Case Report A young man with primary prostatic extra-gastrointestinal stromal tumor: a rare case report and review of the literature

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Received February 8, 2014; Accepted March 10, 2014; Epub March 15, 2014; Published April 1, 2014

Abstract: Mounting evidence demonstrates the presence of extragastrointestinal stromal tumor (EGIST) which originates from tissues outside the gastrointestinal (GI) tract and shares overlapping immunohistological features with gastrointestinal stromal tumor (GIST). GIST emanating from prostate is extremely rare. To our knowledge, there are only 3 definitely reported cases of primary prostatic EGIST. Herein, we report a case of prostatic EGIST in 31-yearold man with low urinary tract symptoms who was initially misdiagnosed as sarcoma of prostate. Imaging studies assist in determining the origin and location of EGIST. Immunohistochemical assessment (DOG-1, CD117, and CD34) helps in differentiating such lesion from other stromal tumors and in addressing an appropriate and optimal therapeutic strategy.

Keywords: Extragastrointestinal stromal tumor (EGIST), prostate, differential diagnosis, imatinib

Introduction

Extragastrointestinal stromal tumor (EGIST) is defined as mesenchymal neoplasms arsing from soft tissues outside the gastrointestinal (GI) tract, which is morphologically, histologically, and immunophenotypically similar to its gastrointestinal counterpart (i.e. gastrointestinal stromal tumor, GIST) [1]. The diagnosis of EGIST relies on the combination of tumor location, histopathologic appearance, and immunohistochemical analysis. Immunohistochemistry study plays a major role in GIST diagnosis. Most of these tumors express KIT (CD117) tyrosine kinase and show the presence of activating mutations in KIT or PDGFRa. Recently, a novel antibody, DOG1, has been identified to be sensitive and specific, in particular, for KITmutation-negative ones. Imatinib mesylate with activity against KIT and PDGFRA is the primary therapeutic candidate. To our best knowledge, there are rare definitive reports on EGIST arising from prostate [2-4]. In this report, we present our unique case and discuss the clinical presentation, differential diagnosis, pathologic characteristics, and therapeutic strategies for primary prostatic EGIST.

Case report

A 31-year-old man was admitted to our hospital with dysuria. He had frequency, urgency for 4 months and intermittent gross hematuria for 2 weeks. Digital rectal examination revealed an enlarged prostate with hard consistency but without bump, tenderness, or bleeding. The surface of prostate was not smooth. Serum level of prostate specific antigen (PSA) was 0.37 ng/ml. Pelvic B-mode ultrasound examination showed an expanded prostate (6.0×6.1) × 6.5 cm) with irregular internal structure and dark area of fluid. Ultrasonic inspection displayed a 1.4 cm protrusion with moderate echo at vesical neck, which showed no clear boundary with the prostate. Pelvic magnetic resonance imaging (MRI) examination (Figure 1) demonstrated 1) abnormal shape of prostate and seminal vesicles with mixed signal (generally, moderate signal was mainly detected), 2) an enlarged prostate with expansive growth and compression of the rectum and bladder. The whole-body bone scan revealed no bone involvement. Besides, no tumor dwelling inside or outside of the rectal wall was detected by enteroscopy and transrectal ultrasound (TRUS)

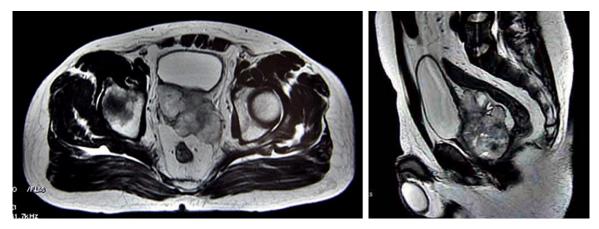


Figure 1. Magnetic resonance imaging showing 1) abnormal morphological appearance of prostate and seminal vesicles with mixed signal, 2) an enlarged prostate with expansive growth and compression of the rectum and bladder.

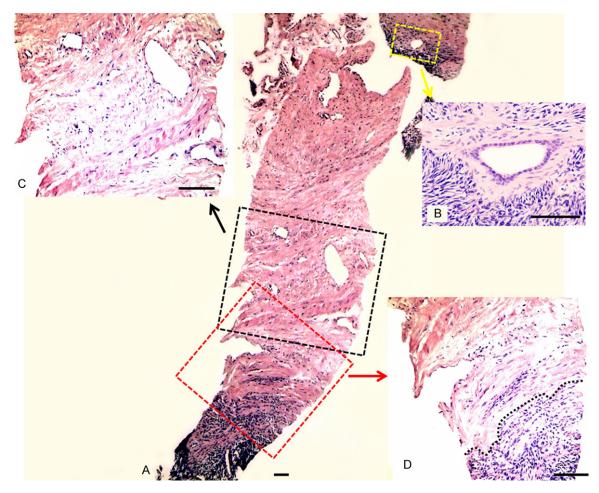


Figure 2. Histopathology of the tumor. H & E staining of tissue section from biopsy showed irregular dense arrangement of tumor cells (A) and the cytological pleomorphism of tumor tissue, composed of spindle-shape (B) and epithelioid cells (C). A boundary (dashed line) between regions of epithelioid or mixed epithelioid/spindle cell was observed (D). (magnification: A, \times 100; B, \times 400; C and D, \times 200; Scale bars = 100 µm).

examination. According to above indications, the initial diagnosis was sarcoma of the pros-

tate. TRUS guided prostate biopsy was performed for pathologic diagnosis. Histologically,

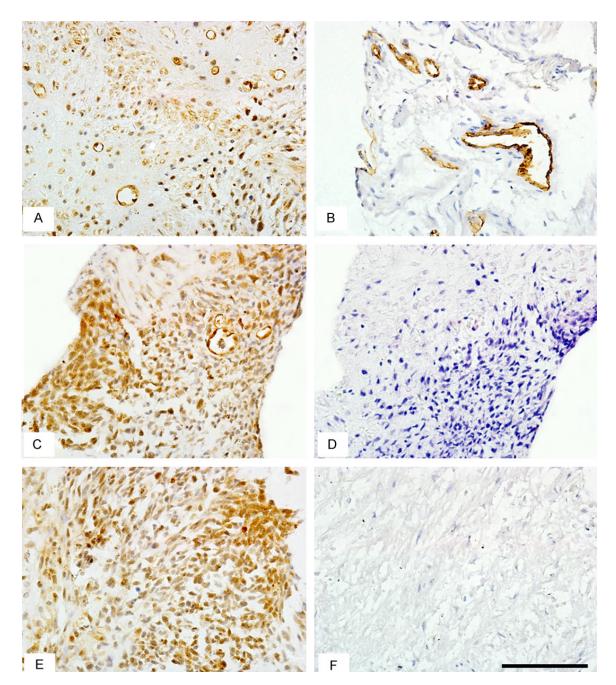


Figure 3. Immunohistochemical staining of the tumor cells. Immunostaining of the lesion displayed strong reactivity for CD34 (A) and diffuse positivity for CD117 (C) and DOG-1 (E). (B) is a positive control for CD34 staining illustrating the vessels, while (D) and (F) are blank controls for CD117 and DOG-1, respectively. (magnification: \times 400; Scale bars = 100 µm).

the tumor consisted of spindle-shape, oval, and epithelioid cells (**Figure 2**). Tumor cells were very big in size, with abundant eosinophilic cytoplasm and spherical/oval nuclei. The nuclei had a varying size, and some of them stained light. The mitotic count was more than 10 per 50 high-power fields (HPF). Tumor hemorrhage or necrosis were frequently seen. Tumor cells showed strong and diffuse immunoreactivity for DOG-1, CD117, and CD34 (**Figure 3**) and were negative for S-100 and smooth muscle actin (SMA) (not shown). The pathologic diagnosis was GIST of the prostate based on histological and immunostaining results. Considering extension of the mass and increased risk of rectal injury, no surgical treatment was per-

Ref.	Age (yrs)	Tumor Size (cm)	Clinical presentation	Immunoreactivity	Treatment	Follow-up Interval (Months)	Outcomes	Metastasis
[2]	49	8	Perineal pain	CD117, CD34, Desmin	RP	14	No recurrence	None
[3]	75	6.7	Dysturia, frequency, hesitancy	CD117, CD34, Desmin	TURP + RP	6	Good condition	None
[4]	49	14.2	Acute urinary retention, body weight loss	CD117, α-SMA	Imatinib	24	Reduced mass volume and liver nodules	Liver (when diagnosed)

Table 1. Review of the Literature on Primary Prostatic EGIST

Abbreviations: TURP, transurethral prostatectomy; RP, radical prostatectomy.

formed. Instead, the patient was administered imatinib (400 mg per day). He took imatinib intermittently due to financial reasons, resulting in a poor response during a 3-month drug treatment. As the mass volume increased apparently ($6.5 \times 7.2 \times 9.0$ cm), leading to urinary retention, he then received indwelling catheter for a 3-month follow-up. The patient ultimately developed intestinal obstruction and died for electrolyte disturbances and multiple organ failure. Due to no informed consented obtained, autopsy was not performed.

Discussion

GISTs may arise anywhere in the gastrointestinal tract, from the esophagus to the rectum. Recent studies have disclosed identical lesions (EGIST) occurring in various locations outside the alimentary tract. As a very rare tumor, EGIST constitutes only 5%-10% of GIST [5]. The mean patient age at diagnosis is 58 years (range: from 31 to 82). The majority of reports on EGIST that is histologically similar to GIST are derived from the mesentery, omentum, and retroperitoneum [6, 7]. However, a significant number of individual cases have also been demonstrated in other sites, encompassing pancreas [8, 9], female genital organs [10, 11], urinary bladder [12, 13] and seminal vesicles [14]. To our knowledge, only a few cases of EGIST presenting as prostatic mass have been reported previously, among which primary prostatic EGIST is less commonly [2-4].

Due to heterogeneous clinical presentation and unusual anatomic locations of prostatic EGIST, it is difficult to differentiate such tumor from other prostatic occupying lesions and to identify the origin of the mass. A variety of symptoms have been demonstrated in the previous reports (**Table 1**). As for our case, occasional hematuria was presented as well apart from low urinary tract symptoms, suggesting diverse manifestation of such disease as a pitfall for differential diagnosis. Although TRUS guided prostate biopsy is widely performed for the purpose of establishing pathologic diagnosis, it plays poorly in determining the location of tumors. Accordingly, prostatic mass diagnosed as EGIST by biopsy may originate from either prostate or rectum. In this context, other approaches including assistant imaging examinations as well as enteroscopy are required to help identifying between such two origins.

Apart from the reported cases of prostatic EGIST, it has been recently shown that some cases of GIST arising from the rectum present clinically as prostatic masses [15]. Herawi and coworkers reported 4 resected tumors (in an 8-case study) following initial diagnosis by needle biopsy [16]. Based on their data, two cases originated from rectum (one is shown to be primary in the rectum without prostatic involvement, the other one extensively involving the prostate), while the origins of the other two were not well defined (one was separated from the prostate, the other one a perirectal mass). More recently, a literature review has been carried by Anagnostou and his group to reveal 20 cases of EGIST occurring in the prostate, diagnosed as either primary EGIST or rectal neoplasms extending to this organ [17]. They recapitulated the previous reports diagnosed as primary prostatic GIST and doubted the existence of such neoplasm. They put a premium on the exclusion of rectal involvement before such diagnosis is made. In our case, the prostatic origin was addressed mainly based on imaging results. What is note, there is no evidence supporting the existence of other GIST inside or outside the GI. Thus, we consider this prostate lesion to be a primary prostatic GIST. Theoretically, as suggested by Loeb, an ideal diagnosis can be drawn when the prostatic tumor is surgically resected with intact capsule, revealing no connection to the rectal wall [18].

Histopathology represents the gold standard in GIST diagnosis and a number of antibodies have been used in the routine practice. CD34

may aid, but it is positive in many other types of soft tumors [19]. Most GIST stain positively for CD117 (C-kit) and harbor a kinase-activating mutation in either KIT or PDGFR α [20, 21]. Because the distribution and strong expression of CD117 is quite limited in sarcoma other than GIST [22], CD117 is a relatively specific marker for GIST. However, approximately ~4% to 15% of GIST fail to stain for CD117 [23, 24]. A large percentage of these tumors possess PDGFRa mutations and respond to tyrosine kinase inhibitors [25, 26]. In this regard, the traditional panel of immunohistochemical stains is not sufficient to address accurate diagnosis of GIST. Recently, DOG-1 has been shown to be a promising antibody used in GIST diagnosis [27, 28]. As the most specific and sensitive marker of GIST [29], DOG-1 can reliably stain cases that are CD117-weak/negative [30]. Moreover, the combination of positivity for both antibodies (CD117 and DOG-1) were demonstrated to be reassuring in histological diagnosis [31]. Mutation screening of KIT or PDGFRα assists in confirming the diagnosis [3] and predicting the likelihood of imatinib response [18], but this technique adds to the time and cost of diagnosis. Screening is required in tumors that are negative for both CD117 and DOG-1 [30].

Due to emerging adjuvant treatment strategies, the need for precise risk stratification and classification is increasingly emphasized to attain an optimal individualized therapy. A decade ago, Flecher et al. suggested that some effective prognostic parameters (e.g. tumor size and the mitotic rate) should be taken into significant consideration in risk assessment and that the definitions for the risk categories (as very low, low, intermediate, or high) should be proved to be clinically useful [32]. According to Flecher's study with combination to a recent work [33] assessing the reliability of the staging system for GIST in the new revision of the AJCC, our case had a high risk of aggressive behavior. Similar to that of GIST, the most effective treatment for EGIST is aggressive surgical intervention associated with the use of imatinib [34]. In patients with localized tumors (whose lesions are very small, even < 2 cm, and lesions with very low mitotic rates, even < 5 per 50 HPF), surgery remains the elective treatment, due to the insensitivity of tumor cells to radiotherapy or chemotherapy. They are considered low risk and have small chance to reoccur with surgery and may not require adjuvant targeted therapy.

For advanced tumors, the role of surgery is very limited and the patients may benefit from imatinib therapy. Although imatinib therapy represents standard treatment, showing continuous improvements in progressive-free and overall survival, the therapeutic efficacy in metastatic cases remains disputed. Surgical intervention was not considered in current patient because the lesion compressed and extended toward bladder and rectum. Instead, we preferred imatinib to surgical treatment in terms of his dismal prognosis. However, the young patient took imatinib only intermittently due to financial reasons, leading to a poor response to the targeted therapy and rapid exacerbation.

In conclusion, we reported an extremely rare case of primary GIST arising from the prostate. The diagnosis depended on imaging studies, pathologic results as well as immunohistochemical findings. Imaging examinations combined with enteroscopy contribute to identifying tumor origin. A broader immunohistochemical panel including DOG-1, CD117, as well as CD34 allows the confirmatory diagnosis of GIST. Because of its rarity, clinicians involved in the assessment of a mass arising in the prostate are suggested to be alert of EGIST in differential diagnosis, even in a young man.

Acknowledgements

This work is funded by grants from the National Natural Science Foundation of China (81172451) and Key Projects of Tianjin Science and Technology Support Programme (112CGYSY02300).

Disclosure of conflict of interest

All authors have no conflicts of interest.

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References

[1] Miettinen M and Lasota J. Gastrointestinal stromal tumors–definition, clinical, histological, immunohistochemical, and molecular genetic features and differential diagnosis. Virchows Arch 2001; 438: 1-12.

- [2] Yinghao S, Bo Y and Xiaofeng G. Extragastrointestinal stromal tumor possibly originating from the prostate. Int J Urol 2007; 14: 869-871.
- [3] Lee CH, Lin YH, Lin HY, Lee CM and Chu JS. Gastrointestinal stromal tumor of the prostate: a case report and literature review. Hum Pathol 2006; 37: 1361-1365.
- [4] Van Der Aa F, Sciot R, Blyweert W, Ost D, Van Poppel H, Van Oosterom A, Debiec-Rychter M and De Ridder D. Gastrointestinal stromal tumor of the prostate. Urology 2005; 65: 388.
- [5] Emory TS, Sobin LH, Lukes L, Lee DH and O'Leary TJ. Prognosis of gastrointestinal smooth-muscle (stromal) tumors: dependence on anatomic site. Am J Surg Pathol 1999; 23: 82-87.
- [6] Mouaqit O, Jahid A, Ifrine L, Omar El Malki H, Mohsine R, Mahassini N and Belkouchi A. Primary omental gastrointestinal stromal tumors. Clin Res Hepatol Gastroenterol 2011; 35: 590-593.
- [7] Miettinen M, Sobin LH and Lasota J. Gastrointestinal stromal tumors presenting as omental masses—a clinicopathologic analysis of 95 cases. Am J Surg Pathol 2009; 33: 1267-1275.
- [8] Kim HH, Koh YS, Park EK, Seoung JS, Hur YH, Kim JC, Cho CK and Kim HJ. Primary extragastrointestinal stromal tumor arising in the pancreas: report of a case. Surg Today 2012; 42: 386-390.
- [9] Cecka F, Jon B, Ferko A, Subrt Z, Nikolov DH and Tycova V. Long-term survival of a patient after resection of a gastrointestinal stromal tumor arising from the pancreas. Hepatobiliary Pancreat Dis Int 2011; 10: 330-332.
- [10] Vazquez J, Perez-Pena M, Gonzalez B and Sanchez A. Gastrointestinal stromal tumor arising in the rectovaginal septum. J Low Genit Tract Dis 2012; 16: 158-161.
- [11] Lam MM, Corless CL, Goldblum JR, Heinrich MC, Downs-Kelly E and Rubin BP. Extragastrointestinal stromal tumors presenting as vulvovaginal/rectovaginal septal masses: a diagnostic pitfall. Int J Gynecol Pathol 2006; 25: 288-292.
- [12] Krokowski M, Jocham D, Choi H, Feller AC and Horny HP. Malignant extragastrointestinal stromal tumor of bladder. J Urol 2003; 169: 1790-1791.
- [13] Lasota J, Carlson JA and Miettinen M. Spindle cell tumor of urinary bladder serosa with phenotypic and genotypic features of gastrointestinal stromal tumor. Arch Pathol Lab Med 2000; 124: 894-897.
- [14] Song W, Yang JR, Wang YH and Liang QC. Primary extragastrointestinal stromal tumor of the seminal vesicles. Urology 2012; 79: e36-37.

- [15] Madden JF, Burchette JL, Raj GV, Daly JT and Tannenbaum M. Anterior rectal wall gastrointestinal stromal tumor presenting clinically as prostatic mass. Urol Oncol 2005; 23: 268-272.
- [16] Herawi M, Montgomery EA and Epstein JI. Gastrointestinal stromal tumors (GISTs) on prostate needle biopsy: A clinicopathologic study of 8 cases. Am J Surg Pathol 2006; 30: 1389-1395.
- [17] Anagnostou E, Miliaras D and Panagiotakopoulos V. Diagnosis of gastrointestinal stromal tumor (GIST) on transurethral resection of the prostate: a case report and review of the literature. Int J Surg Pathol 2011; 19: 632-636.
- [18] Loeb S, Lotan TL, Thornton K, Gearhart SL and Schoenberg MP. A case of gastrointestinal stromal tumor diagnosed on prostate biopsy. Nat Clin Pract Urol 2009; 6: 54-57.
- [19] Parfitt JR, Rodriguez-Justo M, Feakins R and Novelli MR. Gastrointestinal Kaposi's sarcoma: CD117 expression and the potential for misdiagnosis as gastrointestinal stromal tumour. Histopathology 2008; 52: 816-823.
- [20] Hirota S, Isozaki K, Moriyama Y, Hashimoto K, Nishida T, Ishiguro S, Kawano K, Hanada M, Kurata A, Takeda M, Muhammad Tunio G, Matsuzawa Y, Kanakura Y, Shinomura Y and Kitamura Y. Gain-of-function mutations of c-kit in human gastrointestinal stromal tumors. Science 1998; 279: 577-580.
- [21] Heinrich MC, Corless CL, Duensing A, Mc-Greevey L, Chen CJ, Joseph N, Singer S, Griffith DJ, Haley A, Town A, Demetri GD, Fletcher CD and Fletcher JA. PDGFRA activating mutations in gastrointestinal stromal tumors. Science 2003; 299: 708-710.
- [22] Sato O, Wada T, Kawai A, Yamaguchi U, Makimoto A, Kokai Y, Yamashita T, Chuman H, Beppu Y, Tani Y and Hasegawa T. Expression of epidermal growth factor receptor, ERBB2 and KIT in adult soft tissue sarcomas: a clinicopathologic study of 281 cases. Cancer 2005; 103: 1881-1890.
- [23] Debiec-Rychter M, Wasag B, Stul M, De Wever I, Van Oosterom A, Hagemeijer A and Sciot R. Gastrointestinal stromal tumours (GISTs) negative for KIT (CD117 antigen) immunoreactivity. J Pathol 2004; 202: 430-438.
- [24] Medeiros F, Corless CL, Duensing A, Hornick JL, Oliveira AM, Heinrich MC, Fletcher JA and Fletcher CD. KIT-negative gastrointestinal stromal tumors: proof of concept and therapeutic implications. Am J Surg Pathol 2004; 28: 889-894.
- [25] Corless CL, Schroeder A, Griffith D, Town A, Mc-Greevey L, Harrell P, Shiraga S, Bainbridge T, Morich J and Heinrich MC. PDGFRA mutations in gastrointestinal stromal tumors: frequency, spectrum and in vitro sensitivity to imatinib. J Clin Oncol 2005; 23: 5357-5364.

- [26] Heinrich MC, Corless CL, Demetri GD, Blanke CD, Von Mehren M, Joensuu H, McGreevey LS, Chen CJ, Van Den Abbeele AD, Druker BJ, Kiese B, Eisenberg B, Roberts PJ, Singer S, Fletcher CD, Silberman S, Dimitrijevic S and Fletcher JA. Kinase mutations and imatinib response in patients with metastatic gastrointestinal stromal tumor. J Clin Oncol 2003; 21: 4342-4349.
- [27] West RB, Corless CL, Chen X, Rubin BP, Subramanian S, Montgomery K, Zhu S, Ball CA, Nielsen TO, Patel R, Goldblum JR, Brown PO, Heinrich MC and Van De Rijn M. The novel marker, DOG1, is expressed ubiquitously in gastrointestinal stromal tumors irrespective of KIT or PDGFRA mutation status. Am J Pathol 2004; 165: 107-113.
- [28] Miettinen M, Wang ZF and Lasota J. DOG1 antibody in the differential diagnosis of gastrointestinal stromal tumors: a study of 1840 cases. Am J Surg Pathol 2009; 33: 1401-1408.
- [29] Liegl B, Hornick JL, Corless CL and Fletcher CD. Monoclonal antibody DOG1.1 shows higher sensitivity than KIT in the diagnosis of gastrointestinal stromal tumors, including unusual subtypes. Am J Surg Pathol 2009; 33: 437-446.
- [30] Espinosa I, Lee CH, Kim MK, Rouse BT, Subramanian S, Montgomery K, Varma S, Corless CL, Heinrich MC, Smith KS, Wang Z, Rubin B, Nielsen TO, Seitz RS, Ross DT, West RB, Cleary ML and Van De Rijn M. A novel monoclonal antibody against DOG1 is a sensitive and specific marker for gastrointestinal stromal tumors. Am J Surg Pathol 2008; 32: 210-218.

- [31] Novelli M, Rossi S, Rodriguez-Justo M, Taniere P, Seddon B, Toffolatti L, Sartor C, Hogendoorn PC, Sciot R, Van Glabbeke M, Verweij J, Blay JY, Hohenberger P, Flanagan A and Dei Tos AP. DOG1 and CD117 are the antibodies of choice in the diagnosis of gastrointestinal stromal tumours. Histopathology 2010; 57: 259-270.
- [32] Fletcher CD, Berman JJ, Corless C, Gorstein F, Lasota J, Longley BJ, Miettinen M, O'Leary TJ, Remotti H, Rubin BP, Shmookler B, Sobin LH and Weiss SW. Diagnosis of gastrointestinal stromal tumors: a consensus approach. Int J Surg Pathol 2002; 10: 81-89.
- [33] Rutkowski P, Wozniak A, Debiec-Rychter M, Kakol M, Dziewirski W, Zdzienicki M, Ptaszynski K, Jurkowska M, Limon J and Siedlecki JA. Clinical utility of the new American Joint Committee on Cancer staging system for gastrointestinal stromal tumors: current overall survival after primary tumor resection. Cancer 2011; 117: 4916-4924.
- [34] Barros A, Linhares E, Valadao M, Goncalves R, Vilhena B, Gil C and Ramos C. Extragastrointestinal stromal tumors (EGIST): a series of case reports. Hepatogastroenterology 2011; 58: 865-868.