Original Article Association of apolipoprotein E with the progression of hepatitis B virus-related liver disease

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Abstract: Hepatitis B virus (HBV) infection increases the risk of liver decompensation, cirrhosis and hepatocellular carcinoma (HCC). Apolipoprotein E (ApoE), one of the major cholesterol carriers, plays important role in the metabolism of lipoprotein and the regulation of immune response. The present study was aimed to explore whether the genetic variation in ApoE gene affected disease progression in HBV infected individuals. We collected sera samples from healthy volunteers (n=40), inactive HBV carriers (n=30), and patients with acute hepatitis (n=60), severe hepatitis (n=12), HBV-related liver cirrhosis (n=58) or primary HCC (n=39). We found that ApoE and interlukin-6 (IL-6) was progressively increased, while IL-2 was gradually decreased with the increasing grade of disease severity. Furthermore, high ApoE levels in HBV infected individuals were correlated with increased IL-6 and decreased IL-2 levels, indicating immune abnormalities in these patients. The frequency of E3/3 genotype was progressively increased from carriers group, hepatitis group to progressive group (cirrhosis and HCC). The serum levels of low-density lipoprotein cholesterol (LDL-C) differed among ApoE phenotypes, with E3/4, E4/4> E3/3>E2/3. Our study suggested that ApoE may have a role in the pathogenesis and progression of HBV-related liver disease and indicated the possible underlying mechanisms.

Keywords: Hepatitis B virus, apolipoprotein E, polymorphisms

Introduction

Hepatitis B. an infectious disease caused by hepatitis B virus (HBV), is widely prevalent in the world especially in developing country. HBV infection increases the risk of liver decompensation, cirrhosis and hepatocellular carcinoma (HCC). According to World Health Organization (WHO) statistics, more than 2 billion people have been infected with HBV and approximately 378 million are chronic carriers and at risk for HBV-related liver disease worldwide. HBV results in 500,000 to 700,000 deaths each year [1]. HBV infection has a wide clinical spectrum ranging from inactive HBV carrier state to active hepatitis, severe hepatitis, HBV-related liver cirrhosis and primary HCC associated with HBV infection [2, 3]. A variety of risk factors, including older age, male gender, longer duration of HBV infection, high HBV-DNA levels and alcohol consumption, have been identified associated with the progressive course of this disease [4]. Furthermore, previous investigations suggest that host genetic polymorphism, which is responsible for differential immune response during HBV infection, may affect the individual susceptibility to infectious pathogens [5-7].

Apolipoprotein E (ApoE) is one of the major cholesterol carriers and synthesized principally in the liver [8]. ApoE plays important role in the metabolism of lipoprotein and the regulation of immune response [9]. The ApoE gene is mapped to chromosome 19 in band 19g13.2. The ApoE gene is polymorphic with three common codominant alleles (E2, E3 and E4), resulting in six different genotypes (E2/2, E2/3, E2/4, E3/3, E3/4 and E4/4) [10]. The three major isoforms, ApoE2 (cys112, cys158), ApoE3 (cys112, arg158) and ApoE4 (arg112, arg158), are encoded by E2, E3 and E4, respectively. Although these three major isoforms differ from one another by only one or two amino acids substitutions, these differences alter the structure and function of ApoE [11-13]. ApoE polymor-

Probe	Sequence	Size (bp)
rs429358_ modify	P-CACGTCCTCCATGTCCGCGCTTTTTTTTTTTTTTTTTTT	
rs429358_C	TTTTTTTTTTTTTTTTTCGGTACTGCACCAGGCGGCCGCG	92
rs429358_T	TTTTTTTTTTTTTTTTTTTCGGTACTGCACCAGGCGGCCGCA	94
rs7412_ modify	P-CTTCTGCAGGTCATCGGCATTTTTTTTTTTTTTTTTTTT	
rs7412_C	TTTTTTTTTTTTTTTTTTTTCCGGCCTGGTACACTGCCAGGCG	97
rs7412_T	TTTTTTTTTTTTTTTTTTTTCCGGCCTGGTACACTGCCAGGCA	99

Table 1. The probes of ligase detection reactions

Table 2. Distributions of AopE alleles

	Group						_
	Normal	Carrier	Active hepatitis	Severe hepatitis	Cirrhosis	HCC	All
E2	3 (3.8%)	4 (5.7%)	6 (5.0%)	0 (0)	1 (0.9%)	3 (3.9%)	17
E3	70 (87.5%)	56 (80.0%)	105 (87.5%)	20 (83.3%)	111 (97.4%)	67 (88.2%)	429
E4	7 (8.8%)	10 (14.3%)	9 (7.5%)	4 (16.7%)	2 (1.8%)	6 (7.9%)	38
All	80	70	120	24	114	76	484

phisms have been linked to the progression of cardiovascular disease[8] and Alzheimer's disease [11, 14]. Several studies demonstrated the association of ApoE genotypes with the outcomes of viral diseases, including herpex simplex virus (HSV) [15], hepatits C virus (HCV) [16-20] and HBV [7, 21-23]. However, few studies have been performed on the correlation of ApoE genotypes with disease progression in people with HBV infection.

In the present study, we hypothesized that genetic variation in *ApoE* gene could affect disease progression in HBV infected individuals. We determined the ApoE genotype distribution in healthy volunteers, inactive HBV carriers, and patients with acute hepatitis, chronic hepatitis, HBV-related liver cirrhosis or primary HCC, and analyzed the association of serum levels of interleukin-2 (IL-2), IL-6, high-density lipoprotein cholesterol (HDL-C) and low-density lipoprotein cholesterol (LDL-C) with ApoE concentration or polymorphisms. Our study revealed the possible mechanisms of the involvement of ApoE in the pathogenesis and progression of HBV infected individuals.

Materials and methods

Study patients

A total of 199 subjects, which included 30 HBV carriers, 60 patients with active hepatitis, 12 patients with severe hepatitis, 58 patients with HBV-related liver cirrhosis and 39 patients with HCC, admitted to Zhejiang Hospital (Hangzhou, China) from June 2012 to December 2013 were enrolled in this study. There were 107 male (32.14 ± 8.63 years, range 17-79 years) and 92 female $(31.47 \pm 7.16$ years, range 20-73 years). Diagnosis was based on the criteria explained in viral hepatitis prevention and control program revised on the national infectious and parasitic Conference (Xi'an) in 2000. The studies excluded patients with other hepatitis virus infection, other malignant cancer, autoimmune liver disease, alcoholic liver disease, nonalcoholic fatty liver disease, hyperlipidemia, coronary heart disease or atherosclerosis. Sera samples were obtained from these patients before treatment. Samples from 40 healthy volunteer, without history of liver disease, were collected as a control. No significant differences were found in gender and age between the patients and normal control group. Ethical approval for the study was provided by the independent ethics committee, XX. Informed and written consent was obtained from each individual according to the ethics committee guidelines.

ApoE genotyping

To determine the ApoE genotype, we genotyped two single-nucleotide polymorphisms (SNPs; NCBI SNPs rs429358 and rs7412). Genomic DNA was extracted from whole blood samples using a QIAamp DNA Blood Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer's guidelines. ApoE genotyping was performed using the multiplexed ligation de-

	Group						
	Normal (n=40)	Carrier (n=30)	Active hepatitis (n=60)	Severe hepatitis (n=12)	Cirrhosis (n=58)	HCC (n=39)	All (n=239)
E2/3	3 (7.5%)	4 (13.3%)	6 (10.0%)	0 (0)	1 (1.8%)	2 (5.3%)	16
E2/4	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)	1 (2.6%)	1
E3/3	30 (75.0%)	19 (63.3%)	45 (75.0%)	8 (66.7%)	54 (94.7%)	31 (81.6%)	187
E3/4	7 (17.5%)	4 (13.3%)	9 (17.5%)	4 (33.3%)	2 (3.5%)	3 (7.9%)	29
E4/4	0 (0)	3 (10.0%)	0 (0)	0 (0)	0 (0)	1 (2.6%)	4
NA	0 (0)	0 (0)	0 (0)	0 (0)	1 (1.7%)	1 (2.6%)	2

 Table 3. Distributions of ApoE genotypes

Table 4. Association between ApoE polymorphism and HBV infection

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	Normal (n=30)	HBV persistence (n=199)	Р
Genotype			
E2/3	3 (7.5%)	13 (6.5%)	0.7525
E2/4	0 (0)	1 (0.5%)	
E3/3	30 (75.0%)	157 (78.9%)	
E3/4	7 (17.5%)	22 (11.1%)	
E4/4	0 (0)	4 (2.0%)	
NA	0 (0)	2 (1.0%)	

Note: Statistical analysis refers to a chi-square or Fisher's exact test, as appropriate.

tection reaction technique (LDR) as described previously [24]. Firstly, PCR products containing the SNPs were generated using the primers as follows: rs429358, forward: 5'-GCCTACAAATCGGAACTGGA-3' and reverse: 5'-CAGCTCCTCGGTGCTCTG-3'; rs7412, forward: 5'-TAAGCGGCTCCTCCGCGAT-3' and reverse: 5'-GCCCCGGCCTGGTACACTG-3'. Secondly, PCR products were treated with proteinase K at 37°C for 15 min, 55°C for 10 min, and 90°C for 10 min, to destroy any remaining DNA polymerase activity before the LDR. LDRs were performed by using Taq DNA ligase (New England Biolabs, Beverly, MA, USA) with the following reaction conditions: 95°C for 2 minutes, followed by 40 cycles of ligation (94°C for 15s followed 50°C for 25S). The Probe sequences for LDR genotyping were listed in Table 1. Finally, LDR amplification product was run on an ABI prism 3730 sequencer (Applied Biosystems, Foster City, CA, USA) and genotypes analyzed using Genemapper software (Applied Biosystems).

Measurement of serum indices

Serum ApoE levels were measured by a turbidimetric immunoassay with a commercial kit (Wako Pure Chemical Industries, Tokyo, Japan). Serum concentrations of HDL-C and LDL-C were determined by an enzymatic method according to the manufacturer's instructions (Sekisui Medical, Tokyo, Japan). All were performed on an automatic analyzer (AU5400, Olympus, Japan).

Enzyme-linked immunosorbent assay (ELISA) analysis

Serum concentrations of IL-2 and IL-6 were assessed by using ELISA assay (eBioscience, San Diego, CA, USA) following the instructions of the manufacturer. Plates were read at 450 nm using a microplate reader (Multiscan MS[™], Labsystems, Helsinki, Finland).

Statistical analysis

All statistical analyses were performed with SPSS software version 16.0 (Chicago, IL, USA). Statistical comparisons between conditions were conducted using a chi-square or Fisher's exact test, as appropriate. Data were reported as the means \pm SD, and comparisons were conducted using ANOVA. The relationships between two factors were assessed by Pearson correlation analysis. Differences were considered statistically significant when *P*<0.05.

Results

Associations between ApoE polymorphisms and HBV infection or HBV disease progression

The frequency of ApoE allele and genotype in healthy volunteers, HBV carriers, and patients with active hepatitis, severe hepatitis, cirrhosis and HCC was shown in Tables 2 and 3. The ApoE E3 allele and E3/3 genotype were the most frequent in all groups.

	Carrier (n=30)	Hepatitis (n=72)	Р	Progressive (CH+HCC)	Р	
			VS carrier	(n=97)	VS carrier	VS hepatitis
Genotype						
E2/3	4 (13.3%)	6 (8.3%)	0.0395*	3 (3.2%)	0.0106*	0.028*
E2/4	0 (0)	0 (0)		1 (1.1%)		
E3/3	19 (63.3%)	53 (73.6%)		85 (89.5%)		
E3/4	4 (13.3%)	13 (18.1%)		5 (5.3%)		
E4/4	3 (10.0%)	0 (0)		1 (1.1%)		
NA	0 (0)	0 (0)		2 (2.1%)		

 Table 5. Association between ApoE polymorphism and disease progression in the HBV infection group

Note: Statistical analysis refers to a chi-square or Fisher's exact test, as appropriate. **P*<0.05.

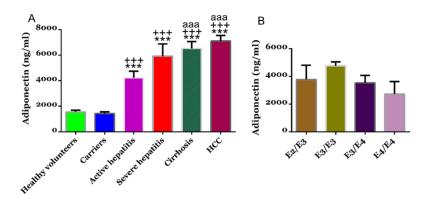


Figure 1. Apolipoprotein E (APOE) serum levels in study subjects. A. Serum was collected from healthy volunteers (n=40), HBV carriers (n=30), patients with active hepatitis (n=60), patients with severe hepatitis (n=12), patients with HBV-related liver cirrhosis (n=58), and patients with HCC (n=39) and APOE concentrations were determined by using a turbidimetric immunoassay. **P*<0.05, ****P*<0.001 VS healthy volunteers; ++*P*<0.01, +++*P*<0.001 VS HBV carriers; aaa*P*<0.001 VS patients with active hepatitis; sss*P*<0.001 VS patients with severe hepatitis; ccc*P*<0.001 VS patients with HBV-related liver cirrhosis. B. Serum APOE levels in subjects with different APOE genotypes.

We then evaluated whether ApoE polymorphism was related to HBV infection or disease progression in HBV infection. We could not find any association between normal control and HBV-infected group (**Table 4**), indicating that ApoE genotypes did not affect the susceptibility of human hepatitis B virus. Hepatitis group and progressive group (cirrhosis and HCC) were found significantly different from carriers group in the distribution of ApoE polymorphism. A significant difference was also found between hepatitis group and progressive group (**Table 5**).

ApoE serum concentration in study subjects

In order to investigate the functional significance of ApoE at protein level, we measured ApoE serum concentrations of the 239 subjects by a turbidimetric immunoassay. There was no significant differin ApoE serum ence healthy levels between volunteers and HBV carriers. Compared with healthy volunteers, the ApoE serum levels of patients with active hepatitis, severe hepatitis, cirrhosis and HCC increased to 2.72, 3.82, 4.21 and 4.58 fold, respectively (Figure 1A). We then analyzed whether ApoE serum levels correlated with ApoE genotypes (Figure 1B). No significant differences were detected ApoE serum levels in among the four ApoE genotypes, E2/E3, E3/E3, E3/E4 and E4/E4.

Serum IL-6 and IL-2 levels

To explore the changes of IL-6 and IL-2 with the progression of HBV-related liver disease, we detected their levels in healthy volunteers and patients with HBV-related liver disease (Figure 2). There were no significant difference in serum levels of IL-6 and IL-2 between healthy volunteers and HBV carriers. Comparing with healthy volunteers, the IL-6 serum levels of patients with active hepatitis, severe hepatitis, cirrhosis and HCC increased to 1.54, 2.56, 3.36 and 4.72 fold, respectively (Figure 2A), while the IL-2 serum levels decreased to 76%, 80%, 64% and 56%, respectively (Figure 2D).

ApoE is known to be involved in the regulation of immune response. We then analysis the correlation of ApoE with both cytokines by Pearson's analysis. ApoE serum levels were

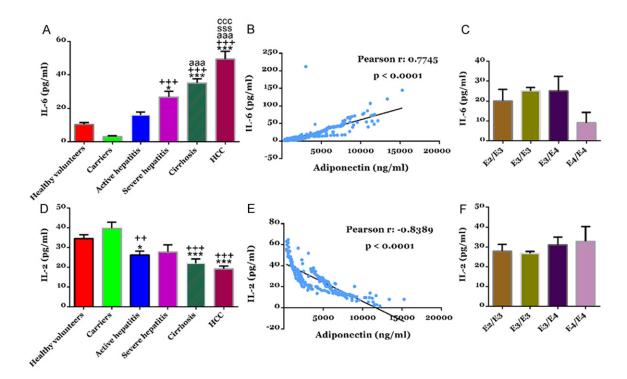


Figure 2. Serum levels of cytokines, IL-6 and IL-2 in study subjects. Serum IL-6 (A) and IL-2 (D) concentration were determined by using ELISA. ***P<0.001 VS healthy volunteers; +++P<0.001 VS HBV carriers; aaaP<0.001 VS patients with active hepatitis. The serum levels of APOE showed a significant positive correlation with IL-6 (B) and a significant negative correlation with IL-2 (E). Serum IL-6 (C) or IL-2 (F) levels in subjects with different APOE genotypes.

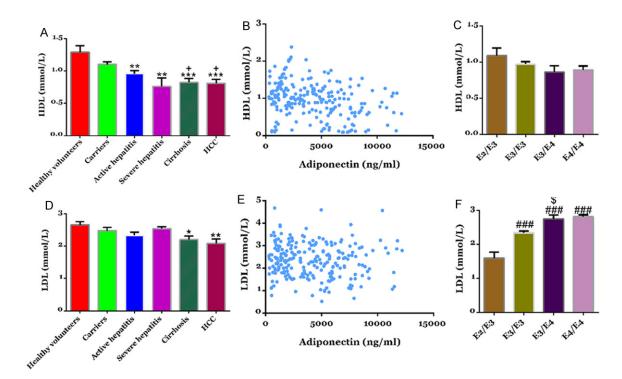


Figure 3. Serum levels of HDL-C and LDL-C in study subjects. Serum IL-6 (A) and IL-2 (D) concentration were determined by using ELISA. ***P*<0.01, ****P*<0.001 VS healthy volunteers; +*P*<0.05 VS HBV carriers. No correlation was observed between the serum levels of APOE and HDL-C (B) or LDL-C (E). Effects of APOE Phenotype on HDL-C (C) or LDL-C (F).

positively correlated with serum levels of IL-6 (r=0.7745, *P*<0.0001, **Figure 2B**), while negatively correlated with serum levels of IL-2 (r=-0.8389, *P*<0.0001, **Figure 2E**).

Further, we analyzed whether ApoE genotypes correlated with serum levels of IL-6 (**Figure 2C**) and IL-2 (**Figure 2F**). No significant differences were detected in the serum levels of both cytokines among the four ApoE genotypes, E2/E3, E3/E3, E3/E4 and E4/E4.

Serum HDL-C and LDL-C concentrations

ApoE is well known cholesterol transporter and plays an important role in lipid metabolism. We then detected serum concentrations of HDL-C and LDL-C in study subjects. As shown in Figure 3A, there were no significant difference in serum concentrations of HDL-C between healthy volunteers and HBV carriers. The HDL-C concentrations of patients with active hepatitis, severe hepatitis, cirrhosis and HCC were significantly lower than that of healthy volunteers. LDL-C concentrations showed no difference among healthy volunteers, HBV carriers, patients with active hepatitis and patients with severe hepatitis, while notable decrease of LDL-C concentrations was observed in the serum of patients with cirrhosis and HCC (Figure 3D). Pearson analysis revealed no correlation between the serum levels of ApoE and HDL-C (Figure 3B) or LDL-C (Figure 3E).

Further, there was no significant difference in the serum levels of HDL-C among the four ApoE genotypes, E2/E3, E3/E3, E3/E4 and E4/E4 (**Figure 3C**). LDL-C was progressively increased from E2/3 to E3/3 to E3/4 (**Figure 3F**).

Discussion

In a previous study, serum levels of ApoE were found significantly higher in patients with HBVrelated liver cirrhosis and HCC compared with that of the healthy controls [22]. Here, we collected serum samples from healthy volunteers, HBV carriers, and patients with active hepatitis, severe hepatitis, cirrhosis and HCC to further explored the changes of ApoE serum concentration with the progression of HBV-relative liver diseases. Our data demonstrated that ApoE was gradually increased with the increasing grade of disease severity (**Figure 1A**). E3 and E3/3 are the most common allele and genetic phenotype of APOE, respectively [22, 23]. Ahn et al. revealed that significantly more patients carrying the E3/3 genotype had developed liver cirrhosis [22]. Here, the frequency of E3/3 genotype was progressively increased from carriers group, hepatitis group to progressive group (cirrhosis and HCC) (**Table 5**). Our data further suggested that the serum levels and genotypes of ApoE were related with the progression of HBV-related liver diseases.

Cytokines are low-molecular-weight proteins regulate various inflammatory responses. It has been demonstrated that cytokine networks perform multiple functions in physiology and pathology [25]. Increased IL-6 [26] and decreased IL-2 [27, 28] were observed in chronic liver disease. In the present study, a progressive increase of IL-6 and a gradual down-regulation of IL-2 was observed with the increasing grade of disease severity (Figure 2). Furthermore, ApoE levels were positively correlated with IL-6 levels, but negatively related with IL-2 levels. IL-6 and IL-2 levels were not significantly different among ApoE phenotypes. These data suggested that high serum levels of ApoE, correlating with high IL-6 and low IL-2 could represent one of the factors leading to immune abnormalities in HBV-related diseases.

The liver plays a critical role in the synthesis and degradation of lipid and lipoprotein [29]. In various liver diseases, liver functions are impaired, leading to the broken of lipid and lipoprotein homeostasis. Here, we find a notable reduction of HDL-C in patients with hepatitis, cirrhosis and HCC (Figure 3A), and a decrease of LDL-C in patients with cirrhosis and HCC (Figure 3D). These results may be ascribed to hepatocellular dysfunction and has been confirmed earlier in other studies [30, 31]. Some reports have indicated that the E4 allele is associated with higher LDL-C and the E2 with lower LDL-C [32, 33]. In the present study, although the serum levels of ApoE was not correlated with the levels of HDL-C or LDL-C (P<0.05), LDL-C differed among ApoE phenotypes, with E3/4, E4/4> E3/3>E2/3 (Figure 3F), while no difference was observed in HDL-C among ApoE phenotypes (Figure 3C). Our data demonstrated the close relation of LDL-C levels with ApoE phenotypes.

Taken together, we demonstrated the association of serum concentration and polymorphisms of ApoE with the progression of HBVrelated liver disease. High serum levels of ApoE in patients with HBV-related liver disease may be related with immune abnormalities manifested by increased IL-6 and decreased IL-2. Moreover, ApoE genotypes influenced LDL-C concentration, which indicated the association of ApoE genotypes and lipid metabolism in these patients.

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Disclosure of conflict of interest

None.

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