Original Article Association of GSTs polymorphisms with risk of gestational diabetes mellitus

Yan Li¹, Shaoru Li², Qianqian Zhai¹, Jie Hai¹, Di Wang¹, Meng Cao¹, Qinggui Zhang¹

Departments of ¹Endocrinology, ²Gynaecology and Obstetrics, The First Affiliated Hospital of Xinxiang Medical University, Weihui 453100, Henan, China

Received September 22, 2015; Accepted October 25, 2015; Epub November 1, 2015; Published November 15, 2015

Abstract: We conducted a case-control study to investigate the association between *GSTM1*, *GSTT1* and *GSTP1* Ile105Val polymorphisms and development of gestational diabetes mellitus in a Chinese population. A total of 320 patients with gestational diabetes mellitus and 358 pregnancy subjects were consecutively collected between January 2013 and December 2014. Genotyping for detection of *GSTM1*, *GSTT1* and *GSTP1* Ile105Val was conducted by using PCR-RFLP (polymerase chain reaction-restriction fragment length polymorphisms) method. By Fisher's exact test, we found that the genotype distributions of *GSTP1* Ile105Val were in line with the Hardy-Weinberg equilibrium in control subjects (P=0.57). By Chi-square test, we found significant differences in the genotype distributions of *GSTM1* (χ^2 =11.49, P=0.001) and *GSTT1* (χ^2 =18.50, P<0.001). Using unconditional logistic analysis, individuals carrying the null genotypes of *GSTM1* and *GSTT1* were associated with an increased risk of gestational diabetes mellitus when compared with the present genotype, and the adjusted Ors (95% CI) were 1.71 (1.24-2.36) and 2.00 (1.44-2.79), respectively. However, the *GSTP1* Ile105Val polymorphism was not associated with an elevated risk of gestational diabetes mellitus. In conclusion, we suggest that the *GSTM1* null genotype and *GSTT1* null genotype are correlated with an increased risk of gestational diabetes mellitus in a Chinese population.

Keywords: GSTM1, GSTT1, GSTP1 IIe105Val, polymorphism, gestational diabetes mellitus

Introduction

Gestational diabetes mellitus is one type of diseases which only occurs in pregnant women [1], which is defined as carbohydrate intolerance during pregnancy. It is estimated that the incidence of gestational diabetes mellitus is between 2.4% and 22.3% among pregnant women worldwide [2, 3]. The development of gestational diabetes mellitus is resulted from complex multifactorial environmental and lifestyle factors, such as pregnancies at older age, prepregnancy high weight and BMI, family history of diabetes mellitus, dietary habit, physical inactivity and a history of gestational diabetes [4-9]. However, not all the pregnancies would develop gestational diabetes mellitus even when they expose to the same risk factors of gestational diabetes mellitus, which suggests that the hereditary factors may contribute to the development of gestational diabetes mellitus. Currently, many studies have reported that the genetic factors play an important role in the development of gestational diabetes mellitus, such as KCNQ1, MTHFR, FVL, G22A, Calpain 10 and TCF7L2 [10-14].

Glutathione-S-transferases (GSTs) belong to the family of cytosolic enzymes, which play an important role in the cellular defense system [15]. The main function of GSTs is to detoxify environmental toxicants and reactive oxygen species mediated cell injury in body, catalyze the harmful compounds in the combined with glutathione, and finally prevent the DNA damage in body [15, 16]. Three functional genes have been found in GSTs, including GSTM1, GSTT1 and GSTP1 IIe105Val. The polymorphisms of GSTM1 and GSTT1 could reduce the enzyme activities and binding activity of the two genes [17]. Only one previous study has investigated the association between the genetic polymorphisms and development of gestational diabetes mellitus [18]. Therefore, we conducted a case-control study to investigate the association between GSTM1, GSTT1 and GSTP1 Ile105Val polymorphisms and development of gestational diabetes mellitus in a Chinese population.

Patients and methods

Patients

Between January 2013 and December 2014, a total of 320 patients with gestational diabetes mellitus were consecutively collected from our hospital. All patients with gestational diabetes mellitus were between 25 and 40 years old. The diagnosis of gestational diabetes mellitus was based on the criteria from American Diabetes Association [19], and the gestational age is between 24 and 28th weeks of gestation with a 75 g, 2 h OGTT after overnight fast. The inclusion criteria for the gestational diabetes mellitus were patients without using oral hypoglycemic agents. The exclusion criteria were patients with pre-existing diabetes and hepatic and kidney diseases as well as endocrine disorders.

A total of 358 pregnancy subjects without gestational diabetes mellitus were randomly selected from individuals who received the regular prenatal in the Department of Obstetrics and Gynecology during the same period time. Controls that had a history of hypertension, diabetes and hepatic and kidney diseases as well as endocrine disorders were excluded from our study.

The demographic and clinical information of patients with gestational diabetes mellitus and control subjects were collected from a self-designed questionnaire or medical records. The demographic information included sex, age, Body Mass Index and a history of gestational diabetes mellitus. The clinical information included systolic blood pressure, diastolic blood pressure, diabetes mellitus, total cholesterol (TC), triglyceride (TG), low density lipopolysaccharide cholesterol (LDL-c) and high density lipopolysaccharide (HDL-c). The signed written informed consents were collected from all patients with gestational diabetes mellitus and control subjects.

DNA extraction and genotyping

Five ml of fasting venous blood were drawn from all patients and control subjects after participating into this study. The blood samples were stored in tubes with ethylene diamine tetraacetic acid (EDTA), and then the blood was centrifuged to separate the plasma content. The genomic DNA was extracted from the EDTA treated whole blood using the TIANamp Blood DNA Kit (Tiangen, Beijing, China). Genotyping for detection of GSTM1, GSTT1 and GSTP1 Ile105Val was conducted by using PCR-RFLP (polymerase chain reaction-restriction fragment length polymorphisms) method. Genotyping for detection of GSTM1, GSTT1 and GSTP1 IIe105Val was amplified by the following forward and reverse primers. For GSTM1, the primers were 5'-CTGCCCTACTTGATTGATGGG-3' (forwards) and 5'-CTGGATTGTAGCAGATCATGC-3' (reverse); For GSTT1, the primers were 5'-TTCCTTACTGGTCCTCACATCTC-3' (forwards) and 5'-TCACCGGATCATGGCCAGCA-3' (reverse): For GSTP1, the primers were the primers were 5'-GAA GAG CCA AGG ACA GGT AC-3' (forwards) and 5'-CAA CTT CAT CCA CGT TCA CC-3' (reverse). The fragment lengths for GSTM1, GSTT1 and GSTP1 IIe105Val were 215 bp, 480 bp and 176 bp. The PCR conditions were as follows: an initial denaturation at 95°C for 5 min, 35 cycles of amplification with denaturation at 95°C for 30 sec, annealing at 56°C for 30 sec, and extension at 72°C for 30 sec, followed by a final extension step of 7 min at 72°C. For quality control, 10% of the samples were randomly selected to evaluate the reproducibility of the genotyping procedure. The genotyping success rate was 100%.

Statistical methods

Continuous variables were expressed as means ± standard deviation, and categorical variables were expressed as N (%) of study participants. Differences of the characteristics between patients with gestational diabetes mellitus and control subjects were compared using the Chisquare test or student t-test. Whether confirming with the Hardy-Weinberg equilibrium was analyzed by the Fisher's exact test. The association between GSTM1, GSTT1 and GSTP1 IIe1-05Val and risk of gestational diabetes mellitus was analyzed using conditional logistic regression analysis. The results were evaluated by the odds ratio (OR) and 95% confidence intervals (CIs). The wide-type genotype of GSTM1, GSTT1 and GSTP1 IIe105Val was considered as reference group. Statistical analysis was conducted using the SPSS 17.0 package (SPSS Inc., Chicago, IL, USA). P<0.05 was considered to indicate a significant difference.

GSTs polymorphisms and gestational diabetes mellitus

		0			,	
Variables	Patients N=320	~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~		%	Chi-square test or student t-test	P value
Age, years	32.	23 ±10.43	27	.35± 11.53	4.50	<0.001
<30	135	42.19	224	62.57		
≥30	185	57.81	134	37.43	28.18	< 0.001
Gestational age		26.16±5.11		26.35±5.35	0.47	0.32
BMI, kg/m ²		30.62±6.14		27.65±5.26	6.72	<0.001
<28	107	33.44	232	64.80		
≥28	213	66.56	126	35.20	66.49	<0.001
Fasting plasma glucosa, mg/dl		92.30±24.25		75.65±15.32	10.80	<0.001
HOMA-IR		4.57±2.75		3.44±2.01	6.15	<0.001
TG, mg/dl		261.10±115.20		265.55±112.75	0.51	0.31
TC, mg/dl		232.47±48.60		238.42±44.42	1.67	0.05
HDL-C, mg/dl		51.52±13.25		52.15±12.68	0.63	0.26
LDL-C, mg/dl		121.43±26.20		124.37±25.36	1.48	0.07

Table 1. Characteristics between patients with gestational diabetes mellitus and control subjects

BMI: body mass index; HOMA-IR: homeostasis model assessment index for insulin resistance; TG: Triglycerides; TC: Total Cholesterol; HDL-C: High-density lipoprotein cholesterol; LDL-C: Low-density lipoprotein cholesterol.

Table 2. Association between GSTM1,	GSTT1 and GSTP1 Ile105Va	I polymorphisms and development
of gestational diabetes mellitus		

Genotypes	Patients N=320	%	Controls N=358	%	Chi-square test	P value	HWE	OR (95% CI) ¹	P value
GSTM1									
Present	171	53.44	237	66.20				1.0 (Reference)	-
Null	149	46.56	121	33.80	11.49	0.001		1.71 (1.24-2.36)	0.001
GSTT1									
Present	178	55.63	256	71.51				1.0 (Reference)	-
Null	142	44.38	102	28.49	18.50	<0.001		2.00 (1.44-2.79)	< 0.001
GSTP1 lle105Val									
Codominant									
lle/lle	145	45.31	178	49.72				1.0 (Reference)	-
lle/Val	142	44.38	152	42.46				1.15 (0.82-1.59)	0.39
Val/Val	33	10.31	28	7.82	2.00	0.37	0.57	1.45 (0.81-2.61)	0.19
Dominant									
lle/lle+lle/Val	287	89.69	330	92.18				1.0 (Reference)	-
Val/Val	33	10.31	28	7.82	1.28	0.26		1.36 (0.77-2.39)	0.26
Recessive									
lle/lle	145	45.31	178	49.72				1.0 (Reference)	-
lle/Val+Val/Val	175	54.69	180	50.28	1.32	0.25		1.19 (0.87-1.63)	0.25

¹Adjusted for age, BMI, fasting plasma glucosa and HOMA-IR.

Results

The characteristics of patients with gestational diabetes mellitus and control subjects were observed in **Table 1**. The mean ages of patients and controls were 32.23 ± 10.43 and 27.35 ± 11.53 years, respectively. By Chi-square test or

student t-test, patients with gestational diabetes mellitus were more likely to have higher age (t=4.50, P<0.001), BMI (t=6.72, P<0.001), fasting plasma glucose (t=10.80, P<0.001) and HOMA-IR (t=6.15, P<0.001), when compared with the control subjects. However, no significant differences between patients with gesta-

Variables	Patients		Controls		OR (95% CI)	P value
	Present	Null	Present	Null	-	
Age, years						
<30	79	56	149	75	1.41 (0.88-2.24)	0.13
≥30	38	147	29	105	1.07 (0.60-1.90)	0.81
BMI, kg/m²						
<28	65	42	145	87	1.08 (0.65-1.77)	0.76
≥28	52	161	33	93	1.10 (0.64-1.87)	0.72
Fasting plasma glucosa, mg/dl						
<80	59	45	131	81	1.23 (0.74-2.04)	0.39
≥80	58	158	47	99	1.29 (0.79-2.10)	0.27
HOMA-IR						
<4.0	75	55	146	92	1.16 (0.73-1.84)	0.49
≥4.0	42	148	32	88	1.28 (0.73-2.25)	0.36

 Table 3. Interaction between GSTM1 polymorphism and demographic and clinical characteristics in the risk of gestational diabetes mellitus

Table 4. Interaction between GSTT1 polymorphism and demographic and clinical characteristics in
the risk of gestational diabetes mellitus

Variables	Patie	Patients		ols	OR (95% CI)	P value
	Present	Null	Present	Null		
Age, years						
<30	82	53	143	81	1.14 (0.72-1.81)	0.56
≥30	35	150	35	99	1.52 (0.86-2.67)	0.13
BMI, kg/m²						
<28	69	38	150	82	1.01 (0.60-1.67)	0.98
≥28	48	165	28	98	0.98 (0.56-1.72)	0.95
Fasting plasma glucosa	, mg/dl					
<80	61	43	131	81	1.14 (0.69-1.89)	0.59
≥80	56	160	47	99	1.36 (0.83-2.21)	0.19
HOMA-IR						
<4.0	83	47	155	83	1.06 (0.66-1.69)	0.81
≥4.0	34	156	23	97	1.09 (0.58-2.03)	0.78

tional diabetes mellitus and control subjects in terms of gestational age (t=0.47, P=0.32), and levels of TG (t=0.51, P=0.31), TC (t=1.67, P=0.05), HDL-C (t=0.63, P=0.26) and LDL-C (t=1.48, P=0.07).

By Fisher's exact test, we found the genotype distributions of *GSTP1* IIe105Val were in line with the Hardy-Weinberg equilibrium in control subjects (P=0.57) (**Table 2**). By chi-square test, we found significant differences between the present and null genotype distributions of *GSTM1* (χ^2 =11.49, P=0.001). Moreover, significant differences were also observed between the present and null genotype distributions of

GSTT1 (χ^2 =18.50, P<0.001). Using unconditional logistic analysis, individuals carrying the null genotypes of GSTM1 and GSTT1 were associated with an increased risk of gestational diabetes mellitus when compared with the present genotype, and the adjusted Ors (95% Cl) were 1.71 (1.24-2.36) and 2.00 (1.44-2.79), respectively. However, the GSTP1 Ile105Val polymorphism was not associated with an elevated risk of gestational diabetes mellitus.

We also performed a gene-environmental association of *GSTM1* and *GSTT1* polymorphisms with age, BMI, fasting plasma glucosa and HOMA-IR for the risk of gestational diabetes mellitus (**Tables 3** and **4**). We found null and present genotypes of *GSTM1* and *GSTT1* were not significant associated with increased risk of gestational diabetes mellitus regardless of age, BMI, fasting plasma glucosa and HOMA-IR.

Discussion

It is well known that individuals would not develop the same type of disease although they are exposing to the same environmental and lifestyle factors. Therefore, inherit factors may contribute to the development of diseases. Single nucleotide polymorphisms refer to the gene sequence of a single nucleotide bases inserting, missing or replacing to cause the polymorphism of the nucleic acid sequence [20]. Among millions of SNPs, the incidence of gene polymorphism is more than 1%, including transition, transversion, and insertion and deletion of single nucleotide. Gene polymorphisms could alter the gene expression, structure and quantity of the products, and thus influence the function of the gene [21, 22]. In the present study, we conducted a case-control study to investigate whether GSTM1, GSTT1 and GSTP1 Ile105Val polymorphisms could influence the susceptibility to gestational diabetes mellitus. We found that the GSTM1 null genotype and GSTT1 null genotype were associated with an increased risk of gestational diabetes mellitus.

The GSTM1 locus is located on chromosome 1p13.3, and the GSTT1 locus existed on chromosome 22q11.2 [23]. The GSTM1 and GSTT1 contribute to catalyzing the conjugation of glutathione to a variety of hydrophobic and electrophilic substrates and carcinogens such as benzpyrene and reactive oxygen species. The polymorphisms in GSTs could reduce or eliminate the detoxification activities of enzyme, and thus increase the susceptibility to disease, such as gestational diabetes mellitus. Previous studies have reported that polymorphisms in GSTs play an important role in the development of many diseases, such as type 2 diabetes, hypertension and nephropathy [24-26]. Abbas et al. conducted a study with 138 essential hypertension patients and 116 control subjects in India, and they reported that GSTM1 null or positive genotype and GSTT1 null or positive genotype were associated with the development of essential hypertension [24]. Pinheiro et al. conducted a study with 120

patients and 147 healthy individuals in Brazil, and they found that GSTM1 null and GSTT1 null genotypes may contribute to the susceptibility of T2DM patients [25]. Petrovič et al. conducted a study in 1015 Slovenian subjects with type 2 diabetes with or without essential arterial hypertension, and they reported that GSTM1-null and GSTT1-null genotypes contributed to the development of the essential arterial hypertension in patients with type 2 diabetes [26]. However, some studies reported inconsistent results [27]. Moasser et al. investigated the association between GSTM1 and GSTT1 gene polymorphisms and risk of diabetic retinopathy in Iranian population, and they suggested that GSTM1 and GSTT1 gene polymorphisms were associated with the pathogenesis of type 2 diabetes mellitus retinopathy [27].

Currently, only one previous study reported the association between polymorphisms of GSTs and risk of gestational diabetes mellitus [18]. This study conducted in a population with 50 gestational diabetes mellitus and 50 control subjects, and they found that GSTM1 and GSTT11 gene polymorphisms are not associated with the risk of gestational diabetes mellitus [18]. In our study, we found that the GSTM1 null genotype and GSTT1 null genotype were correlated with the development of gestational diabetes mellitus, which is inconsistent with the results of Orhan's. The discrepancies of the reported results might be caused by differences in ethnicities, selection of patients and controls, study design, or sample size. Further studies with large sample sizes are greatly needed to confirm our results.

We identified three limitations in our study. First, patients were selected from a single hospital, which might not be representative of the general population. Second, other genetic polymorphisms might have influenced the risk of gestational diabetes mellitus in addition to GSTs gene polymorphisms. Third, the sample size of this study was relatively small, which could limit the statistical power to identify the differences between groups. Further studies with large sample sizes are greatly needed to clarify the association of GSTs gene polymorphisms with the risk of gestational diabetes mellitus.

In conclusion, we suggest that the GSTM1 null genotype and GSTT1 null genotype are corre-

lated with an increased risk of gestational diabetes mellitus in a Chinese population. This finding could be helpful for identifying the genetic characteristics of gestational diabetes mellitus and developing more efficient strategies for prevention and treatment.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Qinggui Zhang, Department of Endocrinology, The First Affiliated Hospital of Xinxiang Medical University, 88 Jiankang Road, Weihui 453100, Henan, China. Tel: +86-373-4402408; Fax: +86-373-4402408; E-mail: zhanggq5@sina.com

References

- [1] Metzger BE, Buchanan TA, Coustan DR, de Leiva A, Dunger DB, Hadden DR, Hod M, Kitzmiller JL, Kjos SL, Oats JN, Pettitt DJ, Sacks DA, Zoupas C. Summary and recommendations of the fifth international workshop-conference on gestational diabetes mellitus. Diabetes Care 2007; 30 Suppl 2: S251-60.
- [2] Schmidt MI, Duncan BB, Reichelt AJ, Branchtein L, Matos MC, Costa e Forti A, Spichler ER, Pousada JM, Teixeira MM, Yamashita T; Brazilian Gestational Diabetes Study Group. Gestational diabetes mellitus diagnosed with a 2-h 75-g oral glucose tolerance test and adverse pregnancy outcomes. Diabetes Care 2001; 24: 1151-5.
- [3] Murgia C, Berria R, Minerba L, Malloci B, Daniele C, Zedda P, Ciccotto MG, Sulis S, Murenu M, Tiddia F, Manai M, Melis GB. Gestational diabetes mellitus in Sardinia: results from an early, universal screening procedure. Diabetes Care 2006; 29: 1713-4.
- [4] Dabelea D, Snell-Bergeon JK, Hartsfield CL, Bischoff KJ, Hamman RF, McDuffie RS; Kaiser Permanente of Colorado GDM Screening Program. Increasing prevalence of gestational diabetes mellitus (GDM) over time and by birth cohort: Kaiser Permanente of Colorado GDM Screening Program. Diabetes Care 2005; 28: 579-584.
- [5] Anna V, van der Ploeg HP, Cheung NW, Huxley RR, Bauman AE. Sociodemographic correlates of the increasing trend in prevalence of gestational diabetes mellitus in a large population of women between 1995 and 2005. Diabetes Care 2008; 31: 2288-2293.
- [6] Sella T, Shalev V, Elchalal U, Chovel-Sella A, Chodick G. Screening for gestational diabetes in the 21st century: a population-based cohort

study in Israel. J Matern Fetal Neonatal Med 2013; 26: 412-416.

- [7] Chu SY, Callaghan WM, Kim SY, Schmid CH, Lau J, England LJ, Dietz PM. Maternal obesity and risk of gestational diabetes mellitus. Diabetes Care 2007; 30: 2070-2076.
- [8] Ben-Haroush A, Yogev Y, Hod M. Epidemiology of gestational diabetes mellitus and its association with Type 2 diabetes. Diabet Med 2004; 21: 103-113.
- [9] Savitz DA, Janevic TM, Engel SM, Kaufman JS, Herring AH. Ethnicity and gestational diabetes in New York City, 1995-2003. BJOG 2008; 115: 969-978.
- [10] Ao D, Wang HJ, Wang LF, Song JY, Yang HX, Wang Y. The rs2237892 Polymorphism in KCNQ1 Influences Gestational Diabetes Mellitus and Glucose Levels: A Case-Control Study and Meta-Analysis. PLoS One 2015; 10: e012-8901.
- [11] Khan IA, Shaik NA, Kamineni V, Jahan P, Hasan Q, Rao P. Evaluation of Gestational Diabetes Mellitus Risk in South Indian Women Based on MTHFR (C677T) and FVL (G1691A) Mutations. Front Pediatr 2015; 3: 34.
- [12] Takhshid MA, Zahediannejad Z, Aboualizadeh F, Moezzi L, Ranjbaran R. G22A Polymorphism of Adenosine Deaminase and its Association with Biochemical Characteristics of Gestational Diabetes Mellitus in an Iranian Population. Iran J Med Sci 2015; 40: 170-4.
- [13] Khan IA, Movva S, Shaik NA, Chava S, Jahan P, Mukkavali KK, Kamineni V, Hasan Q, Rao P. Investigation of Calpain 10 (rs2975760) gene polymorphism in Asian Indians with Gestational Diabetes Mellitus. Meta Gene 2014; 2: 299-306.
- [14] Shi X, Cai Q, Zou M, Shen Y. Correlation between TCF7L2 gene polymorphism and genetic susceptibility in women with gestational diabetes mellitus. Zhonghua Fu Chan Ke Za Zhi 2014; 49: 588-93.
- [15] Hayes JD, Flanagan JU, Jowsey IR. Glutathione transferases. Annu Rev Pharmacol Toxicol 2005; 45: 51-88.
- [16] Strange RC, Lear JT, Fryer AA. Glutathione S-transferase polymorphisms: influence on susceptibility to cancer. Chem Biol Interact 1998; 111-112: 351-364.
- [17] Strange RC, Jones PW, Fryer AA. Glutathione S-transferase: genetics and role in toxicology. Toxicol Lett 2000; 112-113: 357-363.
- [18] Orhan O, Atalay MA, Orhan F, Karkucak M, Centinkaya Demir B, Yakut T, Cengiz C. Glutathione s-transferase m1 and t1 gene polymorphisms are not associated with increased risk of gestational diabetes mellitus development. West Indian Med J 2014; 63: 300-6.

- [19] Association AD. Diagnosis and classification of diabetes mellitus. Diabetes Care 2008; 31 Suppl 1: S55-60.
- [20] Friedberg EC. DNA damage and repair. Nature 2003; 421: 436-440.
- [21] Bartek J, Lukas J. DNA repair: Damage alert. Nature 2003; 421: 486-488.
- [22] Aas PA, Otterlei M, Falnes PO, Vågbø CB, Skorpen F, Akbari M, Sundheim O, Bjørås M, Slupphaug G, Seeberg E, Krokan HE. Human and bacterial oxidative demethylases repair alkylation damage in both RNA and DNA. Nature 2003; 421: 859-863.
- [23] Guengerich FP. Characterization of human cytochrome P450 enzymes. FASEB J 1992; 6: 745-748.
- [24] Abbas S, Raza ST, Chandra A, Rizvi S, Ahmed F, Eba A, Mahdi F. Association of ACE, FABP2 and GST genes polymorphism with essential hypertension risk among a North Indian population. Ann Hum Biol 2014; 30: 1-9.

- [25] Pinheiro DS, Rocha Filho CR, Mundim CA, Júnior Pde M, Ulhoa CJ, Reis AA, Ghedini PC. Evaluation of glutathione S-transferase GSTM1 and GSTT1 deletion polymorphisms on type-2 diabetes mellitus risk. PLoS One 2013; 8: e76262.
- [26] Petrovič D, Peterlin B. GSTM1-null and GSTT1null genotypes are associated with essential arterial hypertension in patients with type 2 diabetes. Clin Biochem 2014; 47: 574-7.
- [27] Moasser E, Azarpira N, Shirazi B, Saadat M, Geramizadeh B. Genetic polymorphisms of glutathione-s-transferase M1 and T1 genes with risk of diabetic retinopathy in Iranian population. Iran J Basic Med Sci 2014; 17: 351-6.