delta$ CT method [14]. Oligonucleotide primers specific for LncRNA ZEB2-AS1 and GAPDH are listed in **Table 1**.

#### Statistical analysis

Descriptive statistics for the patient group were reported as mean  $\pm$  standard deviation (SD) or median and range. The category data was presented as number and percentages. The correlation analysis between relative expression of IncRNA ZEB2-AS1 and clinical data of the patients was performed using a Chi-square Test. The Kaplan-Meier method was used to determine five-year overall survivals. Multivariate analysis was performed using the Cox proportional hazards model. Statistical significance was accepted at the P<0.05 level. Statistical analyses were performed using SPSS 16.0 software (SPSS Inc., Chicago, IL).

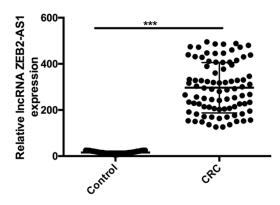
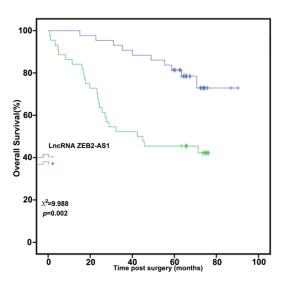


Figure 1. Increased expression of IncRNA ZEB2-AS1 in colorectal cancer tissue. The expression level of IncRNA ZEB2-AS1 was determined by real-time quantitative PCR. A fold of 18.75 was found on colorectal cancer tissues compared to the control tissue.



**Figure 2.** Kaplan-Meier curves for overall survival in patients with colorectal cancer according to the IncRNA ZEB2-AS1 expression.

#### Results

#### Patient demographic data

A total of eighty-seven CRC patients were included in the present study. The median age of the CRC patients was 60 (30-82) years. Of these CRC patients, 41 were male and 46 were female. The mean follow up time of these CRC patients was 55.4±22.3 months, ranging from 6-88 months. The demographic data of these patients are listed in **Table 2**.

Increased IncRNA ZEB2-AS1 expression in CRC tissues

To determine the IncRNA ZEB2-AS1 expression in CRC, we performed quantitative PCR analy-

sis with the tissue samples collected from the above-mentioned patients. As shown in **Figure 1**, a significantly increased level of IncRNA ZEB2-AS1, with a fold change of 18.74, was found on CRC tissues compared to the control tissue (*P*<0.001).

Correlation analysis between IncRNA ZEB2-AS1 expression and clinicopathological characteristics of CRC

We also performed correlation analysis between IncRNA ZEB2-AS1 expression and clinicopathological characteristics of CRC. As shown in **Table 2**, no correlation was found between the IncRNA ZEB2-AS1 expression level and gender (P=0.24), or age (P=0.229), or differentiation (P=0.466), or T stage (P=0.125), or N stage (P=0.761), while significance was found between IncRNA ZEB2-AS1 expression and death (P=0.001) (**Table 2**).

#### Survival analysis

The median OS of the patients with high and low IncRNA ZEB2-AS1 expression was, respectively, 23.2±4.8 months and 40.1±10.8 months (Figure 2). The accumulated survival ratio in patients with high and low IncRNA ZEB2-AS1 expression was, respectively, 43.2% and 76.7%. Significantly prolonged OS was found in patients with low IncRNA ZEB2-AS1 expression compared to those with high IncRNA ZEB2-AS1 expression. Then, we used the Cox proportional hazards model to assess the clinicopathological characteristics and we found that location (P=0.020), N staging (P=0.021), and differential expression levels of IncRNA ZEB2-AS1 (P< 0.001) were the independent risk factors of five-year OS (**Table 3**).

#### Discussion

Searching for effective biomarkers in patients with colorectal cancer could be of great importance in improving the therapeutic benefits of current agents and in the development of individualized therapies [12].

In this present study, we performed real-time quantitative PCR to evaluate the expression level of IncRNA ZEB2-AS1 on CRC tissues to explore its clinical significance in CRC tissues. Our results demonstrated that significantly upregulated IncRNA ZEB2-AS1 expression, with a fold change of 18.75, was found in CRC tissue compared to normal tissue. Moreover, the cor-

**Table 3.** Cox regression analysis of the prognosis in the patients with stage III colorectal cancer

Ob a va ata viati aa	RR	95.0% CI		Р
Characteristics		Upper	Lower	value
Gender	1.423	0.666	0.363	3.044
Age	0.851	0.375	0.701	1.932
Location	0.390	0.176	0.020	0.865
T staging	1.117	0.659	0.682	1.894
N staging	2.347	1.135	0.021	4.850
Differentiation	1.649	0.864	0.129	3.145
LncRNA ZEB2-AS1 level	4.209	1.948	0.000	9.093

RR: risk ratio; CI: confidence interval.

relation analysis revealed that IncRNA ZEB2-AS1 expression in CRC was correlated with death (P<0.001), and the five-year OS was, respectively, 43.2% and 76.7% in patients with higher and lower IncRNA ZEB2-AS1 expression. In addition, Cox regression analysis showed that location (P=0.020), N1 staging (P=0.021) and IncRNA ZEB2-AS1 lower expression (P<0.001) were independent prognosis factors associated with a better OS. To the best of our knowledge, this is the first study concerning the role of IncRNA ZEB2-AS1 in CRC.

As a class of newly discovered genes, IncRNAs, with gene regulatory function but without protein coding ability, are suggested to play a critical role in physiological function regulation. According to previous studies, approximately 18% of IncRNAs are associated with human tumors and have been shown to act as major contributors in the development and progression of human cancers [13]. Multiple mechanisms have been suggested about the regulatory role of IncRNAs in physiological functions, including trans-regulatory and cis-regulatory mechanisms [14-17], and the representative IncRNAs including PTENP1 [18], H19 [19], and CCAT1 [20].

ZEB, as a transcriptional factor, has been identified to play an important role in the process of EMT, which is closely associated with carcinogenesis. Beltran et al. first discovered that a non-coding antisense transcript located from the promoters of ZEB2 (ZEB2-AS1) had the ability to activate ZEB2 expression [21]. Recently, Li et al. reported that the ZEB1-AS1 gene was involved in the tumorigenesis of hepatocellular carcinoma (HCC) and functioned as a noncoding oncogene [22]. They found upregulated

IncRNA ZEB1-AS1 expression level in HCC tissues compared with the adjacent normal tissues. Moreover, Li et al. also described the effects of IncRNA ZEB1-AS1 on cell proliferation, migration and invasion, and cell cycle regulation. In addition, ZEB1-AS1 may promote tumor growth and metastasis in HCC patients through regulating expression levels of ZEB1. However, Lan et al. recently found that downregulation of IncRNA ZEB2-AS1 was associated with reduced tumor growth and metastasis in HCC tissues [23]. However, no study has focused on the role of IncRNA ZEB2-AS1 on CRCs. Here, we report an upregulated pattern of IncRNA ZEB2-AS1 on CRC tissue which was consistent with its pattern in HCC. Moreover, we also found that IncRNA ZEB2-AS1 was correlated with overall survival, which may serve as prognosis marker.

There are also some limitations in our study. First, this is a single-center small study and insufficient sample size could affect the final conclusion. Second, the diagnosis value of IncRNA ZEB2-AS1 was also implicated by the results obtained here, however, the accurate cut-off value of IncRNA ZEB2-AS1 expression quantification should be calculated by an ROC curve in future studies to maximize the diagnostic value of IncRNA ZEB2-AS1. Third, according to previous studies, elevated IncRNA ZEB2-AS1 was associated with early death and high incidence of death. Due to the lack of detailed data of the cause of death, we could not analyze the associations of these features with IncRNA ZEB2-AS1 level. Fourth, the detailed mechanisms that resulted in the upregulation of IncRNA ZEB2-AS should be elucidated for the exact role of IncRNA ZEB2-AS1 in CRC. Taken together, further investigation is of great importance to identify the biomarker role of IncRNA ZEB2-AS1 by a multi-center clinical large sample size study with prognosis results.

In conclusion, our findings suggest that expression of IncRNA ZEB2-AS1 is significantly upregulated in the stage III CRC patients, and thus may be employed as a biomarker for cancer prognosis and malignant diagnosis.

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#### Disclosure of conflict of interest

None.

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#### References

- [1] Chen W, Zheng R, Zuo T, Zeng H, Zhang S, He J. National cancer incidence and mortality in China, 2012. Chin J Cancer Res 2016; 28: 1-11.
- [2] Jaacks LM, Siegel KR, Gujral UP, Narayan KM. Type 2 diabetes: a 21st century epidemic. Best Pract Res Clin Endocrinol Metab 2016; 30: 331-343.
- [3] Yue M, Charles Richard JL, Ogawa Y. Dynamic interplay and function of multiple non-coding genes governing X chromosome inactivation. Biochim Biophys Acta 2016; 1859: 112-120.
- [4] Butler AA, Webb WM, Lubin FD. Regulatory RNAs and control of epigenetic mechanisms: expectations for cognition and cognitive dysfunction. Epigenomics 2016; 8: 135-151.
- [5] Kanduri C. Long non-coding RNAs: lessons from genomic imprinting. Biochim Biophys Acta 2016; 1859: 102-111.
- [6] Tordonato C, Di Fiore PP, Nicassio F. The role of non-coding RNAs in the regulation of stem cells and progenitors in the normal mammary gland and in breast tumors. Front Genet 2015; 6: 72.
- [7] Prensner JR, Iyer MK, Balbin OA, Dhanasekaran SM, Cao Q, Brenner JC, Laxman B, Asangani IA, Grasso CS, Kominsky HD, Cao X, Jing X, Wang X, Siddiqui J, Wei JT, Robinson D, Iyer HK, Palanisamy N, Maher CA, Chinnaiyan AM. Transcriptome sequencing across a prostate cancer cohort identifies pcat-1, an unannotated lincrna implicated in disease progression. Nat Biotechnol 2011; 29: 742-749.
- [8] Li W, Zheng J, Deng J, You Y, Wu H, Li N, Lu J, Zhou Y. Increased levels of the long intergenic non-protein coding RNA pou3f3 promote DNA methylation in esophageal squamous cell carcinoma cells. Gastroenterology 2014; 146: 1714-1726, e1715.

- [9] Yin D, He X, Zhang E, Kong R, De W, Zhang Z. Long non-coding RNA gas5 affects cell proliferation and predicts a poor prognosis in patients with colorectal cancer. Med Oncol 2014; 31: 253.
- [10] Han Y, Yang YN, Yuan HH, Zhang TT, Sui H, Wei XL, Liu L, Huang P, Zhang WJ, Bai YX. Uca1, a long non-coding RNA up-regulated in colorectal cancer influences cell proliferation, apoptosis and cell cycle distribution. Pathology 2014; 46: 396-401.
- [11] Ding J, Lu B, Wang J, Wang J, Shi Y, Lian Y, Zhu Y, Wang J, Fan Y, Wang Z, De W, Wang K. Long non-coding RNA loc554202 induces apoptosis in colorectal cancer cells via the caspase cleavage cascades. J Exp Clin Cancer Res 2015; 34: 100.
- [12] Pu X, Ye Y, Wu X. Development and validation of risk models and molecular diagnostics to permit personalized management of cancer. Cancer 2014; 120: 11-19.
- [13] Khachane AN, Harrison PM. Mining mammalian transcript data for functional long non-coding RNAs. PLoS One 2010; 5: e10316.
- [14] Bhan A, Mandal SS. Lncrna hotair: a master regulator of chromatin dynamics and cancer. Biochim Biophys Acta 2015; 1856: 151-164.
- [15] Dimitrova N, Zamudio JR, Jong RM, Soukup D, Resnick R, Sarma K, Ward AJ, Raj A, Lee JT, Sharp PA, Jacks T. Lincrna-p21 activates p21 in cis to promote polycomb target gene expression and to enforce the g1/s checkpoint. Mol Cell 2014; 54: 777-790.
- [16] Wang K, Long B, Zhou LY, Liu F, Zhou QY, Liu CY, Fan YY, Li PF. Carl Incrna inhibits anoxia-induced mitochondrial fission and apoptosis in cardiomyocytes by impairing mir-539-dependent phb2 downregulation. Nat Commun 2014; 5: 3596.
- [17] Tay Y, Rinn J, Pandolfi PP. The multilayered complexity of cerna crosstalk and competition. Nature 2014; 505: 344-352.
- [18] Yu G, Yao W, Gumireddy K, Li A, Wang J, Xiao W, Chen K, Xiao H, Li H, Tang K, Ye Z, Huang Q, Xu H. Pseudogene ptenp1 functions as a competing endogenous RNA to suppress clear-cell renal cell carcinoma progression. Mol Cancer Ther 2014; 13: 3086-3097.
- [19] Kallen AN, Zhou XB, Xu J, Qiao C, Ma J, Yan L, Lu L, Liu C, Yi JS, Zhang H, Min W, Bennett AM, Gregory RI, Ding Y, Huang Y. The imprinted h19 Incrna antagonizes let-7 microRNAs. Mol Cell 2013; 52: 101-112.
- [20] Ma MZ, Chu BF, Zhang Y, Weng MZ, Qin YY, Gong W, Quan ZW. Long non-coding RNA ccat1 promotes gallbladder cancer development via negative modulation of mirna-218-5p. Cell Death Dis 2015; 6: e1583.

## Long non-coding RNA (IncRNA) ZEB2-AS1 in colorectal cancer

- [21] Beltran M, Puig I, Pena C, García JM, Alvarez AB, Peña R, Bonilla F, de Herreros AG. A natural antisense transcript regulates zeb2/sip1 gene expression during snail1-induced epithelial-mesenchymal transition. Genes Dev 2008; 22: 756-769.
- [22] Li T, Xie J, Shen C, Cheng D, Shi Y, Wu Z, Deng X, Chen H, Shen B, Peng C, Li H, Zhan Q, Zhu Z. Upregulation of long non-coding RNA zeb1-as1 promotes tumor metastasis and predicts poor prognosis in hepatocellular carcinoma. Oncogene 2016; 35: 1575-1584.
- [23] Lan T, Chang L, Wu L, Yuan Y. Downregulation of zeb2-as1 decreased tumor growth and metastasis in hepatocellular carcinoma. Mol Med Rep 2016; 14: 4606-4612.