

## Original Article

# Piwil2 is expressed in various stages of breast cancers and has the potential to be used as a novel biomarker

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**Abstract:** Piwil2, a member of AGO/PIWI family of proteins, has been reported to be expressed in precancerous stem cells (pCSCs), tumor cell lines and various types of human cancers. However, the significance of piwil2 expression in breast cancer has not been investigated. In this study, archival formalin-fixed, paraffin-embedded breast cancer specimens at various developmental stages were prepared as tissue microarrays (TMAs) and examined for the expressions of piwil2, estrogen receptor (ER), progesterone receptor (PR) and a cell proliferation marker Ki67 by immunohistochemical (IHC) staining and human epidermal growth factor receptor 2 (HER2) by fluorescence in situ hybridization (FISH). The correlation of piwil2 expression with ER, PR and Ki67 were analyzed statistically. The piwil2 was detected in all of breast cancer TMA cores. In contrast, ER, PR, HER2, and Ki67 were detected only in 66.1%, 54.5%, 36.0%, and 47% of the TMA cores, respectively. Piwil2 was expressed in cytoplasm (Cyt), nucleus (N) or both cytoplasm and nucleus (C-N). The N pattern was less observed in breast precancers, whereas all three patterns were observed in invasive and metastatic cancers. While the Cyt pattern was significantly correlated with ER expression ( $p = 0.002$ ); N pattern was significantly correlated with Ki67 expression ( $p = 0.001$ ). ER and Ki67 expressions were reduced and increased, respectively, with the expression patterns being shifted from Cyt → C-N → N. In conclusion, piwil2 is expressed in various stages of breast cancers and has the potential to be used a novel biomarker.

**Keywords:** Piwil2, breast cancer, precancer, estrogen receptor, progesterone receptor, HER2, and Ki67, field cancerization

## Introduction

Breast cancer is one of leading causes of women death in the Untied Sates and occurs in about 13% (1/8) of women [1]. Generally, cancer development may undergo the stages of benign proliferation (hyperplasia and low grade dysplasia), precancer (high grade dysplasia and carcinoma *in situ*) and cancer (invasive and metastatic carcinoma) [2,3]. Breast cancer if detected early is curable by prevention or intervention of progression of precancer to invasive and metastatic types [4,5]. Thus, specific biomarker(s) that are expressed in breast precancerous lesions [atypical hyperplasia and carcinoma *in situ* (CIS)] are required for the early detection of

breast cancers. A challenge for the early detection is the current lack of specific biomarkers expressed at the early stages of cancer development. Despite the identification of a number of oncogenes (ONGs) and tumor suppressor genes (TSGs) in breast cancers, most of them are not appropriate to be used as breast cancer biomarkers, because they are redundantly expressed in normal mammary glands and required as well for the control of normal cell cycle, cell growth and/or cell survival [6,7]. Therefore, a critical issue for early detection of breast cancer is to explore biomarkers that are specifically expressed at precancerous or both precancerous and cancerous stages of breast tumors.

The *PIWIL2* gene (alias *mili* in mouse or *hili* in humans) is a member of the P-element-induced wimpy testis/Argonaute (PIWI/AGO) gene subfamily, which is essential for germ-cell development [8-12]. PIWI/AGO genes contain Piwi and PAZ domain (PPD), playing important roles for stem cell self-renewal in *drosophila* [13], gametogenesis [9], small RNA-mediated gene silencing [14,15] and/or chromatin remodeling [16,17]. Recently, *piwil2* was found to bind a novel class of RNA called piwi-interacting RNA (piRNA) or repeat-associated small interfering RNAs (rasiRNAs), in mammal testis [18-23]. It may silence the selfish genetic elements, such as retrotransposons, in the germline stem cells (GSCs) of testis [19,23,24]. Dysregulated piwi protein expression appears to be associated with tumorigenesis [25-27]. Thus, *piwil2* might play an important role in tumor development [2,27].

Recently, *piwil2* was reported to be expressed in tumor cell lines and various types of human cancer [26]. We have also found that *piwil2* transcripts were ectopically expressed in murine precancerous stem cells (pCSCs) [2,3,27,28], suggesting that *piwil2* might be a biomarker for cancer development. Therefore, we examined *piwil2* expression in various stages of human breast cancer and its association with other known breast cancer biomarkers, such as estrogen receptor (ER), progesterone receptor (PR), human epidermal growth factor receptor 2 (HER2/Neu or Erbb-2), and cell proliferation marker Ki67 (also known as MKI67), a cellular marker for proliferation [29]. The results indicate that *piwil2* is expressed in various stages of breast cancer. Especially, it can be detected in breast precancers and its expression pattern is associated with ER and Ki67 expression. The findings suggest a potential for *piwil2* to be developed into a novel biomarker of breast cancer.

## Material and methods

### Specimens and reagents

This study was approved by the Institutional Review Board (IRB #2007E0686) at the Ohio State University (OSU). One hundred and twenty six breast cancer specimens at various developmental stages were obtained from the Tissue Procurement Shared Resource (TPSR), Comprehensive Cancer Center, Ohio State University

**(Table 1).** Tumor specimens were fixed in 10% formalin and embedded in paraffin for pathological and immunohistochemical analysis. Tissue microarrays (TMAs) with 2 mm cores were built by the Histological Core Facility, Department of Pathology. Normal mammary specimens (n=14) from the patients without breast cancer were used as negative controls for *piwil2* expression.

Polyclonal rabbit anti-*mili* antibody was generated and purified as previously described [9]. mAbs to ER (1D5), PR (PgR636) and Ki67 (MIB-1) were purchased from Dako (Carpinteria, California, USA).

### Histological and immunohistochemical (IHC) analysis

IHC analysis was performed as previously described [27,28,30]. TMAs were examined for *piwil2*, ER, PR and Ki67 by immunohistochemistry (IHC). HER2 expression was determined by fluorescence in situ hybridization (FISH). Sections (4 ~ 5  $\mu$ m thick) were stained by H. & E. for pathological analysis, or immunostained with a primary antibody to *mili* (*piwil2*), ER, PR, or Ki67 followed by a horseradish peroxidase (HRP)-conjugated secondary antibody. The immunostained sections were counterstained with hematoxylin. The specimens were analyzed based on the staining score and staining patterns (Cyt, N and N-Cyt). The staining score (0 ~ 3) was determined blindly by two pathologists: 0 (-) or negative, 0 ~ <5% *piwil2*<sup>+</sup> cells; 1 (+) or weak, 5 ~ 25% *piwil2*<sup>+</sup> cells; 2 (++) or medium, 25% ~ 50% *piwil2*<sup>+</sup> cells; 3 (+++) or strong, > 50% *piwil2*<sup>+</sup> cells. ER<sup>+</sup>, PR<sup>+</sup>, and Ki67<sup>+</sup> cells were also scored as for *piwil2*<sup>+</sup> cells. HER2 is based on clinical positive or negative diagnosis.

### Statistical analysis

McNemar Test was used to compare the expression in percentage between *piwil2* and ER, PR, HER2 or Ki67 in breast cancer. This test is used to compare two paired measurements from the same subject. When the sample size is large, the McNemar test follows the same  $\chi^2$  distribution but uses a slightly different formula [31]. Fisher's Exact Test was used to compare difference of *piwil2* expression patterns between various types of breast lesions. The correlation of *piwil2* expression with ER, PR or Ki67 expression, the correlation between ER, PR and Ki67,

and the correlation between each piwil2 expression pattern and ER, PR or Ki67 expression were analyzed by a least-squares linear regression using SAS statistical software (SAS Institute Inc, NC, USA). A value of  $p \leq 0.05$  was considered significant. Data are represented as mean  $\pm$  SD. \*,  $p \leq 0.05$ ; \*\*,  $p \leq 0.01$ .

## Results

Archival formalin-fixed, paraffin-embedded breast cancer tissues ( $n=126$ ) were prepared as tissue microarrays (TMA), and analyzed for piwil2 expression by IHC staining (Table 1). Although these specimens were all procured from the patients with breast cancers, the TMA cores might not be consistent with the clinical diagnosis based on regular histological sections because of multiple primary tumors or field cancerization [32], which might cause technical error. Therefore, the TMA cores were double-blindly re-examined by two pathologists. While six cores from the patients with invasive ductal

**Table 1.** Clinicopathological diagnosis and piwil2 expression patterns

Pathological Diagnosis	Cyt	N	C-N	No.
DH	+	ND	ND	1
ADH	ND	ND	+	1
DCIS	+	-	+	9
IDC	+	+	+	76
ILC	+	+	+	7
MET	+	+	+	7
MET SqCC	ND	ND	+	1
MED CA	ND	ND	+	1
MIC CA	+	ND	ND	1
SRCC	+	ND	ND	1
TC	+	ND	ND	1
IDC+DCIS	+	-	+	14
Benign	+	+/-	+	6
Normal	-	-	-	14

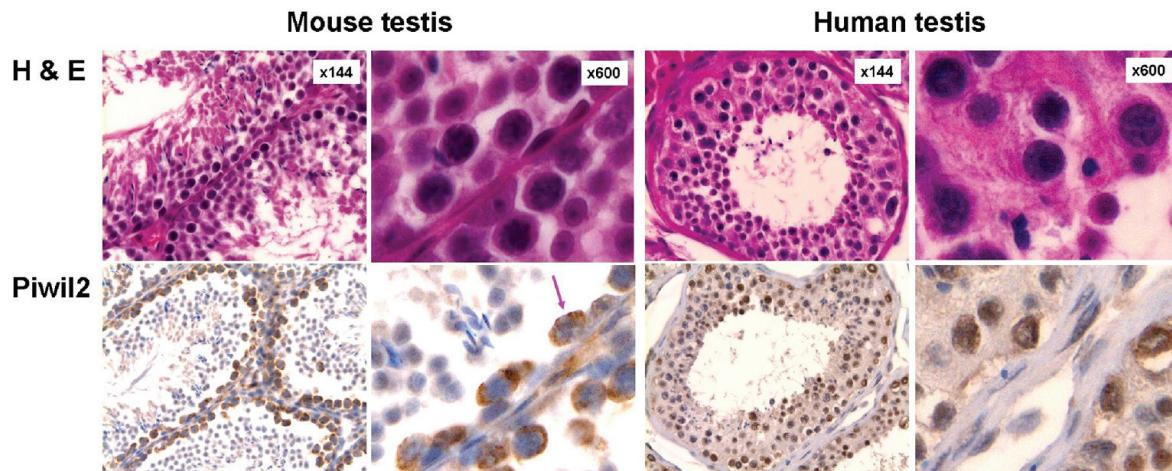
ADH: Atypical Ductal Hyperplasia; DH: Ductal Hyperplasia; DCIS: Ductal Carcinoma In Situ; IDC: Invasive Ductal Carcinoma; IDC + DCIS: IDC plus DCIS; ILC: Invasive Lobular Carcinoma; MET: Metastatic; MET SqCC: Metastatic Squamous Cell Carcinoma; MIC CA: Micropapillary Carcinoma; MED CA: Medullary Carcinoma; TC: Tubular Carcinoma; SRCC: Signet ring cell; ND: not determinable. Benign: Histological normal epithelial cell or tissue within tumors. Normal: patients without breast tumors.

carcinoma exhibited histologically normal appearance, and others ( $n=120$ ) exhibited consistent lesions with clinicopathological diagnosis, including ductal hyperplasia (DH), atypical ductal dysplasia (ADH), ductal carcinoma in situ (DCIS), invasive ductal or lobular carcinoma (IDC or ILC), and metastatic carcinoma (Met-ca) (Table 1).

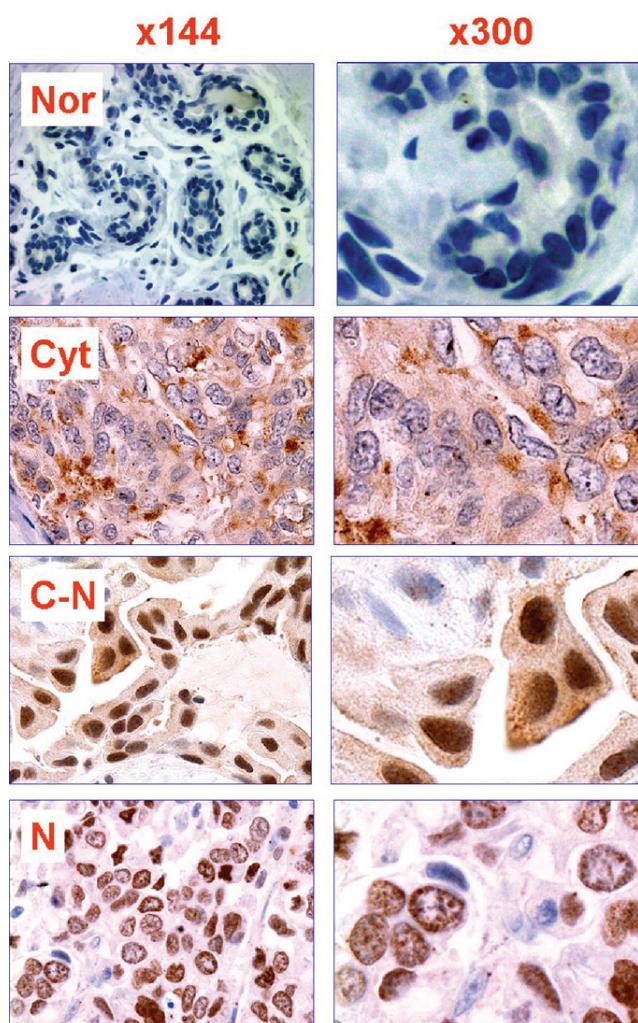
Polyclonal rabbit anti-mili antibody, which was generated as previously described [9] was used to detect piwil2, because it detected not only mili in the GSCs of murine testis but also hili in the GSCs of human testis (Figure 1). The piwil2 was detected essentially in the cytoplasm (Cyt) of murine GSCs with fine granules in some nuclei (N) but mainly in the nuclei or both cytoplasm and nucleus (C-N) of human GSCs. In the human testis, piwil2 was also detected in hyperplastic testicular cells (Figure 1). Similarly, piwil2 expression patterns of Cyt, N and N-C were also observed in breast cancers, but not in normal mammary tissues (Figure 2).

Piwil2 was detected in 126/126 TMA cores of breast cancers, regardless of their developmental stages (Figure 3 and Table 1). Among them, six cores exhibited normal histology. Since these specimens were all procured from the patients with breast tumors, the sensitivity for detection of piwil2 in breast tumors was 100% (Figure 3). However, the level of piwil2 expression in the breast lesions was highly variable with individuals rather than with the types of lesion (from about 5% to >50% of cancer cells) (Figure 4). To further exclude the false positivity of piwil2 expressed in some histologically normal cores, we further examined piwil2 expression in the normal breast tissues without hyperplasia ( $n=14$ ); none of them was detected with piwil2 (Figure 2 and not shown). The results suggest that piwil2 may express in various types and various stages of breast lesions but unlikely in normal mammary tissue.

To compare sensitivity of piwil2 as a breast biomarker with other known breast cancer markers, such as ER, PR, HER2 and ki67, breast cancer TMAs were immunohistochemically stained with anti-ER, PR or Ki67 antibody, and the data of HER2 detected by FISH was procured from clinicopathological records. The ER, PR, and Ki6 expression in the cores were consistent with clinicopathological data. In striking contrast to piwil2, ER, PR, HER2, and Ki67



**Figure 1.** Expression pattern of piwil2 in the testis. The murine testes were derived from C57BL/6 mice showing normal histology; and the human testes were derived from TPSR, Ohio State University, exhibiting hyperplasia. The piwil2 was detected essentially in the cytoplasm of murine germline stem cells with fine granular in nucleus, and in both cytoplasm and nucleus of human germline stem cells as well as in hyperplastic cells.

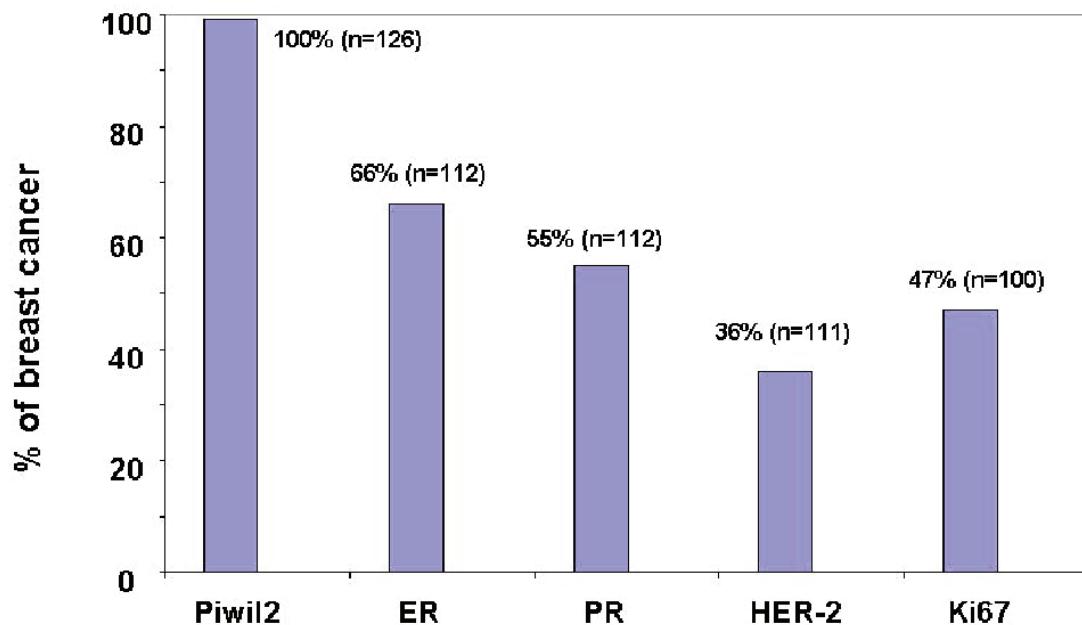


were detected only in 66.1 (74/112), 54.5% (61/112), 36.0% (40/111), and 47% (47/100) of breast cancer, respectively. The differences between groups were statistically significant ( $p < 0.0001$ ) (Figure 3). The total case reduction for each marker in the Figure 3 was due to TMA cores that dropped off slides during staining. These data suggest that piwil2 as a breast cancer biomarker is broader in expression than HER-2, ER, PR and Ki67. Because ER, PR, HER2 and Ki67 are required as well for normal cell development but piwil2 is silenced in adult tissues except in testis [8,10,26], piwil2 as a breast cancer biomarker is obviously more tumor-specific than HER2, ER, PR, and Ki67.

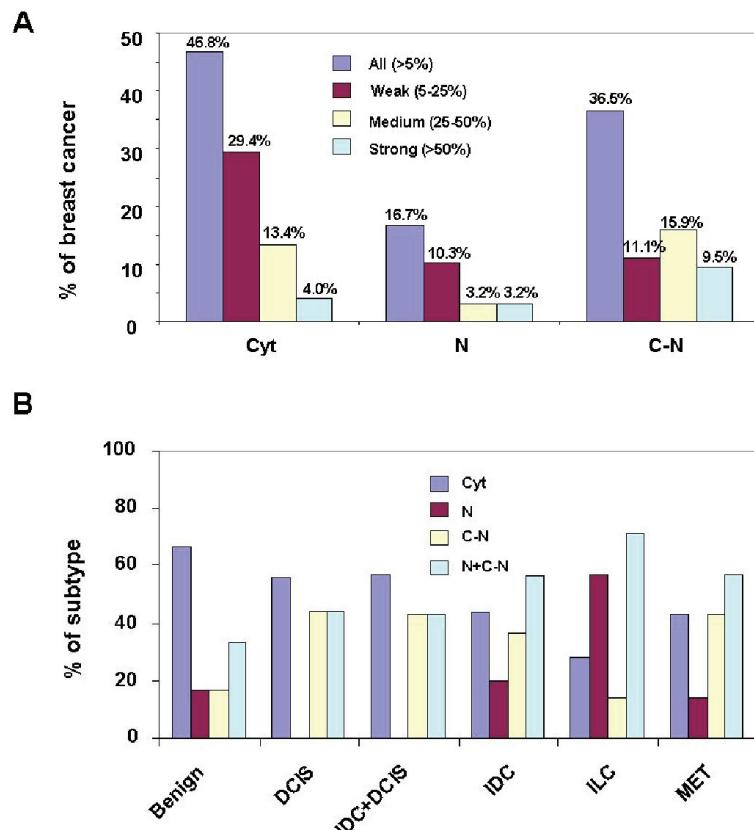
As mentioned above, three distinct expression patterns (Cyt, N and Cyt-N) of piwil2 in breast cancer were identified by IHC staining (Figure 2 & 4). The expression patterns were highly variable between individuals, because not all three patterns were observed in the same core. About 46.8%, 16.7% and 36.5% of the TMA cores was

**Figure 2.** Expression pattern of Piwil2 in human breast cancer. Typical staining patterns of Piwil2 in human breast cancer include cytoplasmic (Cyto), nuclear (N) and nuclear and cytoplasmic (C-N). The examples of staining patterns are derived from the tissues of IDC, which were stained with rabbit anti-mili IgG. Nor: normal mammary tissue.

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**Figure 3.** Expression of piwil2 in breast cancer. The breast cancer TMAs were stained with antibodies to piwil2, ER, PR, and Ki67. The TMA cores with >5% positive cells were considered as positive one for each marker. The data of HER2 were derived from clinicopathological records of the patients with breast cancer and revealed by FISH. The percentage of piwil2 expression is significantly higher than that of ER, PR, HER2 or Ki67, as determined by McNemar Test. The p-value of all comparisons (piwil2:ER, PR, HER2 or Ki67) are < 0.0001.



**Figure 4.** Piwil2 expression pattern in various stages of breast cancer. Piwil2 can be detected in cytoplasm (Cyt), nucleus (N) or both cytoplasm and nucleus (C-N), as demonstrated in Figure 2. Each pattern was scored as weak (5-25%), medium (25-50%), and strong (>50%), based on the percentage of piwil2-expressing cells. A, proportion of each piwil2 expression pattern in breast cancer (n=126). B, distribution of piwil2 expression pattern in various types of breast cancer: Benign (histological normal tissue within breast cancer): n=6; DCIS: n=9; DCIS + IDC: n=14; IDC: n=76; ILC: n=7; MET (metastatic carcinoma) n=7. N + C-N: the summation of N and C-N pattern. The percentage between groups is not significantly different as tested by Fisher's Exact Test.

expressed with the pattern of Cyt, N and C-N, respectively (**Figure 4A**). Interestingly, the expression patterns of piwil2 appeared to be associated with the developmental stages of breast cancer (**Figure 4B**). In the histologically normal cores, Cyt pattern was dominant compared to breast precancer and invasive/metastatic cancer. The N pattern was almost absent in ductal carcinoma in situ (DCIS) and invasive ductal carcinoma (IDC) + DCIS (both IDC and DCIS were detected in the same cores), and all three patterns were detected in IDC, invasive lobular carcinoma (ILC) and metastatic cancers (MET) (**Figure 4B & Table 1**). In addition, piwil2 was also detected in ductal hyperplasia (DH), atypical ductal hyperplasia (ADH) and other histological types of breast cancer, although the sample size is very small (**Table 1**). Although the percentage of the N + C-N pattern was lower in precancers (benign lesions: 33.4%, DCIS: 44.44% and IDC + DCIS: 42.86%) than in invasive/metastatic cancers (IDC: 56.58%; ILC: 71.39%; and MET: 57.15%), the difference was not statistically significant, as tested by Fisher's Exact Test (**Figure 4B**).

To determine the significance of piwil2 expression patterns in breast cancer development, we statistically analyzed the correlation of piwil2 with the prognostic markers of breast cancer including ER, PR or Ki67. As shown in **Figure 5**, Cyt pattern of piwil2 expression was positively correlated with ER and PR expressions, especially significantly correlated with ER expression ( $p = 0.002$ ), but inversely correlated with Ki67 expression ( $p = 0.03$ ) (**Figure 5A**). In contrast, N pattern of piwil2 expression was positively correlated with Ki67 expression ( $p = 0.001$ ) and had no correlation with ER and PR expression (**Figure 5A**). C-N pattern of piwil2 expression appeared not to be correlated significantly with ER, PR and Ki67 expression. Interestingly, the coefficient of correlation was decreased between piwil2 and ER but increased between piwil2 and Ki67, with the expression pattern being shifted from Cyt → C-N → N, suggesting an important role of piwil2 in regulation of ER and Ki67 expression in breast cancer (**Figure 5B**).

## Discussion

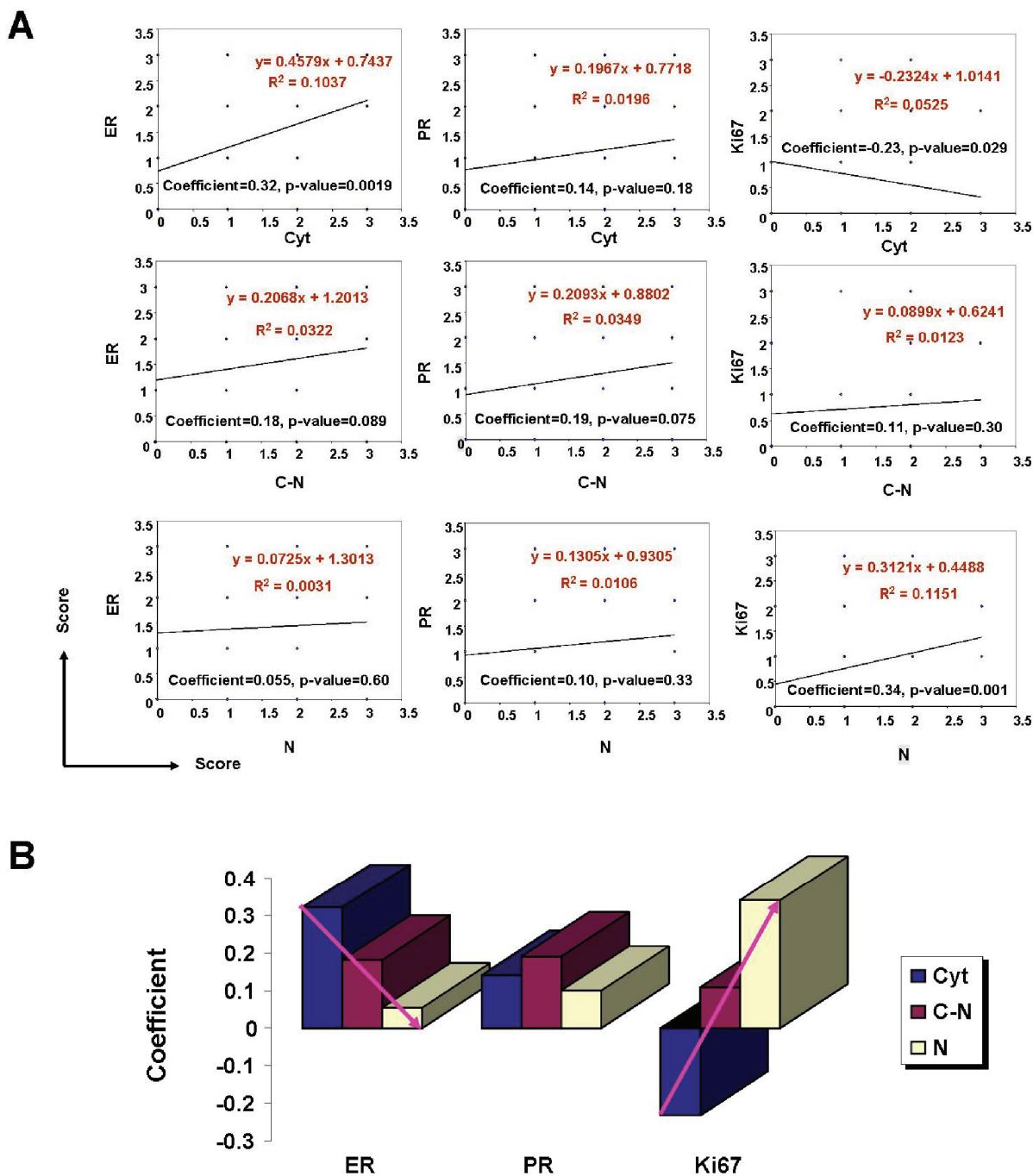
Identification of biomarkers for detection of breast cancer is a critical issue for the treatment and cure of breast cancer [4,5]. To reach the goal, it is a necessary step to identify a bio-

marker widely expressed in various developmental stages of breast cancer, especially at the initial stage of cancer development [4,33]. Recently, it has been demonstrated that piwil2 is widely expressed in various types of human cancers [26], but not in normal tissues [2,8,26]. The significance of piwil2 expression in breast cancer, however, has not been defined.

In this study, Archival formalin-fixed, paraffin-embedded breast cancer tissues were prepared for tissue microarray (TMA), and analyzed for piwil2, ER, PR and Ki67 expression by IHC staining as well as HER2 by FISH. The correlation of piwil2 expression with ER, PR, and Ki67 was also analyzed statistically. Our results indicate that the piwil2 was not expressed in normal mammary tissues but in all breast cancer TMA cores (100%), including some histologically normal cores derived from the tissues adjacent to carcinoma, precancerous cores, and invasive/metastatic cancers. In contrast, ER, PR, HER2 and Ki67 were detected only in 66.1 (74/112), 54.5% (61/112), 36.0% (40/111), and 47% (47/100) of breast cancers, respectively. While ER expression was significantly correlated with PR expression ( $p < 0.001$ ), there was no significant expression correlation between piwil2 and ER, PR, or HER2 ( $p > 0.05$ ); however, the expression pattern of piwil2 was significantly correlated with ER and Ki67 expressions. Three expression patterns of piwil2 were observed in breast cancer cells, including cytoplasmic (Cyt), nuclear (N) and cytoplasmic and nuclear expression (C-N). The Cyt pattern of piwil2 was positively correlated with ER expression ( $p = 0.02$ ); whereas the N pattern was positively correlated with Ki67 expression ( $p = 0.001$ ). Interestingly, the expression pattern of piwil2 from Cyt → C-N → N was associated with the decreased expression of ER but with the increased expression of Ki67. N pattern of piwil2 expression was less observed in breast precancers. The results suggest an important role of piwil2 for breast cancer development. Because piwil2 can be expressed at the initiation stages of breast tumors, it is likely that piwil2 could be further developed into a biomarker for diagnosis and/or prognosis of breast cancers when complemented with other biomarkers and a therapeutic target for cure of breast cancers. In addition, piwil2 might be an ideal target for breast cancer therapy, because it is expressed in all stages of breast tumors.

Generally, a cancer may undergo the develop-

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**Figure 5.** Correlation of piwi2 expression pattern with the expressions of ER, PR and Ki67. Breast cancer TMA cores ( $n=126$ ) were immunohistochemically stained with antibody to mili, ER, PR, or Ki67. The intensity of staining for each marker was scored as negative (0); weak (1), moderate (2), and strong (3). The correlation of piwi2 expression patterns with ER, PR and Ki67 was determined by least-squares linear regression. A, correlation of each piwi2 expression pattern with ER, PR, or Ki67; B, Correlation coefficient between piwi2 expression patterns and ER, PR or Ki67. Cyt: cytoplasm; N: nucleus; C-N: both cytoplasm and nucleus. The red arrows indicate the trends of correlation coefficient between piwi2 expression and ER or Ki67 expression in breast cancer.

mental stages of benign proliferation (initiation), precancer and malignant cancer [2,3,34-36], which is supposed corresponding to pathological changes of hyperplasia and atypical hyperplasia (benign proliferations), dysplasia and carcinoma in situ (precancer), and invasive / metastatic carcinoma (malignant cancer) [2,3,36,37]; and is mediated by tumor stem cells (TSCs), including tumor initiating stem cells (TISCs), pCSCs, and cancer stem cells (CSCs) [2,3,27,28]. Tumor initiation stage and precancer stage are highly reversible and thus are the ideal targets for cancer prevention and therapy [2,3,34,38]. In this study, piwil2 was detected in breast benign proliferations [DH and histological normal tissues within cancer (benign)], precancers (ADH and DCIS), and malignant cancers (IDC, ILD and MET), suggesting that *PIWIL2* can be activated as early at the initial stage (benign proliferation) of breast cancer. This feature is of importance for piwil2 to be developed into a biomarker for detection, prevention and therapy of breast cancer. One potential concern is that detection of piwil2 in the histologically normal tissues within malignant breast cancers might lead to false-positivity in practice. This issue needs to be further addressed with a large cohort of patients, despite unlikely to happen.

Detection of piwil2 in the histologically normal tissues adjacent to cancers might be related to field cancerization [39,40] or “field effects” of cancer, which may reflect the precancerous epigenetic alteration of normal epithelial cells or mammary stem cells [41,42], or the seeding of breast tumor cells such as pCSCs and CSCs in the distant areas [3,43]. In fact, the detection of piwil2 in the histologically normal tissues further confirms the “field effect” of epigenetic and oncogenetic alterations in breast cancer [41,42]. Because piwil2 transcripts are constitutively expressed in pCSCs [27], it is likely that the piwil2-expressing “normal” tissues surrounding or within malignant breast cancer reflect that the precancerous lesions occur at molecular levels rather than at cellular and/or histological levels [3,38]. The specificity of piwil2 for early breast lesions was further supported by the failure to detect piwil2 in the normal mammary tissues without hyperplastic lesions, which were derived from the patients with lumpectomy. Because the case number for normal mammary tissues was relatively small, further experiments are warranted to validate the sensitivity and specificity of piwil2 for detection of

breast cancers.

High sensitivity and high specificity are essential criteria for a biomarker to be used for detection of cancer. Currently, none of the known breast cancer biomarkers such as ER, PR, HER2, BRST2 [anti-gross cystic disease fluid protein (GCDFP15)] and Ki67 meets the criteria. HER2, ER, PR, BRST2 or Ki67 have been used in clinic as complementary markers for prognosis of breast cancer [4,44,45]. However, these markers are not appropriate for diagnosis of breast cancer. In this study, piwil2 was detected in all TMA cores of breast cancers with various developmental stages but not in normal mammary tissues, suggesting that piwil2 as a biomarker is more sensitive and specific than other known breast cancer biomarkers. Thus, piwil2 has the potential of developing into a novel biomarker for detection of various stages of breast cancers.

Based on the expression of HER2, ER and PR, breast cancer can be categorized by IHC staining into the following subtypes: luminal A: ER<sup>+</sup> or PR<sup>+</sup> and HER2<sup>-</sup>; luminal B: ER<sup>+</sup> or PR<sup>+</sup> and HER2<sup>+</sup> (triple positive); HER2: ER<sup>-</sup> and PR<sup>-</sup> and HER2<sup>+</sup>; and basal: ER<sup>-</sup> and PR<sup>-</sup> and HER2<sup>-</sup> (triple negative) [46,47]. Although the classification is useful for determining therapeutic mode for breast cancer patients, it can not be used to precisely predict progression of a breast cancer. Piwil2 might also have the potential to be used as a novel marker for breast cancer classification when complemented with other biomarkers. In this study, we demonstrate that the frequency and patterns of piwil2-expressing cells are individually variable in breast cancer. This might reflect differential developmental status of breast cancer between individuals. The association of piwil2 expression with ER and Ki67 expression further suggests that piwil2 expression pattern might be an indicator of tumor progression. For example, the shift of piwil2 expression pattern from Cyt → C-N → N may lead to an increase of Ki67-expressing cells in breast cancer; as a consequence, tumor growth is accelerated. Interestingly, N pattern was not detected in the TMA cores of DCIS or DCIS + IDC. Thus, the N pattern might be a demarcation to distinguish true IDC from false IDC. The hypothesis needs to be verified in a larger cohort of breast cancer patients.

Our studies have demonstrated that piwil2 is

ubiquitously and uniquely expressed in various stages of breast cancers and its expression patterns are associated with ER and Ki67 as well as cancer development, suggesting that piwi2 plays an important role in breast cancer development. Therefore, piwi2 can be targeted for the study of the mechanisms underlying breast cancer development and has the potential to be developed into a novel biomarker for detection, prognosis and therapy of breast cancers.

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