### Review Article

## A Practical and Comprehensive Immunohistochemical Approach to the Diagnosis of Superficial Soft Tissue Tumors

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Abstract: Soft tissue tumors include neoplasms of specific and unknown lineages, and, therefore, lineage markers of smooth muscle, skeletal muscle, endothelial, epithelial and Schwann cells have proven useful in everyday practice. However, groups of tumors remain that are defined essentially on grounds of histology; others can be defined by molecular genetic studies. The complex distribution patterns of many antigens and loss of some differentiation antigens in malignant tumors often necessitate the use of panels of antibodies. Optimally such panels should address all significant differential diagnostic alternatives. There is little doubt that numerous new differentiation markers will appear in the future. The evaluation of tumor proliferation, apoptosis, and cell cycle control will give new information related to tumor biology and prognosis.

Key Words: Soft tissue tumors, skin, immunohistochemistry

#### Introduction

Immunohistochemistry (IHC) is presently the most important adjunct tool in the evaluation of soft tissue tumors because of its practicability and relatively low cost [1]. As more data has accumulated over the years, typical diagnostic patterns for many tumor types have emerged. Nevertheless, many diagnoses, such as those of fibrous, fibrohistiocytic and lipomatous tumors, are still based on histology. Furthermore, a small of soft tissue tumors remains unclassified despite extensive ancillary studies. Because of the complex patterns of expression of many antigens, the use of panels of antibodies is often necessary. New modalities of antigen recovery, especially heat induced antigen/epitope retrieval, have made possible the application of many antibodies that previously were believed reactive only in frozen tissues. Standardization of reagents, optimization of techniques, and proactive quality assurance are vitally important for the success of diagnostic IHC [2]. This review summarizes the application of the most important IHC tools as applied to the diagnosis

of specific types of superficial soft tissue tumors. Earlier reviews discussed the biology of specific antigens, such as intermediate filament proteins, S100 protein, muscle cell, and neural antigens [3-5]. Other ancillary tests, such as molecular genetic tests for translocation-specific fusion transcripts, are becoming useful in the evaluation of such specific tumor types as myxoid liposarcoma, alveolar rhabdomyosarcoma, synovial sarcoma, clear cell sarcoma, Ewing's sarcoma, and desmoplastic small round cell tumor [6].

#### Fibroblastic/Myofibroblastic Tumors

There are no universally applicable markers for fibroblastic lineage. However, similar to subsets of dermal and soft tissue fibroblasts, some fibroblastic neoplasms, such as dermatofibrosarcoma protuberans (DFSP) and solitary fibrous tumors are positive for CD34, the haematopoietic progenitor cell antigen also expressed in endothelial cells [7-10]. In contrast, other fibroblastic tumors including benign fibrous histiocytoma (FH), desmoid and nodular fasciitis are generally negative for CD34, although occasional benign FHs (5-

10%) have been CD34-positive [7, 11]. DFSPs have undergone fibrosarcomatous transformation commonly lose the CD34 expression [2, 13]. Benign FH typically shows a significant factor XIIIa-positive cell population, whereas DFSP is negative [7, 14]. Another potentially useful marker in the differential diagnosis of FH, cellular FH and DFSP is CD163 as it was reported to be positive in most FHs and cellular FHs with DFSPs being mostly negative [15]. In addition, CD44 has been claimed to be positive in dermatofibroma rather than DFSP [16]. These preliminary data is still to be validated. A panel composed of CD34, factor XIIIa, CD163 and CD44 will have a significantly higher sensitivity and specificity in this differential diagnosis. However, if any histologic doubts exist, FISH techniques will give the answer [17].

**Fibromatoses** are locally aggressive myofibroblastic proliferations that characterised by higher rate of recurrence. Higher levels of nuclear β-catenin have been reported in all types of fibromatoses (superficial and deep) [18, 19]. However, a subset of other soft tissue lesions such as solitary fibrous tumor. low grade myofibroblastic sarcoma and synovial sarcoma may show B-catenin immunoreactivity [20, 21].

Fibroblastic/myofibroblastic sarcoma is a controversial group of low grade sarcomas prominent myofibroblastic with differentiation [22]. The immunophenotype of these lesions is variable but they are consistently positive for at least one myogenic marker 23]. In one study, all cases of myofibroblastic sarcoma were positive for smooth muscle actin (SMA) and 2/3 of cases were positive for calponin, whereas none reacted with either smooth muscle myosin (SMMS) or h-caldesmon. This reaction pattern was similar to that seen in nodular fasciitis and fibromatosis [24].

The haemosiderotic fibrolipomatous tumor is histologically characterised by the presence of mature adipose tissue and spindle cell component accompanied by haemosidern pigment deposition within macrophages, in the cytoplasm of some of the spindle cells and also within the extracellular stroma. The spindle cell component of these lesions is generally positive for CD34 and negative for CD68, S100, SMA and desmin [25].

Recently, CD34, EMA and CD99 were found very useful in the diagnosis of the newly described entity "Superficial Acral Fibromyxoma" [26].

#### So-called Fibrohistiocytic Tumors

Histiocytic markers have limited applications in the diagnosis of soft tissue tumors with CD163 as an exception [27, 28]. True histiocytic differentiation occurs only in a few tumors, and can be identified in the lesional cells of juvenile xanthogranuloma and in true histiocytic neoplasms (true histiocytic lymphoma/sarcoma) [29]. It can also be seen in most cases of primitive myeloid tumors and in soft tissues named as extramedullary myeloid tumors or granulocytic sarcoma. Histiocyte-rich non-neoplastic lesions include necrobiotic granulomas, xanthomas. fibroxanthomas and foreign body and other granulomas. In addition, high numbers of reactive histiocytes are seen in the so-called fibrohistiocytic tumors. However, the presence of large numbers of histiocytes is not specific for fibrohistiocytic tumors as they may occur in other types of sarcomas, especially in high grade and undifferentiated tumors.

Malignant fibrous histiocytoma (MFH) is a designation used for poorly differentiated sarcomas that do not show any specific differentiation, except perhaps fibroblastic differentiation. This diagnosis is, therefore, made by exclusion of other specific diagnoses, leiomyosarcoma, including metastatic sarcomatoid carcinoma, and melanoma [30, 31]. Because this tumor does not display true histiocytic differentiation, histiocytic markers have no role in its diagnosis. In two studies employing multiple lineage markers for histiomonocytic cells, MFH cells were negative, suggesting their fibroblast-like phenotype [32, 33]. However, MFH cells often express CD68, which is not specific for histiocytes, together with SMA and CD34 [34]. It is also possible that MFHs contain higher numbers of reactive histiocytes than other sarcomas; such a high content of histiocytes has probably led previous electron microscopy studies to conclude that MFH shows histiocytic differentiation.

Among the best histiocytic markers is lysozyme. However, it does not stain all histiocytes as the lysozyme content is dependent on the functional status of the

histiocytes. Moreover, some non-histiocytic cells are considered to be rich in lysozyme. CD68 (KP1) and NKI-C3 react with most histiocytes, but it also reacts with lysosomal components in cells of any lineage. Therefore, it is positive in many lysosome-rich tumors, for example, granular cell tumors, Schwann cell tumors, melanomas, and high grade sarcomas of different phenotypes. Some carcinomas may also be CD68-positive, limiting the value of this marker, except in narrow settings [35].

Factor XIIIa, a component in the coagulation pathway, is expressed in histiocytes and related cells and has been shown in a number of fibrous and fibrohistiocytic tumors [36]. Our impression on the numerous Factor XIIIa-positive cells in the fibrohistiocytic tumors, such as MFH, is that they represent abundant non-neoplastic histiocytes that typically infiltrate these tumors.

#### **Lipomatous Tumors**

No specific markers are currently available for the diagnosis of fatty tumors. Normal fat cells and some neoplastic adipocytes are positive for \$100-protein, and 1 study showed that \$100 can be useful in the diagnosis of such poorly differentiated tumors as round cell liposarcoma [37]. We have found that \$100 reactivity is inconsistent in fatty tumors, even in the lipogenic components; we therefore do not routinely apply this marker.

Recently, an immunohistochemical panel composed of MDM2 and CDK4 was recommended differentiate atypical to lipomatous tumor/well differentiated liposarcomas (ALT/WDLS) from benign adipose tumors and to separate dedifferentiated liposarcomas (DDLS) from poorly differentiated sarcomas [38, 39]. In studies, majority of ALT/WDLS these expressed both markers, whereas benign adipose tumors were predominantly negative. In addition, the dedifferentiated component of DDLS usually retained the expression of MDM2 and CDK4 [38].

Spindle cell lipoma and the closely related pleomorphic lipoma are relatively uncommon subcutaneous tumors typically seen in the posterior neck of older men. These lesions contain foci of bland (or pleomorphic) spindle cells which are strongly CD34-positive [40]. Often fatty tumors such as well-differentiated

liposarcoma and spindle cell non-lipogenic components of fatty tumors (including some dedifferentiated liposarcoma) may show CD34 [41].

# Angiomyolipoma and Related Tumors (PEComas)

Angiomyolipoma most commonly occur in the kidney, but similar tumors may also present in the retroperitoneum, skin and liver. Recently, sugar tumor of the lung and similar tumors identified in other organs have been found closely related to angiomyolipoma. It was suggested that they represent neoplasms (sometimes referred to as hamartomas) of a unique perivascular epithelioid cell population (PEComas). The lymphangioleiomyomas of lung and retroperitoneum also belong to this group [42, 43].

Immunohistochemically angiomyolipomas. lymphangiomyomas, and related tumors are distinctive and coexpress markers of smooth muscle (consistently SMA-positive, variably desmin-positive) and melanocytes (HMB-45, Melan-A, MART). Therefore, such constellation of markers is useful in the diagnosis of angiomyolipoma-related mesenchymal tumors, especially when they occur in an unusual clinical setting [42, 44].

#### **Smooth Muscle Tumors**

Leiomyomas are typically positive for muscle actin when evaluated with the monoclonal antibody HHF-35 or with antibodies to alpha-SMA. However, both antibodies also react with myoepithelial cells and the latter also with myofibroblasts, as seen, for example, in the mvofibroblast-rich nodular fasciitis. Therefore. strong SMA-reactivity itself is not diagnostic of a smooth muscle cell tumor [45]. Typical leiomyosarcomas generally show prominent actin reactivity similar to that seen in benign leiomyomas, but desmin reactivity is variable and may be absent. In our experience, approximately 70% of leiomyosarcomas are desmin positive, and the reactivity is often focal [46-48].

Newer smooth muscle markers of diagnostic interest include various isoforms of myosin, especially smooth muscle myosin. Our experience has shown that the results of antibodies to smooth muscle myosin are essentially similar to muscle actin antibodies.

Heavy molecular weight isoform of caldesmon (HCD) is a cytoskeleton-associated calmodulin and actin binding protein that is expressed in most smooth muscle tumors but not in rhabdomyosarcoma [49]. Similar to muscle actins, HCD is also expressed in myoepithelial cells but not in myofibroblasts, and it can be considered a valuable alternative marker for the diagnosis of smooth muscle tumors [50]. Calponin is yet another cytoskeletonassociated protein shared by smooth muscle subsets of myofibroblasts, myoepithelial cells. It appears to have less value in the diagnosis of smooth muscle tumors, as it is present in reactive and neoplastic myofibroblasts and curiously, in subsets of spindle cells in synovial sarcoma [49].

Neither typical leiomyomas nor leiomyosarcomas react with CD34. Some nonneoplastic, benign neoplastic, and malignant smooth muscle tumors express keratins [51, 52]. For example, myometrium commonly dot-like keratin reactivity shows antibodies that react to keratins 8, 18 and 19, and keratin expression in myometrial cells has been confirmed by western blotting [53]. Epithelial membrane antigen expression may also occur in leiomyosarcomas; the expression epithelial markers in 20-30% leiomyosarcomas should be considered in the differential diagnosis of sarcomas, and spindle cell carcinomas [52].

#### Skeletal Muscle Tumors

Primary cutaneous rhabdomyosarcoma is extremely rare. Only scattered case reports are found in the literature [54-56]. Most rhabdomyosarcomas and rhabdomyoblastic components in other tumors are positive for desmin and muscle actins (HHF-35), but are typically negative for alpha-SMA [5, 57, 58]. Myoglobin can be demonstrated only in differentiated rhabdomyoblasts, and may be useful in the search of rhabdomyoblasts and in the verification of scattered rhabdomyoblasts. However, it is of limited value in the diagnosis of poorly differentiated rhabdomyosarcomas because such tumors, including most cases of alveolar rhabdomyosarcomas are typically negative. In our experience, the applicability of myoglobin immunostaining is also limited by the common background problems with many of the antibodies available. Cells that are not positive for desmin and actin are almost never

positive for myoglobin.

MyoD1 is a skeletal muscle specific transcription factor that is located in the nucleus. MyoD1 has been shown to be a sensitive and specific adjunct for the diagnosis of paediatric rhabdomyosarcoma. Pleomorphic sarcomas in adults have been verified as rhabdomyosarcomas based on MyoD1 nuclear expression [59-61]. Cytoplasmic MyoD1 staining has also been described in rhabdomyosarcomas; however, this pattern may occur in other tumors and cannot be considered specific for rhabdomyosarcoma [62].

Many childhood rhabdomyosarcomas exhibit polyphenotypic features, as they may express keratins, neurofilaments and non-specific enolase (NSE). These features should be considered in the differential diagnosis of epithelial and keratin-positive or neural and neurofilament-positive tumors [63]. Reactivity for \$100 protein commonly occurs in skeletal muscle, especially in atrophic muscle fibres, and it is therefore not surprising that rhabdomyosarcomas may also be \$100 protein positive [63].

#### Vascular Tumors

The diagnosis of endothelial differentiation is often challenging, especially in cases of angiosarcoma showing limited vasoformation or morphologically simulating epithelial tumors.

Von Willebrand factor (VWF, also called factor VIII-related antigen) is expressed in vascular cells. megakaryocytes endothelial platelets, and most cases of benign vascular tumors. Although it is very useful in the differential diagnosis of borderline epithelioid hemangioendotheliomas and epithelioid angiosarcomas from carcinomas. VWF is often undetectable in other types of angiosarcomas, as it is only partially conserved in malignant endothelial cells. Because the antibody to VWF also reacts with the soluble antigen present in hemorrhagic and necrotic tissues, such tissue components in VWF immunostains are often uninterpretable [64].

CD34 (the haematopoietic progenitor cell antigen) is a glycosylated transmembrane protein of unknown function, which is constitutionally expressed in haematopoietic

stem cells, most types of vascular endothelial cells, and subsets of fibroblasts, especially observed in perivascular and periadnexal locations in the skin and soft tissues. Lymphatics often show a weaker expression [64, 65]. CD34 is consistently expressed in Kaposi sarcoma, but is variably present in angiosarcomas and epithelioid hemangioendotheliomas (approximately 50%). The reactivity of CD34 in a variety of fibroblastic, lipomatous, and gastrointestinal stromal tumors and epithelioid sarcoma also limits its value as an endothelial cell marker, but does extends the application of this marker to the subtyping of other mesenchymal tumors [66].

CD31 (platelet-endothelial cell adhesion molecule 1, PE-CAM-1) is a cell surface molecule with immunoglobulin homology. CD31 is expressed in all endothelial cells, platelets and in subsets of haematopoietic cells, including haematopoietic stem cells and some histiocytes [67]. CD31 is considered the most sensitive marker for endothelial cell: however, histiocytes can be positive, which is a hidden pitfall [68]. It is present in most angiosarcomas and Kaposi's sarcomas and currently is the single most useful marker in diagnosis of poorly differentiated the angiosarcomas in terms of sensitivity and specificity. In typical cases, distinct membrane staining is observed. Also, epithelioid vascular tumors. including epithelioid hemangioendotheliomas and epithelioid angiosarcomas, are typically positive in approximately 85-95% of cases [64, 67, 69]. Reactivity with platelets causes staining in areas of thrombosis and haemorrhage. CD31 reactivity in occasional epithelial tumors has been described [70].

FLI-1 protein is a nuclear transcription factor which is frequently used in the immunehistochemical diagnosis of Ewing's sarcoma/primitive neuroectodermal tumors. It is also expressed in endothelial cells and was reported to be highly sensitive in the detection of vascular neoplasms [71, 72]. However, it is also expressed in lymphocytes which may produce a background staining and lead to the erroneous identification of intratumoural lymphocytes as endothelial cells. Moreover, its expression in other, non-endothelial tumors, limits its use as a stand-alone vascular marker [73].

Human erythrocyte-type glucose transporter protein (GLUT-1) was found to be a reliable immunohistochemical marker in distinguishing haemangiomas (GLUT-1 positive) from vascular malformations and pyogenic granuloma (GLUT-1 negative) [74, 75].

Kaposi's sarcoma is a human herpes virus-8 (HHV-8) induced vascular tumor. The 4 epidemiological forms of Kaposi's sarcoma (classic, endemic, iatrogenic and HIV-related) show nuclear expression of HHV-8 latent nuclear antigen-1 [76, 77], which is considered a highly sensitive and specific marker for differentiating Kaposi's sarcoma from other spindle cell tumors [78].

#### **Perivascular Tumors**

(HPC) is defined Hemangiopericytoma histologically, and it comprises a group of benign and borderline tumors that present in a wide variety of locations, the most common being retroperitoneum and extremities. Malignant overtly sarcomatous tumors with a hemangiopericytoma-like pattern usually represent synovial sarcomas or other type of soft tissue sarcomas. HPC differs from the normally actin-positive pericytes, because the lesional spindle cells, with rare exceptions, do not express actins [79, 80]. However, the spindle cells are typically CD34-positive but negative for desmin and keratins. Only the endothelial cells in these tumors react with CD31. Therefore, hemangiopericytoma probably is a tumor of primitive perivascular mesenchymal cells rather than tumor of differentiated pericytes.

Solitary fibrous tumor (SFT), which commonly shows hemangiopericytoma-like histologic patterns in some areas of the tumors, is immunohistochemically inseparable from hemangiopericytoma and is defined by histology. Both SFT and HPC are CD34-positive and actin and desmin-negative tumors, which probably overlap, especially when presenting in non-serosal sites [8, 10]. Some SFTs express CD99, Bcl-2 and less frequently SMA but they are consistently negative for desmin, factor XIIIa, keratin, EMA and S100 [81].

Myopericytoma is a tumor originating from perivascular myoid cells sharing features of both smooth muscle cells and glomus cells. This tumor usually displays diffuse positivity with SMA with occasional focal and weak positivity for desmin. Immunostaining for S100, HMB45, CD34 and keratin is negative [82].

Glomus tumors are considered to be arising from modified smooth muscle cells and therefore they show a smooth muscle-like phenotype by their consistent muscle actin (HHF-35) and alpha-SMA positivity. A significant proportion of these tumors are positive for CD34 but they are consistently negative for CD31. They rarely express desmin and are negative for S100 and keratins; the latter finding is useful in the differential diagnosis from epithelial skin adnexal tumors with round cell morphology [79, 83, 84].

#### **Schwann Cell Tumors**

Benign neurofibromas are typically composed of admixed Schwannian and fibroblastic elements. The former are positive for \$100 protein, whereas the latter (often an extensive component) are positive for CD34 [85]. Combined use of these 2 markers is, therefore, useful in the diagnosis of neurofibroma. According to our experience, preservation of the S100+ and CD34+ cell populations is typical in benign neurofibromas, and such a dual population is typically absent in malignant peripheral nerve sheath tumors. differential diagnosis of diffuse neurofibroma and DFSP, both of which often show diffuse fat infiltration, is aided by the S100 protein reactivity of the former.

Benign schwannomas are composed of compact sheets of Schwann cells, variably prominent blood vessels and loose, degenerative foci often rich in histiocytes. The neoplastic spindle cell components are almost uniformly S100 protein positive, whereas CD34 is seen only in the pericapsular area and loose, degenerative areas. Schwannomas commonly express glial fibrillary acidic protein (GFAP) and are sometimes positive for keratins [86].

Additional markers expressed in Schwann cells include basement membrane components collagen type IV and laminin. Schwannomas show abundant basement membrane deposition around the thin cell processes of the Schwann cells and are typically strongly positive for basement membrane proteins, with apparent diffuse staining in histological sections [87, 88]. Granular cell tumor of soft

tissues is believed to be of Schwannian derivation. This tumor is consistently \$100 protein positive and also reacts with CD68 (KP1) because of its high content of lysosomes [35, 89, 90].

Malignant peripheral nerve sheath tumors (MPNSTs) can be definitively diagnosed when they occur in connection with a pre-existent neurofibroma or nerve trunk, often in connection with neurofibromatosis type 1. These tumors can be tentatively diagnosed by histologic appearance and expression of Schwannian markers, especially \$100 protein. However, less than 50% of MPNSTs are S100positive. Low grade MNPSTs show a more diffuse S-100 staining compared to high grade tumours, which tend to show a patchy and weak staining pattern [91]. In our experience, epithelioid subtypes tend to be diffusely positive for S100 [4, 92]. MNPSTs are negative for keratin, EMA, GFAP. neurofilament, desmin and CD34 [92]. Occasional cases showed focal positivity for SMA [93], 93, Ki67 index ranged between 5%-65% in MPNSTs, while none of the benign peripheral sheath tumors had a labelling index greater than 5% [94]. Another useful marker is the percentage of p53 expression as it ranges from 5-100% in MPNSTs and not more than 1% in benign peripheral nerve sheath tumors [94].

Epithelial sheath neuroma is a recently described entity representing a superficial dermal proliferation of enlarged nerve fibers ensheathed by squamous epithelium. The nerve fibers are immunoreactive for \$100 protein, neurofilaments, CD57, and nerve growth factor receptor, whereas the perineural epithelial sheaths are positive for cytokeratins [95].

Neurothekeoma is a benign tumor of uncertain histogenesis. It has been postulated to be arising from fibroblasts/myofibroblasts [96]. These tumors are typically positive for NKI/C3, vimentin, NSE, CD10, and microphthalmia transcription factor, with occasional tumors showing focal reactivity for SMA and CD68. All tumors were negative for \$100 protein, GFAP, and Melan A [96, 97]. It has to be noted that NKI/C3 is not entirely specific neurothekeoma as it can be expressed in a wide range of histiocytic lesions and therefore correlation with the morphological picture is required [98].

#### **Perineurial Cell Tumors**

Perineuriomas are very rare soft tissue tumors that show cellular differentiation similar to perineurial (and meningeal) cells and express EMA, Glut-1 and claudin-1; they are negative for S100 protein [99, 100]. Such an IHC constellation, similar to that observed in meningiomas, has been used to identify examples of benign spindle cell and epithelioid and malignant perineuriomas [101, 102].

#### **Paragangliomas**

Paragangliomas are a group of neural neuroendocrine tumors. Most common examples of this group include pheochromocytomas of the adrenal. retroperitoneal extra-adrenal paragangliomas, carotid body, tympanic, and vagal paragangliomas. Skin is one of the rare and unusual sites for paragangliomas [103]. Although the organoid pattern is often sufficiently distinctive, it may be poorly developed in malignant variants. On the other hand, other tumors such as neuroendocrine carcinomas. smooth muscle tumors. meningiomas, and melanomas may show organoid patterns simulating those of paraganglioma.

Common to most paragangliomas is the expression of neural markers chromogranin, synaptophysin, and neuro-filaments and negativity for keratins and EMA in the chief cell component. Benign paragangliomas typically contain delicate, elongated \$100-positive Schwann cell-like, so-called sustentacular cells in the periphery of the spherical organoid clusters [104]. This component may also be GFAP-positive. Analysis of malignant paragangliomas suggests common loss of the sustentacular cell component [104].

#### **Synovial Sarcoma**

The occurrence of primary cutaneous synovial sarcoma is extremely rare [105]. The biphasic malignant tumors composed of glandular and spindle cell elements in soft tissues usually represent synovial sarcomas and can usually be diagnosed without special studies. The monophasic spindle cell type is probably the most common variant of synovial sarcoma. Although the uniform pattern of spindle cell with pointed nuclei and scattered microcalcifications is highly suggestive for synovial

sarcoma, differential diagnosis from nerve sheath or smooth muscle tumors may be difficult. Both the epithelial and the spindle cell components of biphasic synovial sarcoma show pancytokeratin and EMA positivity. In monophasic tumors, scattered spindle cells are positive for both markers. In contrast to haemangiopericytoma/solitary fibrous tumor CD34 is consistently negative [106].

EMA may be a more sensitive marker than keratins for monophasic and poorly differentiated synovial sarcomas, and most cases show patchy or streaky reactivity [107]. The possible, usually focal, S100 reactivity in synovial sarcoma should not be interpreted to indicate nerve sheath differentiation. Similarly, CD99 reactivity occurs in all variants of synovial sarcoma and should not lead to the diagnosis of Ewing's sarcoma and related tumors [108]. Analysis of poorly differentiated synovial sarcomas in comparison with other tumors have shown that keratin 7, typically seen in poorly differentiated as well as other synovial sarcomas, is absent in MPNSTs [109, 1101.

#### Clear Cell Sarcoma and Metastatic Melanoma

Clear cell sarcoma of tendons and aponcuroses is a rare soft tissue sarcoma. which predominantly occurs in the distal extremities of young adults. Occasional case reports of primary cutaneous clear cell sarcoma have been reported [111, 112]. This tumor shows melanocytic differentiation and compartmentalization of tumor cells between thick fibrous septa. Clear cell sarcoma is typically variably positive for S100 protein and usually strongly positive for HMB-45, while negative for keratin and muscle cell markers [113]. Although the histologic clinicopathologic features are distinctive, its diagnosis requires exclusion of (metastatic) melanoma.

or bulky primary malignant Metastatic melanoma is a common and often difficult problem in soft tissue diagnosis as these tumors are often non-pigmented and can show various combinations of sarcomatous spindle cell, carcinoma-like epithelioid or lymphomalike round cell patterns. The demonstration of strong S100 protein reactivity in a malignant soft tissue tumor always necessitates consideration of malignant melanoma in the differential diagnosis.

Curiously, there remains a significant group of melanoma-like soft tissue tumors without apparent primary tumors elsewhere. They may represent primary epithelioid nerve sheath tumors, but such a diagnosis requires clinical of melanoma. Metastatic exclusion melanomas have been stated to show HMB-45-reactivity in over 90% of cases, but lack of this marker should not detract from the diagnosis of melanoma; there is some evidence that HMB-45 may be deleted in metastases [114]. New melanoma markers tyrosinase and Melan-A/MART (melanoma antigen recognized by T-cells) have been shown equally as sensitive as HMB-45, and have been detected in approximately 85% of amelanotic melanomas [114, 115]. Keratinexpression has been described in some melanomas (in our experience as often as in 10-20%), and the presence of keratins has been confirmed by western blotting [116].

#### **Epithelioid Sarcoma**

Epithelioid sarcoma is an uncommon soft tissue sarcoma that typically occurs in the distal extremities of young adults; but it may also occur in more proximal locations where it often shows large cells, sometimes with rhabdoid cytoplasm [117]. Keratins and EMA are present in most if not all cases [118, 119].

In our experience, all variants of epithelioid sarcoma typically show reactivity for EMA and low molecular weight keratins 8, 18, and 19, but usually not for keratin 7 (if present only focally). Keratins of higher molecular weight, such as K5 and K14, occur focally in the majority of tumors. A feature that was found useful when using K5/6 in differentiating epithelioid sarcoma which showed focal and weak staining from spindle cell squamous cell carcinoma which showed diffuse and strong staining [120]. Approximately 50% of epithelioid sarcomas are positive for CD34, and one-third displays muscle immunoreactivity. Although the biologic significance of the latter 2 findings is open, such findings are diagnostically useful. Vimentin negativity of epithelioid sarcoma has been occasionally described, but according to our experience, such a finding is very rare if heat-induced epitope retrieval is employed [121]. S100 and desmin are usually negative [122].

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#### References

- [1] Erlandson RA and Rosai J. A realistic approach to the use of electron microscopy and other ancillary diagnostic techniques in surgical pathology. Am J Surg Pathol 1995; 19:247-250.
- [2] Elias JM, Gown AM, Nakamura RM, Wilbur DC, Herman GE, Jaffe ES, Battifora H and Brigati DJ. Quality control in immunohistochemistry. Report of a workshop sponsored by the Biological Stain Commission. Am J Clin Pathol 1989;92:836-843.
- [3] Miettinen. Immunohistochemistry of softtissue tumors. Possibilities and limitations in surgical pathology. *Pathol Annu* 1990;25 Pt 1:1-36.
- [4] Wick MR, Swanson PE, Ritter JH and Fitzgibbon JF. The immunohistology of cutaneous neoplasia: a practical perspective. *J Cutan Pathol* 1993;20:481-497.
- [5] Ordonez NG. Application of immunocytochemistry in the diagnosis of soft tissue sarcomas: a review and update. *Adv Anat Pathol* 1998;5:67-85.
- [6] Ladanyi. The emerging molecular genetics of sarcoma translocations. *Diagn Mol Pathol* 1995:4:162-173.
- [7] Abenoza P and Lillemoe T. CD34 and factor XIIIa in the differential diagnosis of dermatofibroma and dermatofibrosarcoma protuberans. Am J Dermatopathol 1993;15: 429-434.
- [8] Hanau CA and Miettinen M. Solitary fibrous tumor: histological and immunohistochemical spectrum of benign and malignant variants presenting at different sites. Hum Pathol 1995;26:440-449.
- [9] Kutzner H. Expression of the human progenitor cell antigen CD34 (HPCA-1) distinguishes dermatofibrosarcoma protuberans from fibrous histiocytoma in formalin-fixed, paraffin-embedded tissue. J Am Acad Dermatol 1993;28:613-617.
- [10] Westra WH, Gerald WL and Rosai J. Solitary fibrous tumor. Consistent CD34 immunoreactivity and occurrence in the orbit. Am J Surg Pathol 1994;18:992-998
- [11] Hsi ED and Nickoloff BJ. Dermatofibroma and dermatofibrosarcoma protuberans: an immunohistochemical study reveals distinctive antigenic profiles. *J Dermatol Sci* 1996;11:1-9.
- [12] Goldblum JR. CD34 positivity in fibrosarcomas which arise in dermato-

- fibrosarcoma protuberans. Arch Pathol Lab Med 1995; 119:238-241.
- [13] Goldblum JR, Reith JD and Weiss SW. Sarcomas arising in dermatofibrosarcoma protuberans: a reappraisal of biologic behavior in eighteen cases treated by wide local excision with extended clinical follow up. Am J Surg Pathol 2000;24:1125-1130.
- [14] Altman DA, Nickoloff BJ and Fivenson DP. Differential expression of factor XIIIa and CD34 in cutaneous mesenchymal tumors. J Cutan Pathol 1993;20:154-158.
- [15] Sachdev R and Sundram U. Expression of CD163 in dermatofibroma, cellular fibrous histiocytoma, and dermatofibrosarcoma protuberans: comparison with CD68, CD34, and Factor XIIIa. J Cutan Pathol 2006;33: 353-360.
- [16] Calikoglu E, Augsburger E, Chavaz P, Saurat JH and Kaya G. CD44 and hyaluronate in the differential diagnosis of dermatofibroma and dermatofibrosarcoma protuberans. J Cutan Pathol 2003;30:185-189.
- [17] Kaur S, Vauhkonen H, Bohling T, Mertens F, Mandahl N and Knuutila S. Gene copy number changes in dermatofibrosarcoma protuberans - a fine-resolution study using array comparative genomic hybridization. Cytogenet Genome Res 2006;115:283-288.
- [18] Varallo VM, Gan BS, Seney S, Ross DC, Roth JH, Richards RS, McFarlane RM, Alman B and Howard JC. Beta-catenin expression in Dupuytren's disease: potential role for cellmatrix interactions in modulating betacatenin levels in vivo and in vitro. Oncogene 2003;22:3680-3684.
- [19] Montgomery E, Torbenson MS, Kaushal M, Fisher C and Abraham SC. Beta-catenin immunohistochemistry separates mesenteric fibromatosis from gastrointestinal stromal tumor and sclerosing mesenteritis. Am J Surg Pathol 2002;26:1296-1301.
- [20] Ng TL, Gown AM, Barry TS, Cheang MC, Chan AK, Turbin DA, Hsu FD, West RB and Nielsen TO. Nuclear beta-catenin in mesenchymal tumors. *Mod Pathol* 2005;18:68-74.
- [21] Carlson JW and Fletcher CD. Immunohistochemistry for beta-catenin in the differential diagnosis of spindle cell lesions: analysis of a series and review of the literature. Histopathology 2007;51:509-514.
- [22] Diaz-Cascajo C, Borghi S, Weyers W and Metze D. Fibroblastic/myofibroblastic sarcoma of the skin: a report of five cases. J Cutan Pathol 2003;30:128-134.
- [23] Mentzel T, Dry S, Katenkamp D and Fletcher CD. Low-grade myofibroblastic sarcoma: analysis of 18 cases in the spectrum of myofibroblastic tumors. Am J Surg Pathol 1998;22:1228-1238.
- [24] Perez-Montiel MD, Plaza JA, Dominguez-Malagon H and Suster S. Differential expression of smooth muscle myosin, smooth

- muscle actin, h-caldesmon, and calponin in the diagnosis of myofibroblastic and smooth muscle lesions of skin and soft tissue. *Am J Dermatopathol* 2006:28:105-111.
- [25] Browne TJ and Fletcher CD. Haemosiderotic fibrolipomatous tumour (so-called haemosiderotic fibrohistiocytic lipomatous tumour): analysis of 13 new cases in support of a distinct entity. *Histopathology* 2006;48: 453-461.
- [26] Fetsch JF, Laskin WB and Miettinen M. Superficial acral fibromyxoma: a clinicopathologic and immunohistochemical analysis of 37 cases of a distinctive soft tissue tumor with a predilection for the fingers and toes. *Hum Pathol* 2001;32:704-714.
- [27] Miettinen M and Fetsch JF. Reticulohistiocytoma (solitary epithelioid histiocytoma): a clinicopathologic and immunohistochemical study of 44 cases. *Am J Surg Pathol* 2006;30:521-528.
- [28] Nguyen TT, Schwartz EJ, West RB, Warnke RA, Arber DA and Natkunam Y. Expression of CD163 (hemoglobin scavenger receptor) in normal tissues, lymphomas, carcinomas, and sarcomas is largely restricted to the monocyte/macrophage lineage. Am J Surg Pathol 2005;29:617-624.
- [29] Sangueza OP, Salmon JK, White CR Jr and Beckstead JH. Juvenile xanthogranuloma: a clinical, histopathologic and immunohistochemical study. J Cutan Pathol 1995;22: 327-335.
- [30] Fletcher CD. Pleomorphic malignant fibrous histiocytoma: fact or fiction? A critical reappraisal based on 159 tumors diagnosed as pleomorphic sarcoma. Am J Surg Pathol 1992;16:213-228.
- [31] Tos AP. Classification of pleomorphic sarcomas: where are we now? *Histopathology* 2006;48:51-62.
- [32] Wood GS, Beckstead JH, Turner RR, Hendrickson MR, Kempson RL and Warnke RA. Malignant fibrous histiocytoma tumor cells resemble fibroblasts. Am J Surg Pathol 1986;10:323-335.
- [33] Roholl PJ, Kleyne J and Van Unnik JA. Characterization of tumor cells in malignant fibrous histiocytomas and other soft-tissue tumors, in comparison with malignant histiocytes. II. Immunoperoxidase study on cryostat sections. *Am J Pathol* 1985;121: 269-274.
- [34] Nascimento AF and Raut CP. Diagnosis and management of pleomorphic sarcomas (socalled "MFH") in adults. J Surg Oncol 2008; 97:330-339.
- [35] Tsang WY and Chan JK. KP1 (CD68) staining of granular cell neoplasms: is KP1 a marker for lysosomes rather than the histiocytic lineage? *Histopathology* 1992;21:84-86.
- [36] Nemes Z and Thomazy V. Factor XIIIa and the

- classic histiocytic markers in malignant fibrous histiocytoma: a comparative immunohistochemical study. *Hum Pathol* 1988;19:822-829.
- [37] Dei Tos AP, Mentzel T and Fletcher CD. Primary liposarcoma of the skin: a rare neoplasm with unusual high grade features. Am J Dermatopathol 1998;20:332-338.
- [38] Binh MB, Sastre-Garau X, Guillou L, de Pinieux G, Terrier P, Lagace R, Aurias A, Hostein I and Coindre JM. MDM2 and CDK4 immunostainings are useful adjuncts in diagnosing well-differentiated and dedifferentiated liposarcoma subtypes: a comparative analysis of 559 soft tissue neoplasms with genetic data. Am J Surg Pathol 2005;29:1340-1347.
- [39] Binh MB, Garau XS, Guillou L, Aurias A and Coindre JM. Reproducibility of MDM2 and CDK4 staining in soft tissue tumors. Am J Clin Pathol 2006;125:693-697.
- [40] Templeton SF, Solomon AR, Jr.: Spindle cell lipoma is strongly CD34 positive. An immunohistochemical study. J Cutan Pathol 1996;23:546-550.
- [41] Suster S and Fisher C. Immunoreactivity for the human hematopoietic progenitor cell antigen (CD34) in lipomatous tumors. Am J Surg Pathol 1997;21:195-200.
- [42] Bonetti F, Pea M, Martignoni G, Doglioni C, Zamboni G, Capelli P, Rimondi P and Andrion A. Clear cell ("sugar") tumor of the lung is a lesion strictly related to angiomyolipoma--the concept of a family of lesions characterized by the presence of the perivascular epithelioid cells (PEC). Pathology 1994;26: 230-236.
- [43] Chan JK, Tsang WY, Pau MY, Tang MC, Pang SW and Fletcher CD. Lymphangiomyomatosis and angiomyolipoma: closely related entities characterized by hamartomatous proliferation of HMB-45-positive smooth muscle. Histopathology 1993;22:445-455.
- [44] Fetsch PA, Fetsch JF, Marincola FM, Travis W, Batts KP and Abati A. Comparison of melanoma antigen recognized by T cells (MART-1) to HMB-45: additional evidence to support a common lineage for angiomyolipoma, lymphangiomyomatosis, and clear cell sugar tumor. Mod Pathol 1998;11:699-703.
- [45] Schurch W, Skalli O, Seemayer TA and Gabbiani G. Intermediate filament proteins and actin isoforms as markers for soft tissue tumor differentiation and origin. I. Smooth muscle tumors. Am J Pathol 1987;128:91-103.
- [46] Truong LD, Rangdaeng S, Cagle P, Ro JY, Hawkins H and Font RL. The diagnostic utility of desmin. A study of 584 cases and review of the literature. Am J Clin Pathol 1990;93: 305-314.

- [47] Rangdaeng S and Truong LD. Comparative immunohistochemical staining for desmin and muscle-specific actin. A study of 576 cases. Am J Clin Pathol 1991;96:32-45.
- [48] Parham DM, Reynolds AB and Webber BL. Use of monoclonal antibody 1H1, anticortactin, to distinguish normal and neoplastic smooth muscle cells: comparison with anti-alpha-smooth muscle actin and antimuscle-specific actin. *Hum Pathol* 1995; 26:776-783.
- [49] Miettinen MM, Sarlomo-Rikala M, Kovatich AJ and Lasota J. Calponin and h-caldesmon in soft tissue tumors: consistent h-caldesmon immunoreactivity in gastrointestinal stromal tumors indicates traits of smooth muscle differentiation. *Mod Pathol* 1999;12:756-762.
- [50] D'Addario SF, Morgan M, Talley L and Smoller BR. h-Caldesmon as a specific marker of smooth muscle cell differentiation in some soft tissue tumors of the skin. J Cutan Pathol 2002;29:426-429.
- [51] Norton AJ, Thomas JA and Isaacson PG. Cytokeratin-specific monoclonal antibodies are reactive with tumours of smooth muscle derivation. An immunocytochemical and biochemical study using antibodies to intermediate filament cytoskeletal proteins. Histopathology 1987;11:487-499.
- [52] Miettinen M. Immunoreactivity for cytokeratin and epithelial membrane antigen in leiomyosarcoma. Arch Pathol Lab Med 1988; 112:637-640.
- [53] Gown AM, Boyd HC, Chang Y, Ferguson M, Reichler B, Tippens D. Smooth muscle cells can express cytokeratins of "simple" epithelium. Immunocytochemical and biochemical studies in vitro and in vivo. Am J Pathol 1988;132:223-232.
- [54] Tari AS, Amoli FA, Rajabi MT, Esfahani MR and Rahimi A. Cutaneous embryonal rhabdomyosarcoma presenting as a nodule on cheek; a case report and review of literature. Orbit 2006;25:235-238.
- [55] Brecher AR, Reyes-Mugica M, Kamino H and Chang MW. Congenital primary cutaneous rhabdomyosarcoma in a neonate. *Pediatr Dermatol* 2003;20:335-338.
- [56] Gong Y, Chao J, Bauer B, Sun X and Chou PM. Primary cutaneous alveolar rhabdomyosarcoma of the perineum. Arch Pathol Lab Med 2002;126:982-984.
- [57] Azumi N, Ben-Ezra J and Battifora H. Immunophenotypic diagnosis of leiomyosarcomas and rhabdomyosarcomas with monoclonal antibodies to muscle-specific actin and desmin in formalin-fixed tissue. *Mod Pathol* 1988;1:469-474.
- [58] Schmidt RA, Cone R, Haas JE, Gown AM: Diagnosis of rhabdomyosarcomas with HHF35, a monoclonal antibody directed

- against muscle actins. Am J Pathol 1988; 131:19-28.
- [59] Wesche WA, Fletcher CD, Dias P, Houghton PJ and Parham DM. Immunohistochemistry of MyoD1 in adult pleomorphic soft tissue sarcomas. Am J Surg Pathol 1995;19:261-269.
- [60] Wang NP, Marx J, McNutt MA, Rutledge JC and Gown AM. Expression of myogenic regulatory proteins (myogenin and MyoD1) in small blue round cell tumors of childhood. Am J Pathol 1995;147:1799-1810.
- [61] Tallini G, Parham DM, Dias P, Cordon-Cardo C, Houghton PJ and Rosai J. Myogenic regulatory protein expression in adult soft tissue sarcomas. A sensitive and specific marker of skeletal muscle differentiation. Am J Pathol 1994;144:693-701.
- [62] Wang NP, Bacchi CE, Jiang JJ, McNutt MA and Gown AM. Does alveolar soft-part sarcoma exhibit skeletal muscle differentiation? An immunocytochemical and biochemical study of myogenic regulatory protein expression. Mod Pathol 1996;9:496-506.
- [63] Coindre JM, de Mascarel A, Trojani M, de Mascarel I and Pages A. Immunohistochemical study of rhabdomyosarcoma. Unexpected staining with S100 protein and cytokeratin. J Pathol 1988;155:127-132.
- [64] Miettinen M, Lindenmayer AE and Chaubal A. Endothelial cell markers CD31, CD34, and BNH9 antibody to H- and Y-antigensevaluation of their specificity and sensitivity in the diagnosis of vascular tumors and comparison with von Willebrand factor. Mod Pathol 1994;7:82-90.
- [65] Traweek ST, Kandalaft PL, Mehta P and Battifora H. The human hematopoietic progenitor cell antigen (CD34) in vascular neoplasia. Am J Clin Pathol 1991;96:25-31.
- [66] Cohen PR, Rapini RP and Farhood AI. Dermatopathologic advances in clinical research. The expression of antibody to CD34 in mucocutaneous lesions. *Dermatol Clin* 1997;15:159-176.
- [67] Kuzu I, Bicknell R, Harris AL, Jones M, Gatter KC and Mason DY. Heterogeneity of vascular endothelial cells with relevance to diagnosis of vascular tumours. J Clin Pathol 1992;45: 143-148.
- [68] McKenney JK, Weiss SW and Folpe AL. CD31 expression in intratumoral macrophages: a potential diagnostic pitfall. Am J Surg Pathol 2001;25:1167-1173.
- [69] DeYoung BR, Swanson PE, Argenyi ZB, Ritter JH, Fitzgibbon JF, Stahl DJ, Hoover W and Wick MR. CD31 immunoreactivity in mesenchymal neoplasms of the skin and subcutis: report of 145 cases and review of putative immunohistologic markers of endothelial differentiation. J Cutan Pathol 1995;22:215-222.

- [70] De Young BR, Frierson HF Jr., Ly MN, Smith D and Swanson PE. CD31 immunoreactivity in carcinomas and mesotheliomas. Am J Clin Pathol 1998;110:374-377.
- [71] Pusztaszeri MP, Seelentag W, Bosman FT: Immunohistochemical expression of endothelial markers CD31, CD34, von Willebrand factor, and Fli-1 in normal human tissues. J Histochem Cytochem 2006;54: 385-395.
- [72] Folpe AL, Chand EM, Goldblum JR and Weiss SW. Expression of Fli-1, a nuclear transcription factor, distinguishes vascular neoplasms from potential mimics. Am J Surg Pathol 2001;25:1061-1066.
- [73] Rossi S, Orvieto E, Furlanetto A, Laurino L, Ninfo V and Dei Tos AP. Utility of the immunohistochemical detection of FLI-1 expression in round cell and vascular neoplasm using a monoclonal antibody. *Mod Pathol* 2004;17:547-552.
- [74] Dyduch G, Okon K and Mierzynski W. Benign vascular proliferations—an immunohistochemical and comparative study. Pol J Pathol 2004;55:59-64.
- [75] Leon-Villapalos J, Wolfe K and Kangesu L. GLUT-1: an extra diagnostic tool to differentiate between haemangiomas and vascular malformations. Br J Plast Surg 2005:58:348-352.
- [76] Robin YM, Guillou L, Michels JJ and Coindre JM. Human herpesvirus 8 immunostaining: a sensitive and specific method for diagnosing Kaposi sarcoma in paraffin-embedded sections. Am J Clin Pathol 2004;121:330-334
- [77] Cheuk W, Wong KO, Wong CS, Dinkel JE, Ben-Dor D and Chan JK. Immunostaining for human herpesvirus 8 latent nuclear antigen-1 helps distinguish Kaposi sarcoma from its mimickers. Am J Clin Pathol 2004;121:335-342.
- [78] Patel RM, Goldblum JR and Hsi ED. Immunohistochemical detection of human herpes virus-8 latent nuclear antigen-1 is useful in the diagnosis of Kaposi sarcoma. *Mod Pathol* 2004;17:456-460.
- [79] Porter PL, Bigler SA, McNutt M, Gown AM: The immunophenotype of hemangio-pericytomas and glomus tumors, with special reference to muscle protein expression: an immunohistochemical study and review of the literature. *Mod Pathol* 1991;4:46-52.
- [80] Schurch W, Skalli O, Lagace R, Seemayer TA and Gabbiani G. Intermediate filament proteins and actin isoforms as markers for soft-tissue tumor differentiation and origin. III. Hemangiopericytomas and glomus tumors. Am J Pathol 1990;136:771-786.
- [81] Erdag G, Qureshi HS, Patterson JW and Wick MR. Solitary fibrous tumors of the skin: a clinicopathologic study of 10 cases and

- review of the literature. *J Cutan Pathol* 2007; 34:844-850.
- [82] Dray MS, McCarthy SW, Palmer AA, Bonar SF, Stalley PD, Marjoniemi V, Millar E and Scolyer RA. Myopericytoma: a unifying term for a spectrum of tumours that show overlapping features with myofibroma. A review of 14 cases. J Clin Pathol 2006;59:67-73.
- [83] Mentzel T, Hugel H and Kutzner H. CD34positive glomus tumor: clinicopathologic and immunohistochemical analysis of six cases with myxoid stromal changes. J Cutan Pathol 2002;29:421-425.
- [84] Hatori M, Aiba S, Kato M, Kamiya N and Kokubun S. Expression of CD34 in glomus tumors. Tohoku J Exp Med 1997;182:241-247.
- [85] Weiss SW and Nickoloff BJ. CD-34 is expressed by a distinctive cell population in peripheral nerve, nerve sheath tumors, and related lesions. Am J Surg Pathol 1993;17: 1039-1045.
- [86] Gould VE, Moll R, Moll I, Lee I, Schwechheimer K and Franke WW. The intermediate filament complement of the spectrum of nerve sheath neoplasms. Lab Invest 1986;55:463-474.
- [87] Bhatnagar S, Banerjee SS, Mene AR, Prescott RJ and Eyden BP. Schwannoma with features mimicking neuroblastoma: report of two cases with immunohistochemical and ultrastructural findings. J Clin Pathol 1998; 51:842-845.
- [88] Johnson MD, Glick AD and Davis BW. Immunohistochemical evaluation of Leu-7, myelin basic-protein, S100-protein, glialfibrillary acidic-protein, and LN3 immunoreactivity in nerve sheath tumors and sarcomas. Arch Pathol Lab Med 1988;112: 155-160.
- [89] Buley ID, Gatter KC, Kelly PM, Heryet A and Millard PR. Granular cell tumours revisited. An immunohistological and ultrastructural study. Histopathology 1988;12:263-274.
- [90] Mazur MT, Shultz JJ and Myers JL. Granular cell tumor. Immunohistochemical analysis of 21 benign tumors and one malignant tumor. Arch Pathol Lab Med 1990;114:692-696.
- [91] Yamaguchi U, Hasegawa T, Hirose T, Chuman H, Kawai A, Ito Y and Beppu Y. Low grade malignant peripheral nerve sheath tumour: varied cytological and histological patterns. J Clin Pathol 2003;56:826-830.
- [92] Allison KH, Patel RM, Goldblum JR and Rubin BP. Superficial malignant peripheral nerve sheath tumor: a rare and challenging diagnosis. Am J Clin Pathol 2005;124:685-692.
- [93] Rodriguez FJ, Scheithauer BW, Abell-Aleff PC, Elamin E and Erlandson RA. Low grade malignant peripheral nerve sheath tumor with smooth muscle differentiation. Acta Neuropathol 2007;113:705-709.

- [94] Kindblom LG, Ahlden M, Meis-Kindblom JM and Stenman G. Immunohistochemical and molecular analysis of p53, MDM2, proliferating cell nuclear antigen and Ki67 in benign and malignant peripheral nerve sheath tumours. Virchows Arch 1995;427: 19-26.
- [95] Requena L, Grosshans E, Kutzner H, Ryckaert C, Cribier B, Resnik KS and LeBoit PE. Epithelial sheath neuroma: a new entity. Am J Surg Pathol 2000;24:190-196.
- [96] Fetsch JF, Laskin WB, Hallman JR, Lupton GP and Miettinen M. Neurothekeoma: an analysis of 178 tumors with detailed immunohistochemical data and long-term patient follow-up information. Am J Surg Pathol 2007;31:1103-1114.
- [97] Hornick JL and Fletcher CD. Cellular neurothekeoma: detailed characterization in a series of 133 cases. Am J Surg Pathol 2007;31:329-340.
- [98] Sachdev R and Sundram UN. Frequent positive staining with NKI/C3 in normal and neoplastic tissues limits its usefulness in the diagnosis of cellular neurothekeoma. *Am J Clin Pathol* 2006;126:554-563.
- [99] Hirose T, Tani T, Shimada T, Ishizawa K, Shimada S and Sano T. Immunohistochemical demonstration of EMA/Glut1positive perineurial cells and CD34-positive fibroblastic cells in peripheral nerve sheath tumors. Mod Pathol 2003;16:293-298.
- [100] Ariza A, Bilbao JM and Rosai J. Immunohistochemical detection of epithelial membrane antigen in normal perineurial cells and perineurioma. Am J Surg Pathol 1988; 12:678-683.
- [101] Fetsch JF and Miettinen M. Sclerosing perineurioma: a clinicopathologic study of 19 cases of a distinctive soft tissue lesion with a predilection for the fingers and palms of young adults. Am J Surg Pathol 1997;21: 1433-1442.
- [102] Hirose T, Scheithauer BW and Sano T. Perineurial malignant peripheral nerve sheath tumor (MPNST): a clinicopathologic, immunohistochemical, and ultrastructural study of seven cases. *Am J Surg Pathol* 1998; 22:1368-1378.
- [103] Saadat P, Cesnorek S, Ram R, Kelly L and Vadmal M. Primary cutaneous paraganglioma of the scalp. J Am Acad Dermatol 2006;54: S220-223.
- [104] Wasserman PG and Savargaonkar P. Paragangliomas: classification, pathology, and differential diagnosis. Otolaryngol Clin North Am 2001;34:845-862.
- [105] Flieder DB and Moran CA. Primary cutaneous synovial sarcoma: a case report. *Am J Dermatopathol* 1998;20:509-512.
- [106] Murphey MD, Gibson MS, Jennings BT, Crespo-Rodriguez AM, Fanburg-Smith J and Gajewski DA. From the archives of the AFIP:

- Imaging of synovial sarcoma with radiologic-pathologic correlation. *Radiographics* 2006; 26:1543-1565.
- [107] van de Rijn M, Barr FG, Xiong QB, Hedges M, Shipley J and Fisher C. Poorly differentiated synovial sarcoma: an analysis of clinical, pathologic, and molecular genetic features. Am J Surg Pathol 1999;23:106-112.
- [108] Fisher C. Synovial sarcoma. Ann Diagn Pathol 1998;2:401-421.
- [109] Folpe AL, Schmidt RA, Chapman D and Gown AM. Poorly differentiated synovial sarcoma: immunohistochemical distinction from primitive neuroectodermal tumors and highgrade malignant peripheral nerve sheath tumors. Am J Surg Pathol 1998;22:673-682.
- [110] Machen SK, Fisher C, Gautam RS, Tubbs RR and Goldblum JR. Utility of cytokeratin subsets for distinguishing poorly differentiated synovial sarcoma from peripheral primitive neuroectodermal tumour. *Histopathology* 1998;33:501-507.
- [111] Ansai S, Takeda H, Koseki S, Hozumi Y and Kondo S. A patient with rhabdomyosarcoma and clear cell sarcoma of the skin. J Am Acad Dermatol 1994;31:871-876.
- [112] Seike T, Matsumoto K, Nakanishi H, Hashimoto I, Kubo Y and Arase S. Malignant melanoma of soft parts (clear cell sarcoma): a case report. J Dermatol 2003;30:550-555.
- [113] Swanson PE and Wick MR. Clear cell sarcoma. An immunohistochemical analysis of six cases and comparison with other epithelioid neoplasms of soft tissue. Arch Pathol Lab Med 1989;113:55-60.
- [114] Blessing K, Sanders DS and Grant JJ. Comparison of immunohistochemical staining of the novel antibody melan-A with S100 protein and HMB-45 in malignant melanoma and melanoma variants. *Histopathology* 1998;32:139-146.
- [115] Kaufmann O, Koch S, Burghardt J, Audring H

- and Dietel M. Tyrosinase, melan-A, and KBA62 as markers for the immunohistochemical identification of metastatic amelanotic melanomas on paraffin sections. *Mod Pathol* 1998;11:740-746
- [116] Zarbo RJ, Gown AM, Nagle RB, Visscher DW and Crissman JD. Anomalous cytokeratin expression in malignant melanoma: one- and two-dimensional western blot analysis and immunohistochemical survey of 100 melanomas. *Mod Pathol* 1990;3:494-501.
- [117] Guillou L, Wadden C, Coindre JM, Krausz T and Fletcher CD. "Proximal-type" epithelioid sarcoma, a distinctive aggressive neoplasm showing rhabdoid features. Clinicopathologic, immunohistochemical, and ultrastructural study of a series. *Am J Surg Pathol* 1997;21:130-146.
- [118] Daimaru Y, Hashimoto H, Tsuneyoshi M and Enjoji M. Epithelial profile of epithelioid sarcoma. An immunohistochemical analysis of eight cases. *Cancer* 1987;59:134-141.
- [119] Manivel JC, Wick MR, Dehner LP and Sibley RK. Epithelioid sarcoma. An immunohistochemical study. Am J Clin Pathol 1987;87: 319-326.
- [120] Lin L, Skacel M, Sigel JE, Bergfeld WF, Montgomery E, Fisher C and Goldblum JR. Epithelioid sarcoma: an immunohistochemical analysis evaluating the utility of cytokeratin 5/6 in distinguishing superficial epithelioid sarcoma from spindled squamous cell carcinoma. J Cutan Pathol 2003;30:114-117.
- [121] Arber DA, Kandalaft PL, Mehta P and Battifora H. Vimentin-negative epithelioid sarcoma. The value of an immuno-histochemical panel that includes CD34. *Am J Surg Pathol* 1993;17:302-307.
- [122] Fisher C. Epithelioid sarcoma of Enzinger. Adv Anat Pathol 2006;13:114-121.