Original Article CREPT expression correlates with poor prognosis in patients with retroperitoneal leiomyosarcoma

Yaoguang She¹, Jiao Liang², Lin Chen², Ying Qiu², Na Liu¹, Xudong Zhao¹, Xiaohui Huang¹, Yinyin Wang², Fangli Ren², Zhijie Chang², Peiyu Li¹

¹Department of General Surgery, Chinese PLA General Hospital, Beijing 100853, China; ²State Key Laboratory of Biomembrane and Membrane Biotechnology, School of Medicine, National Engineering Laboratory for Anti-tumor Therapeutics, Tsinghua University, Beijing 100084, China

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Abstract: Retroperitoneal leiomyosarcomas (LMSs) are rare gynecological malignancies that display poor prognosis and high mortality. Cell cycle-related and expression-elevated protein in tumor (CREPT) is an oncogene that is involved in the regulation of many cell cycle-related proteins. However, its distribution and clinical significance in retroperitoneal LMS remains poorly understood. This study assessed the histological classifications of postoperative tumor samples from 71 cases of retroperitoneal LMS that were collected at The General Hospital of the People's Liberation Army from January 1998 to December 2012. We found that more than half of the patients displayed positive expressions of CREPT, Ki-67 and PCNA via immunohistochemical analysis. The expression of CREPT correlated with histological grade (P = 0.044), and the PCNA expression level correlated with the differentiation of tumor cells and histological grade (P < 0.001 and P = 0.009, respectively). Multivariate analysis showed that survival was associated with histological grade and the expression level of CREPT (P = 0.011 and P = 0.012, respectively). Kaplan-Meier analysis showed that the patients lacking CREPT expression exhibited significantly longer overall postoperative survival (median, 60.0 months) than the patients displaying CREPT expression (median, 33.0 months), and CREPT expression correlated with distant recurrence within 5 years after surgery (P = 0.004). Western blot analyses showed that CREPT was more strongly expressed in the retroperitoneal LMS tumor tissue than in paired control tissue. Based on the above data, we concluded that CREPT displays unique immunostaining for retroperitoneal LMS tissue and can be used to supplement other currently available retroperitoneal LMS markers.

Keywords: CREPT, retroperitoneal neoplasms, leiomyosarcoma, prognosis

Introduction

The retroperitoneum can host a wide spectrum of pathologies, including a variety of benign and malignant neoplasms [1]. Retroperitoneal tumors can create a diagnostic dilemma and present several therapeutic challenges because of their rarity, relatively late presentation and anatomical location, which is often proximal to several vital structures in the retroperitoneal space [2]. The prognosis of patients with retroperitoneal sarcoma is poor with a 12-40% overall5-yearsurvival rate [3]. Leiomyosarcomas (LMSs), a type of retroperitoneal sarcoma, are the 2nd most common primary retroperitoneal neoplasm [4]. The most effective treatment for this disease is generally considered to be surgery, but less than 50% of these patients receive this treatment [5]. Failure to accomplish complete excision is attributed to several factors: the size of the tumor, its location and the number of organs involved [6]. Consequently, the survival of patients diagnosed with retroperitoneal neoplasms is very low, and there is an urgent need to understand the pathogenesis of retroperitoneal neoplasms and to develop new diagnostic markers and treatment modalities.

CREPT (cell-cycle related and expression-elevated protein in tumor, also named RPR1B) is a novel gene that belongs a new family of proteins within the RPR domain and was recently identified to promote tumorigenesis by up-regulating the expression of genes related to the cell cycle. Our group revealed a mechanism through which CREPT promotes cell proliferation by enhancing the transcription of CYCLIN D1 via preventing RNAPII from "reading" through" and possibly promoting the recycling of RNAPII to the promoter of genes; this mechanism occurs in a manner similar to that of the chromatin loop. We previously identified that CREPT is highly expressed in several types of gastroenteric tumors based on immunohistochemistry (IHC) analysis. We demonstrated that stomach cancer patients displaying strong CREPT expression exhibited a poor survival rate after surgery [7]. However, in retroperitoneal LMS, the relationship between CREPT expression and prognosis (particularly that relating to survival) remains unclear.

In the present study, we expanded our immunohistochemical analysis of the expression of CREPT, Ki-67 and PCNA in a set of retroperitoneal LMS samples to further explore the diagnostic value of these markers. We also investigated the relationship between the expression of CREPT and clinical prognosis and the clinicopathological characteristics of retroperitoneal LMS. Our data showed that CREPT could be used as a potential marker for the prognosis of retroperitoneal LMS.

Patients and methods

Patients and tissue samples

A set of 71 retroperitoneal LMS patients who underwent curative surgery without preoperative chemotherapy or radiotherapy were selected for this study from January 1998 to December 2012 at The General Hospital of the People's Liberation Army (PLA) in Beijing, China. The histological diagnosis of retroperitoneal LMS was established and confirmed by two pathologists. Patients displaying GISTs were excluded from the analysis. The baseline clinical and staging data were retrieved from the hospital database for reviewing.

There were 71 retroperitoneal LMS patients; 57 females and 14 males (4:1) ranging from 21 to 79 years of age (median, 48 years). At the time of surgical resection, the tumors ranged in size from 3.5 to 45 cm (median, 12.9 cm). Based on the Federation National des Centres de Lutte contre le Cancer (FNCLCC) standards [8], 48 patients (67.6%) were histological grade II, 63 (88.7%) contained moderately differentiated tumors, and 39 (54.9%) displayed low mitotic counts. A total of 64 (90.1%) retroperitoneal LMS patients with available follow-up data were selected to generate the Kaplan-Meier survival curves. Additionally, 5 paired fresh frozen samples that included the retroperitoneal LMS and adjacent noncancerous tissues were collected for Western blot analysis. The specimen collection and study procedures were approved by The Ethics Committee of the Chinese PLA General Hospital.

IHC and staining evaluation

The paraffin-embedded tumor tissues were sliced into 3 µm sections and deparaffinized. The sections were heated in a microwave oven for antigen retrieval, and a standard streptavidin/peroxidase complex method (SP) was used for immunostaining, as previously described [9]. The monoclonal mouse anti-human CREPT (1:60) and monoclonal rabbit anti-human Ki-67 (1:300) and PCNA antibodies (1:1,000) (Santa Cruz, USA) were used as primary antibodies. After counterstaining with Meyer's hematoxylin, the sections were observed under a light microscope.

All immunohistochemically stained sections were examined in a blinded manner without any knowledge of the clinicopathological parameters or patient outcomes.

The immunoreactivity for CREPT, Ki-67 and PCNA was recorded as strong or weak based on the staining intensity score and the percentage score. The proportion score was assigned according to the percentage of the tumor cells displaying positive nuclear staining (0, < 10%); 1, 11-30%; 2, 31-80%; or 3, > 80%). The intensity score was assigned according to the average intensity of the immunopositive tumor cells (0, none; 1, weak; 2, moderate; or 3, strong). The expression score was calculated using the percentage and intensity scores, which ranged from 0 to 9. The expression levels were categorized as negative (score 0), 1+ (score 1-3), 2+ (score 4-6) and 3+ (score 7-9). Any positive expression level (from 1+ to 3+) was regarded as positive expression.

Western blot analysis

Total protein samples from both tumor and adjacent normal tissues were extracted using

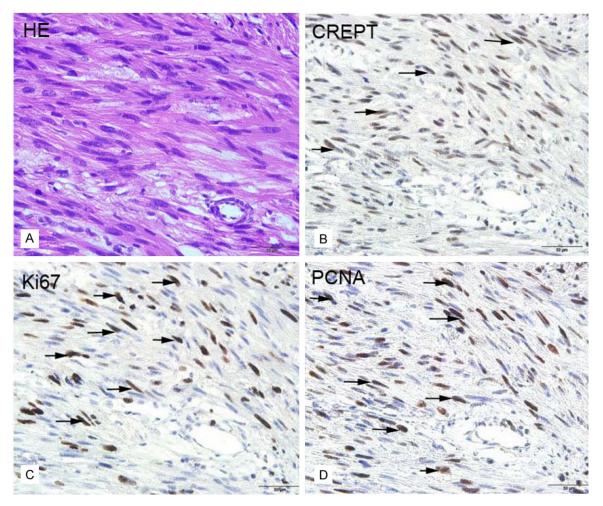


Figure 1. Representative results of the H&E and immunohistochemical staining for CREPT, Ki-67 and PCNA in retroperitoneal LMS tissues. A: H&E staining. B-D: Immunohistochemical staining for CREPT, Ki-67 and PCNA, respectively. The arrows indicate positively stained cells. (Original magnification × 400, bar = 50 µm).

RIPA buffer, and the protein concentration of each sample was determined using the Bradford method (Thermo). Equal amounts of protein (40 ig) were separated via SDS-PAGE and transferred to a PVDF membrane (Millipore, Bedford, MA, USA). The membrane was blocked in 8% non-fat dry milk for 1 h at 37°C prior to incubation in the monoclonal anti-human CREPT, Ki-67, PCNA or β-actin antibody (Abcam, Cambridge, MA, USA) at room temperature for 2 h. After washing with Tris-buffered saline, the membrane was incubated in anti-rabbit or antimouse IgG (Santa Cruz Biotechnology, USA) for 1 h at room temperature. The blots were visualized via enhanced chemiluminescence (ECL) according to the manufacturer's protocol. The experiments were independently repeated at least three times.

Statistical analysis

The statistical analyses were performed using Chi-square and Fisher's exact tests to determine the differences between groups. Overall survival (OS) was defined as the time of surgery until the time of death, and disease-free survival (DFS) was defined as the time of surgery until the appearance of evidence of radiological recurrence or metastasis. Kaplan-Meier analysis followed by the log rank test was performed to estimate the OS of each group. For the univariate and multivariate analyses, independent prognostic factors of patient survival were determined using Cox regression methods. The statistical analyses of the clinical samples were performed using SPSS 16.0 software (SPSS, Inc., Chicago, IL, USA). A two-tailed P-value <

Characteristic	Total	CREPTe, n expression (%) Positive, n = 56	Ρ	Ki-67e, n expression (%) Positive, n = 49	Ρ	PCNA expression n (%) Positive, n = 63	Ρ
Age, years							
< 60	63 (88.7)	49 (49.7)	1.000	43 (43.5)	1.000	50 (48.8)	0.368
≥60	8 (11.3)	7 (6.30)	1.000	6 (5.50)		5 (6.20)	
Sex							
Female	57 (80.3)	44 (45.0)	0.719	38 (39.3)	0.526	42 (44.2)	0.166
Male	14 (19.7)	12 (11.0)	0.719	11(9.70)		13 (10.8)	
Tumor size, cm							
≤ 10	28 (39.4)	20 (22.1)	0.146	18 (19.3)	0.608	21 (21.7)	0.781
> 10	43 (45.1)	36 (33.9)		31 (29.7)		34 (33.3)	
Tumor differentia	tion score						
1 score	7 (9.90)	4 (5.50)	0.299	3 (4.80)	0.085	1 (5.40)	<0.001
2 score	63 (88.7)	51 (49.7)		43 (43.5)		53 (8.8)	
3 score	1 (1.40)	1 (0.80)		1 (0.70)		1 (0.80)	
Mitotic count sco	ore						
1	37 (52.1)	30 (30.8)	0.109	22 (26.9)	0.050	28 (30.2)	0.856
2	16 (22.5)	15 (11.8)		14 (10.4)		13 (11.6)	
3	18 (25.4)	11 (13.4)		13 (11.7)		14 (13.2)	
Tumor necrosis s	core						
0	22 (31.0)	18 (20.5)	0.059	15 (17.9)	0.468	18 (20.1)	0.438
1	25 (35.2)	19 (16.6)		16 (14.5)		17 (16.3)	
2	24 (33.8)	19 (18.96)		18 (16.6)		20 (18.6)	
Histological grade	e						
1	10 (14.1)	7 (10.3)	0.044	6 (9.0)	0.064	5 (10.1)	0.009
2	48 (67.6)	40 (36.3)		33 (31.7)		40 (35.6)	
3	13 (18.3)	9 (9.50)		10 (8.3)		10 (9.38)	

Table 1. Correlations between Ki-67, CREPT and PCNA expression and clinicopathological characteristics	teris-
tics	

Statistical analyses were performed using the Pearson χ^2 test.

0.05 was considered to be statistically significant.

Results

Expression of CREPT, Ki-67, and PCNA in retroperitoneal LMS samples

Histopathological examination showed that the retroperitoneal LMS tumor tissues consisted of spindle cells that formed sheets or fascicles [10]. These cells contained oval nuclei and an elongated hyperchromatic and abundant eosinophilic cytoplasm (H&E, × 400, Figure 1A). To investigate the potential roles of CREPT, Ki-67 and PCNA in retroperitoneal LMS, we examined the expression of CREPT, Ki-67 and PCNA in paraffin-embedded sections from 71 patients via immunohistochemistry. We detected the expression of CREPT, Ki-67 and PCNA only in the nucleus (Figure 1B-D). A total of 56 patients (78.9%) displayed positive CREPT expression. Positive expression of Ki-67 was detected in 49 cases (69.0%), and positive PCNA expression was detected in 63 cases (88.7%). There was a significant association between the expression of CREPT and the mitotic count, the histological grade, Ki-67 expression and PCNA expression (P = 0.046, P = 0.044, P = 0.002 and P < 0.001, respectively, **Table 1**).

Univariate and multivariate analysis for prognostic factors

A total of 64 retroperitoneal LMS patients with available follow-up data were selected to generate the Kaplan-Meier survival curves. The OS rangeds from 5.0 to 74.0 (mean 34.0) months. The five-year OS rate of the retroperitoneal LMS patients was 28.1%. Univariate analysis revealed that the expression levels of CREPT, Ki-67 and PCNA were significant prognostic factors for OS (**Table 2**). The patients displaying positive CREPT expression (mean, 33 months) experienced a shorter OS than those lacking CREPT expression (mean, 60 months). The tumor size, histological grade, and expression

	Univariate analysis			Multivariate analysis		
	RR	P Value	95.0% CI for Exp (B)	RR	P Value	95.0% CI for Exp (B)
Age	1.130	0.780	0.479-2.669			
Sex	1.804	0.116	0.846-3.766			
Size	1.878	0.038	1.036-3.405			
Tumor differentiation	1.330	0.540	0.535-3.308			
Mitotic count	1.293	0.127	0.930-1.800			
Tumor necrosis	1.142	0.422	0.826-1.579			
Histological grade	1.719	0.024	1.074-2.751	1.858	0.011	1.153-2.995
Ki-67 expression	2.041	0.047	1.011-4.123			
CREPT expression	2.381	0.022	1.132-5.006	2.606	0.012	1.236-5.494
PCNA expression	2.442	0.042	1.033-5.773			

Table 2. Univariate and multivariate survival analyses of 71 patients with retroperitoneal LMS

of CREPT, Ki-67 and PCNA correlated with poor OS (**Figure 2**; **Table 2**). Furthermore, a multivariate analysis revealed that CREPT expression and the histological grade are independent prognostic factors of OS (**Table 2**; P = 0.012and P = 0.011, respectively). Taken together, these results suggested that CREPT is strongly correlated with the prognosis of retroperitoneal LMS patients.

Comparison of the expression of CREPT, Ki-67, and PCNA in retroperitoneal LMS tumors and adjacent noncancerous tissues via WB.

Five paired fresh frozen samples, which included retroperitoneal LMS tumor and adjacent noncancerous tissues, were collected for CREPT, Ki-67 and PCNA protein expression analysis (Figure 3). The results showed that CREPT was highly expressed in retroperitoneal LMS tumor tissues (Figure 3, top panel). In contrast, low levels of CREPT expression were detected in the adjacent normal tissues (Figure 3, top panel). Additionally, we found that the expression levels of Ki-67 and PCNA in the retroperitoneal LMS tumor tissue were significantly higher than those in the adjacent noncancerous tissues (Figure 3, middle panel). These results suggested that CREPT expression is correlated with the tumorigenesis of retroperitoneal LMS.

Correlation between CREPT expression and initial recurrence

A total of 18 patients survived for the median follow-up duration of 41 (range, 1-127) months. Based on univariate analysis, histological

grade, tumor size and CREPT expression were significant prognostic factors (Table 2). Clinical follow-up was performed, and the survival analysis suggested a close correlation between CREPT expression and initial recurrence of retroperitoneal LMS. The 5-year disease-specific survival (DSS) was 34% for primary LMS. Among the 45 patients experiencing recurrence, the pattern of recurrence was predominately distant recurrence (DR), which occurred in 19 patients (42.2%). Local recurrence (LR) occurred in only 16 patients (35.6%); 10 patients (22.2%) exhibited both LR and DR. As shown in Figure 4, CREPT expression was a prognostic factor for distant primary LMS recurrence (P = 0.04).

Discussion

Soft tissue sarcomas constitute 0.7% of adult malignancies [11]. Ten to twenty percent of soft tissue sarcomas occur in the retroperitoneum, and LMS, liposarcoma and fibrosarcoma are the most common histological types [12-16]. The retroperitoneum provides a broadly expandable anatomic location for tumor growth, and these tumors often attain a large size before symptoms manifest [1, 17]. Relevant symptoms are often indistinguishable from those of a mass in the abdominal viscera or reproductive organs. LMSs are relatively insensitive to radiotherapy and chemotherapy, and broad surgical excision is the preferred treatment. Retroperitoneal LMSs display relatively late presentation and are often proximal to several vital structures in the retroperitoneal space [18, 19]. Although the standard treatment

CREPT expression correlates to poor prognosis for leiomyosarcoma

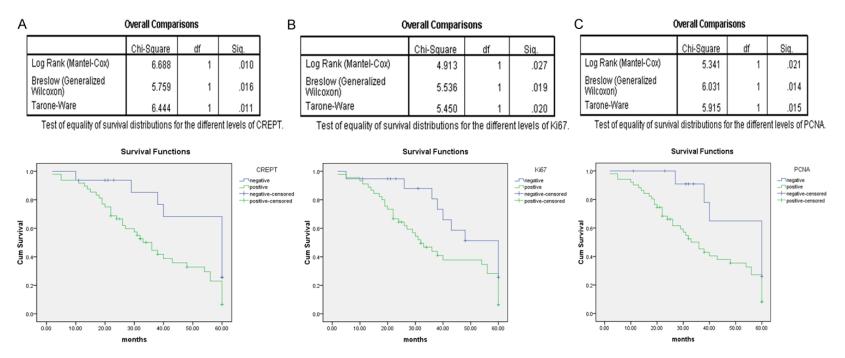


Figure 2. Kaplan-Meier analyses of the overall survival of 64 retroperitoneal LMS patients according to CREPT (A), Ki-67 (B) and PCNA (C) expression.

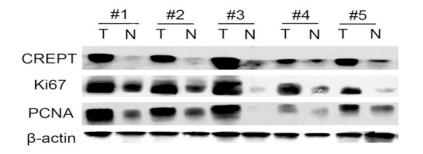


Figure 3. Western blot analysis of CREPT, Ki-67 and PCNA expression in retroperitoneal LMS tumor and normal adjacent tissues. T refers to the tumor tissue, and N refers to the paired normal tissue from the same patient.

remains margin-negative surgical resection, the 5-year survival rate after surgical resection remains low, ranging from 29% to 67% [20, 21]. However, the exact molecular mechanisms of the progression and recurrence of retroperitoneal LMSs are unclear, and there is a lack of valid and reliable biomarkers to predict retroperitoneal LMS recurrence.

Ki-67, a nuclear antigen present during the G1, S, G2, and M phases of all proliferating human cells, is a biomarker of tumor proliferation [22]. Patients displaying increased Ki-67 expression exhibit a lower cancer-specific survival rate [23]. The level of Ki-67 expression has been used in clinical analyses to predict the survival rate in a scenario with several neoplasms. Previous studies have identified Ki-67 as a tool in determining the malignancy of smooth muscle neoplasms (exemplified by myxoid LMS of the uterus) [24, 25]. PCNA is central to many essential cellular processes, such as DNA replication, DNA damage repair, chromatin structure maintenance and cell cycle progression [26]. PCNA has been widely used in studies assessing the growth rate of human malignancies. These previous studies indicated a significant expression level of PCNA in low- and intermediate-grade LMS [27].

CREPT has been reported to be a highly expressed oncogene in a variety of tumors [7]. CREPT expression accelerates malignant cell growth and tumorigenesis. Lu D *et al.* revealed that CREPT mRNA expression was up-regulated in various human malignant tumors and that the CREPT protein expression level was closely associated with the degree of differentiation and the clinical stage of the tumor [7].

In this report, we first detected the protein expression of CREPT in retroperitoneal LMS

tumor tissue. Immunohistochemical staining results showed that CREPT, Ki-67 and PCNA were positively expressed in more than half of the retroperitoneal LMS patients. Further investigation showed that the expression of CREPT correlated with the expression of Ki-67 and PCNA. Our findings were in line with previous results for sarcoma [27]. We observed CREPT protein expression in retroperitoneal LMS samples and paired normal

tissues. The expression of CREPT in the retroperitoneal LMS tumor tissue was significantly stronger than that in the adjacent noncancerous tissue. Therefore, we speculated that CREPT may play an important role in retroperitoneal LMS.

Our statistical analysis showed that the expression of CREPT, Ki-67 and PCNA closely correlated with 5-year overall survival and the distant recurrence of retroperitoneal LMS. Patients displaying high CREPT, Ki-67 or PCNA expression exhibited relatively short recurrence-free survival and overall survival; the patients displaying low CREPT expression exhibited relatively longer survival (P = 0.010, P= 0.027 and P = 0.021, respectively).

Univariate analyses identified tumor size, histological grade, and the expression level of CREPT, Ki-67 and PCNA as significant prognostic factors (P = 0.038, P = 0.024, P = 0.022, P = 0.047 and P = 0.042, respectively), and multivariate Cox regression analysis revealed that the histological grade and expression levels of CREPT were independent prognostic factors of poor overall survival (OS) among retroperitoneal LMS patients (P = 0.011 and P = 0.012, respectively). The retroperitoneal LMS patients in our study experienced a higher incidence of DR (42.2% at 5 years) than LR. With respect to DR, we found that CREPT expression is a prognostic factor for the distant recurrence of retroperitoneal LMS (*P* = 0.04, **Figure 4**). The above results strongly suggested that CREPT plays an important role in retroperitoneal LMS progression and that CREPT represents a valuable biomarker in predicting the prognosis of patients with retroperitoneal LMS.

CREPT has been suggested to display unique advantages for predicting the prognosis and

A Local recurrence

	Chi-Square	df	Siq.		
Log Rank (Mantel-Cox)	1.595	1	.207		
Breslow (Generalized Wilcoxon)	1.071	1	.301		
Tarone-Ware	1.417	1	.234		

Test of equality of survival distributions for the different levels of CREPT.

Overall Comparisons

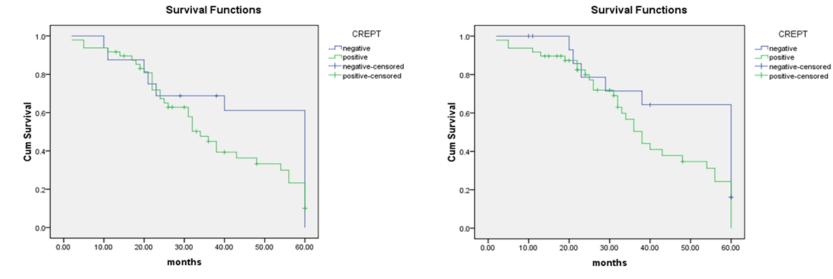


Figure 4. The expression of CREPT is an independent prognostic factor for distant recurrence-free survival (B) but not local recurrence-free survival of primary LMS (A). (P = 0.022 and P = 0.207, respectively).

B Di

Distant recurrence

Overall Comparisons

	Chi-Square	df	Siq.	
Log Rank (Mantel-Cox)	5.275	1	.022	
Breslow (Generalized Wilcoxon)	2.616	1	.106	
Tarone-Ware	3.793	1	.051	

Test of equality of survival distributions for the different levels of CREPT.

distant recurrence of retroperitoneal LMS compared with conventional biomarkers such as Ki-67 and PCNA. Therefore, CREPT serves as a candidate biomarker that could be used in combination with the conventional clinical biomarkers mentioned above. In clinical studies, accurate and reliable prognostic markers are crucial for providing comprehensive prognostic information and an accurate basis for treatment decisions. In this study, we identified the novel potential biomarker CREPT to be a prognostic factor in cancer progression and distant recurrence among patients with retroperitoneal LMS.

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Disclosure of conflict of interest

None.

Address correspondence to: Dr. Peiyu Li, Department of General Surgery, Chinese PLA General Hospital, Beijing 100853, China. E-mail: peiyuli301@ 163.com

References

- [1] Strauss DC, Hayes AJ and Thomas JM. Retroperitoneal tumours: review of management. Ann R Coll Surg Engl 2011; 93: 275-280.
- [2] Strauss DC, Hayes AJ, Thway K, Moskovic EC, Fisher C and Thomas JM. Surgical management of primary retroperitoneal sarcoma. Br J Surg 2010; 97: 698-706.
- [3] Jenkins MP, Alvaranga JC and Thomas JM. The management of retroperitoneal soft tissue sarcomas. Eur J Cancer 1996; 32A: 622-626.
- [4] Neville A and Herts BR. CT characteristics of primary retroperitoneal neoplasms. Crit Rev Comput Tomogr 2004; 45: 247-270.
- [5] Karakousis CP and Perez RP. Soft tissue sarcomas in adults. CA Cancer J Clin 1994; 44: 200-210.
- [6] Alvarenga JC, Ball AB, Fisher C, Fryatt I, Jones L and Thomas JM. Limitations of surgery in the

treatment of retroperitoneal sarcoma. Br J Surg 1991; 78: 912-916.

- [7] Lu D, Wu Y, Wang Y, Ren F, Wang D, Su F, Zhang Y, Yang X, Jin G, Hao X, He D, Zhai Y, Irwin DM, Hu J, Sung JJ, Yu J, Jia B and Chang Z. CREPT accelerates tumorigenesis by regulating the transcription of cell-cycle-related genes. Cancer Cell 2012; 21: 92-104.
- [8] Pautier P, Floquet A, Penel N, Piperno-Neumann S, Isambert N, Rey A, Bompas E, Cioffi A, Delcambre C, Cupissol D, Collin F, Blay JY, Jimenez M and Duffaud F. Randomized multicenter and stratified phase II study of gemcitabine alone versus gemcitabine and docetaxel in patients with metastatic or relapsed leiomyosarcomas: a Federation Nationale des Centres de Lutte Contre le Cancer (FNCLCC) French Sarcoma Group Study (TAXO-GEM study). Oncologist 2012; 17: 1213-1220.
- [9] Esposito V, Baldi A, De Luca A, Tonini G, Vincenzi B, Santini D, Persichetti P, Mancini A, Citro G and Baldi F. Cell cycle related proteins as prognostic parameters in radically resected non-small cell lung cancer. J Clin Pathol 2005; 58: 734-739.
- [10] Weiss SW, Goldblum JR and Enzinger FM. Enzinger and Weiss's Soft Tissue Tumors. St. Louis: Mosby; 2001.
- [11] Todd CS, Michael H and Sutton G. Retroperitoneal leiomyosarcoma: eight cases and a literature review. Gynecol Oncol 1995; 59: 333-337.
- [12] Adam YG, Oland J, Halevy A and Reif R. Primary retroperitoneal soft-tissue sarcomas. J Surg Oncol 1984; 25: 8-11.
- [13] Cody HS 3rd, Turnbull AD, Fortner JG and Hajdu SI. The continuing challenge of retroperitoneal sarcomas. Cancer 1981; 47: 2147-2152.
- [14] McGrath PC, Neifeld JP, Lawrence W Jr, DeMay RM, Kay S, Horsley JS 3rd and Parker GA. Improved survival following complete excision of retroperitoneal sarcomas. Ann Surg 1984; 200: 200-204.
- [15] Catton CN, O'Sullivan B, Kotwall C, Cummings B, Hao Y and Fornasier V. Outcome and prognosis in retroperitoneal soft tissue sarcoma. Int J Radiat Oncol Biol Phys 1994; 29: 1005-1010.
- [16] Hassan I, Park SZ, Donohue JH, Nagorney DM, Kay PA, Nasciemento AG, Schleck CD and Ilstrup DM. Operative management of primary retroperitoneal sarcomas: a reappraisal of an institutional experience. Ann Surg 2004; 239: 244-250.
- [17] Ludwig JA and Weinstein JN. Biomarkers in cancer staging, prognosis and treatment selection. Nat Rev Cancer 2005; 5: 845-856.
- [18] Wang LD, Hong JY, Qiu SL, Gao H and Yang CS. Accumulation of p53 protein in human esophageal precancerous lesions: a possible early biomarker for carcinogenesis. Cancer Res 1993; 53: 1783-1787.

- [19] Liles JS, Tzeng CW, Short JJ, Kulesza P and Heslin MJ. Retroperitoneal and intra-abdominal sarcoma. Curr Probl Surg 2009; 46: 445-503.
- [20] Clary BM, DeMatteo RP, Lewis JJ, Leung D and Brennan MF. Gastrointestinal stromal tumors and leiomyosarcoma of the abdomen and retroperitoneum: a clinical comparison. Ann Surg Oncol 2001; 8: 290-299.
- [21] Gladdy RA, Qin LX, Moraco N, Agaram NP, Brennan MF and Singer S. Predictors of survival and recurrence in primary leiomyosarcoma. Ann Surg Oncol 2013; 20: 1851-1857.
- [22] Gerdes J, Lemke H, Baisch H, Wacker HH, Schwab U and Stein H. Cell cycle analysis of a cell proliferation-associated human nuclear antigen defined by the monoclonal antibody Ki-67. J Immunol 1984; 133: 1710-1715.
- [23] Rioux-Leclercq N, Turlin B, Bansard JY, Patard JJ, Manunta A, Moulinoux JP, Guillé F, Ramée MP and Lobel B. Value of immunohistochemical Ki-67 and p53 determinations as predictive factors of outcome in renal cell carcinoma. Urology 2000; 55: 501-505.

- [24] Mittal K and Demopoulos RI. MIB-1 (Ki-67), p53, estrogen receptor, and progesterone receptor expression in uterine smooth muscle tumors. Hum Pathol 2001; 32: 984-987.
- [25] Sprogoe-Jakobsen S and Holund B. Immunohistochemistry (Ki-67 and p53) as a tool in determining malignancy in smooth muscle neoplasms (exemplified by a myxoid leiomyosarcoma of the uterus). APMIS 1996; 104: 705-708.
- [26] Moldovan GL, Pfander B and Jentsch S. PCNA, the maestro of the replication fork. Cell 2007; 129: 665-679.
- [27] Steck K and el-Naggar AK. Comparative flow cytometric analysis of Ki-67 and proliferating cell nuclear antigen (PCNA) in solid neoplasms. Cytometry 1994; 17: 258-265.