# Original Article

# RUNX3 polymorphisms and the susceptibility to cervical cancer and cervical intraepithelial neoplasia in Western China

Qian-Qian Gao<sup>1</sup>, Bin Zhou<sup>2</sup>, Xiu-Zhang Yu<sup>1</sup>, Xi Zeng<sup>1</sup>, Zhu Zhang<sup>1</sup>, Yi Quan<sup>1</sup>, Yan-Yun Wang<sup>2</sup>, Yan Pu<sup>2</sup>, Peng Chen<sup>2</sup>, Ya-Ping Song<sup>2</sup>, Lin Zhang<sup>2</sup>, Ming-Rong Xi<sup>1</sup>

<sup>1</sup>Department of Obstetrics and Gynecology, West China Second University Hospital, Sichuan University, Chengdu, Sichuan, P. R. China; <sup>2</sup>Laboratory of Molecular Translational Medicine, West China Institute of Women and Children's Health, Key Laboratory of Obstetric & Gynecologic and Pediatric Diseases and Birth Defects of Ministry of Education, West China Second University Hospital, Sichuan University, Chengdu, Sichuan, P. R. China

Received February 27, 2016; Accepted May 22, 2016; Epub October 1, 2016; Published October 15, 2016

Abstract: Runt-related transcription factor 3 (RUNX3) is recognized to play essential roles in various tumors. But the effect between genetic variations and the susceptibility to tumor remains unclear, so does it in cervical cancer (CC) and cervical intraepithelial neoplasia (CIN). Three single nucleotide polymorphisms (SNP) of the RUNX3 gene were selected to evaluate whether they were associated with CC and CIN. The three polymorphisms were genotyped in 260 CC patients, 212 CIN patients and 286 healthy controls in a hospital based case-control study in Western China. The three SNPs of RUNX3 were analyzed between every two groups. We found TT genotype of rs760805 (P=0.01, OR=0.55, 95% CI=0.35-0.88) compared with AA/AT genotype and AG genotype of rs2236852 (P=0.0024, OR=1.72, 95% CI=1.25-2.5) compared with AA/GG genotype were significantly associated with susceptibility to CC. The former was also associated with the clinical stage of CC. We also found SNPs rs760805 (P=0.0042, OR=8.33, 95% CI=1.12-50) and rs7528484 (P=0.027, OR=0.14, 95% CI=0.027-0.72) were closely connected with HPV infection in CIN patients. Moreover, we found rs2236852 AG genotype of RUNX3 was more related with the progression from CIN to CC (P<0.0001, OR=2.27, 95% CI=1.54-3.45). RUNX3 mRNA expression was significantly decreased in CC (P<0.0001) while no significant relationship between RUNX3 mRNA expression and the three SNPs. Those results probably mean that the SNPs of RUNX 3 influence not only the genetic susceptibility to CC, but also the pathogenesis of CC and CIN in Western China. The decreased expression of RUNX3 mRNA might indicate the inhibition of CC.

Keywords: RUNX3, cervical cancer, cervical intraepithelial neoplasia, polymorphism

# Introduction

In less developed countries, cervical cancer (CC) is the second most common cancer and the third leading cause of cancer death among female until year 2012 [1]. It was estimated that there were 527,600 new CC cases and 265,700 deaths worldwide in 2012 [1]. Cervical intraepithelial neoplasia (CIN) is a precancerous lesion of CC [2]. Both CC and CIN are closely related to human papillomavirus (HPV), which is regarded to be the leading etiology [3]. However, not all HPV infections will induce cervical lesions-only with persistent HPV infection, some will develop to CIN or even progress to CC

[3, 4]. In previous studies, researchers showed that the polymorphisms in genes was associated with the susceptibility to HPV [5, 6] and also with the susceptibility to CIN and CC [5, 7-10]. In addition, in our previous studies, we also found interleukin 1 (*IL-1*) and interleukin 6 (*IL-6*) genes were associated with the susceptibility to CC [11, 12]. These findings strongly implied that genetic polymorphisms may have potential associations with CC.

Runt-related transcription factor 3 (RUNX3) is a member of Runt domain family [13] and is a key downstream effector of transforming growth factor beta ( $TGF\beta$ ) signaling pathway [14]. In

many previous studies, such as gastric, hepatocellular, and breast cancers, RUNX3 was recognized to play an essential role in tumor suppressing [15-18]. The primary inhibiting inactivation of RUNX3, which might induce oncogenesis, included promoter hypermethylation, inactivating mutations, gene deletions, protein mislocalization and so on [15, 18-22]. While some researchers thought RUNX3 was an oncogene [23, 24], and Lotem et al. speculated the gene played important functions in immunity and inflammation and might indirectly influence epithelial tumor development [25]. For figure out the relationship between RUNX3 and cancer, some studies found the polymorphisms of RUNX3 (rs760805 and rs2236852) were connected with bladder cancer and gastric cancer, and concluded the gene polymorphisms may affect tumor occurrence [26-28]. Byungho et al. found a risk-associated allele of rs-7528484 increased the distal promoter activity and possibly stimulated nuclear factor kappa-light-chain-enhancer of activated B cells (NF-kB) activity which may be associated with susceptibility to intestinal-type gastric cancer [26]. But so far, it is still not fully understand how RUNX3 regulate the signal pathway and affect tumorigenesis. Further, we knew little about the association of RUNX3 with CC and CIN. Therefore, in this study, we focus on exploring the association of two tag SNPs (rs760805, rs226852 at proximal promoter) and one dbSNP (rs7528484 at distal promoter) in the RUNX3 gene with the individual susceptibility to CC and CIN. And a hospital based casecontrol study was performed in Western China.

# Materials and methods

### Subjects

The study was approved by the ethics committee of the West China Second University Hospital of Sichuan University and all women signed informed consents before the research. This hospital based case-control study enrolled 260 women with CC (mean  $\pm$  SD, 45.89 $\pm$ 8.78) and 212 with CIN (mean  $\pm$  SD, 39.15 $\pm$ 9.15) (Table 1) from 22 to 72 years old in the West China Second University Hospital from January 2012 to December 2014. All patients met the study criteria as follows (i) had not been previously diagnosed with CC or other cancer; (ii) had a cervix; (iii) had no history of cervical sur-

gery; (iv) were not pregnant; and (v) were physically able to undergo routine pelvioscopy. Women were excluded if they never had sexual intercourse or had history of serious disease or cancer. 286 healthy women aged from 18 to 72 years old were recruited as control group when they had a regular gynecological examination (mean ± SD, 40.07±10.34). Diagnoses of all cases were confirmed by pathological diagnosis. All subjects were living in western China. We reviewed medical records for patients' characteristics, including age at diagnosis, menstrual status, pathological type, clinical stage, tumor differentiation, lymph node status, and parametrial invasion, vessel invasion and HPV infection status.

# Genotyping

DNA was extracted from patients' cervical smear using a DNA isolation kit from Bioteke (Peking, China) according to the instruction of manufacturer. Genotyping of RUNX3 polymorphisms was analyzed using the polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) method. The SNPs information of RUNX3, such as primers [27, 28], annealing temperature and restriction enzymes, were all summarized in Table 2. The PCR reactions were performed in a total volume of 10 μl, including 5 μl 2×Taq PCR Mastermix (Tiangen, Peking, China) and 50 ng of genomic DNA. PCR products for SNPs were digested for 3 hours and the digested PCR products were separated by a 6% polyacrylamide gel and stained with 1.0 mg/ml argent nitrate. We randomly selected about 20% samples to perform the repeated assays and the results were 100% concordant. The genotypes were confirmed by DNA sequencing analysis (TsingKe, Peking, China).

### RNA isolation and qRT-PCR

RUNX3 mRNA expression was analyzed in 114 CC and 66 controls. Total RNA was extracted and purified from blood samples using TRIzol® Reagent (Life Technologies, USA) according to the manufacturer's protocol. Reverse transcription-PCR (RT-PCR) were performed by one step RT-PCR kit (BIONEER, South Korea) followed the manufacturer instructions. Quantitative real-time PCR was carried out using SYBR green PCR Master Mix (Roche, Switzerland) and samples were amplified in a thermocycler

Table 1. The socio-demographic characteristics of the study subjects

		Cases	Р					
Characteristics	Controls	Cervical cancer (CC)	CIN	Controls vs CC	Controls vs CIN	CIN vs CC		
Sample size	286	260 (squamous=234, adeno- carcinoma=17, other=9)	212 (CIN1=55, CIN2=40, CIN3=117)	-	-			
Age (years), mean (SD) Menstrual status	40.07 (10.34)	45.89 (8.78)	39.15 (9.15)	<0.001***	0.318	<0.001***		
Premenopausae	275	182	209	<0.001***	0.17	<0.001***		
Postmenopause	11	78	3					
***P<0.001.	,							

Table 2. The SNPs informations of RUNX3

SNP ID	Primer (5'-3')	Annealing tem- perature (°C)	Restriction enzymes	Cuttable allele	Uncuttable allele
rs760805	F: TCTCCCACTCAGCAGTTCACAC	58.7°C	BstZ17I	A (152 and 22 bp)	T (174 bp)
	R: TACAGCTCTCAATATGCGCCAG				
rs2236852	F: TGGAGTGGCTCCCCTCTTTCTG	63.6°C	Ndel	A (100 and 20 bp)	G (120 bp)
	R: TATGGCAGGGCTGCCACCTC				
rs7528484	F: TGCGAGGCCCAGGGTGTTGA	60°C	HincII	C (107 and 18 bp)	T (125 bp)
	R: CATGGAAGGGCACTCTGGTG				

as follows: 95°C for 10 min (1 cycle), 95°C for 15 s, 60°C for 1 min (48 cycles). The primer information of *RUNX3* was as follows: sense GGGCGAGGGAAGAGTTTCAC and antisense GTCTGGTCCTCCAGCTTCTG (product=140 bp). Data were normalized for beta-actin ( $\beta$ -actin) expression with comparative threshold cycle method. Triplicate Ct values were averaged and the relative expression levels were determined as  $2^{\Delta\Delta Ct}$ .

# Statistical analysis

All data analyses were calculated by SPSS 13.0 statistical software (SPSS Inc, Chicago, IL, USA). The baseline characteristics of participants were assessed by Student's t test and Single-factor Pearson chi-square. The genotype and allele frequencies of SNPs rs760805, rs2236852 and rs7528484 were obtained by direct counting. Hardy-Weinberg equilibrium was evaluated by the chi-square test. Genotypic association tests including codominant, dominant, recessive and overdominant genetic models were completed using SNPstats [29] in a case-control pattern. The linkage disequilibrium (LD) between the polymorphisms was implemented by using SHEsis software http:// analysis.bio-x.cn/myAnalysis.php. Odds ratio (OR) and respective 95% confidence intervals (95% CI) were reported to assess the different effects between alleles and genotypes. *RUNX3* mRNA expression levels were compared between CC and controls using Mann-Whitney nonparametric test. Statistical comparisons of the relative expression of mRNA between the different genotypes of the three SNPs were performed with the Kruskal-Wallis test. It was regarded as statistically significant if *P* value < 0.05.

# Results

The socio-demographic characteristics of the study subjects are shown in **Table 1**. There were significant differences in the distribution of age between CC and controls (P<0.001), CC and CIN (P<0.001). The menstrual status was also significantly different. We analyzed the data after adjustments for age and menstrual states, but there were no significant differences between crude and adjusted values (the crude values are not shown in **Tables 3-5**).

## Analysis of allele frequencies

The three polymorphisms of rs760805, rs22-36852 and rs7528484 were all successfully genotyped in 260 CC patients, 212 CIN patients and 286 healthy control subjects. The genotype distribution of the three variants in con-

# RUNX3 polymorphisms in cervical cancer and CIN

**Table 3.** Genotype distributions of rs760805 in cases/controls and the association with CC/CIN risk estimates

					CC vs Cont	rol	CIN vs Contr	ol	CC vs CIN	
rs760805	Geno- type	CC cases N (%)	CIN cases N (%)	Controls N (%)	Logistic regression OR (95% CI) <sup>a</sup>	Р	Logistic regression OR (95% CI)	Р	Logistic regression OR (95% CI) <sup>a</sup>	Р
Genetic model										
Codominant	A/A	101 (38.9%)	74 (34.9%)	85 (29.7%)	1.00	0.01*	1.00	0.47	1.00	0.2
	A/T	122 (46.9%)	92 (43.4%)	133 (46.5%)	1.38 (0.93-2.06)		0.79 (0.53-1.19)		1.04 (0.68-1.61)	
	T/T	37 (14.2)	46 (21.7%)	68 (23.8%)	0.46 (0.27-0.76)		0.78 (0.48-1.27)		1.63 (0.92-2.86)	
Dominant	A/A	101 (38.9%)	74 (34.9%)	85 (29.75%)	1.00	0.016*	1.00	0.22	1.00	0.42
	A/T-T/T	159 (61.1%)	138 (65.1%)	201 (70.3%)	0.63 (0.44-0.92)		0.79 (0.54-1.15)		0.85 (0.56-1.27)	
Recessive	A/A-A/T	223 (85.8%)	166 (78.3%)	218 (76.2%)	1.00	0.01*	1.00	0.58	1.00	0.076
	T/T	37 (14.2%)	46 (21.7%)	68 (23.8%)	0.55 (0.35-0.88)		0.88 (0.58-1.35)		0.63 (0.38-1.05)	
Overdominant	A/A-T/T	138 (53.1%)	120 (56.6%)	153 (53.5%)	1.00	0.79	1.00	0.49	1.00	0.57
	A/T	122 (46.9%)	92 (43.4%)	133 (46.5%)	1.05 (0.74-1.49)		1.13 (0.79-1.46)		1.12 (0.76-1.64)	
Allele										
	Α	324 (62.3%)	240 (56.6%)	303 (53.0%)	1.00	0.002**	1.00	0.27	1.00	0.08
	Т	196 (37.7%)	184 (43.4%)	269 (47.0%)	0.68 (0.53-0.87)		0.86 (0.67-1.11)		0.79 (0.61-1.02)	

\*P<0.05, \*\*P<0.01, \*Adjusted by age and menstrual status.

**Table 4.** Genotype distributions of rs2236852 in cases/controls and the association with CC/CIN risk estimates

					CC vs Con	itrol	CIN vs Conti	rol	CC vs C	CIN		
rs2236852	Geno- type	CC cases N (%)	CIN cases N (%)	Controls N (%)	Logistic regression OR (95% CI) <sup>a</sup>	Р	Logistic regression OR (95% CI)	Р	Logistic regression OR (95% CI) <sup>a</sup>	Р		
Genetic model												
Codominant	G/G	60 (23.1%)	76 (35.9%)	90 (31.5%)	1.00	0.0098**	1.00	0.28	1.00	<0.0001**		
	A/G	165 (63.5%)	90 (42.5%)	142 (49.6%)	1.69 (1.12-2.56)		0.75 (0.50-1.12)		2.27 (1.45-3.57)			
	A/A	35 (13.5%)	46 (21.7%)	54 (18.9%)	0.93 (0.53-1.64)		1.01 (0.60-1.67)		0.98 (0.54-1.79)			
Dominant	G/G	60 (23.1)	76 (35.9%)	90 (31.5%)	1.00	0.05*	1.00	0.31	1.00	0.0049**		
	A/G-A/A	200 (76.9%)	136 (64.2%)	196 (68.5%)	1.49 (1.00-2.22)		0.82 (0.56-1.19)		1.85 (1.20-2.86)			
Recessive	G/G-A/G	225 (86.5%)	166 (78.3%)	232 (81.1%)	1.00	0.086	1.00	0.44	1.00	0.04*		
	A/A	35 (13.5%)	46 (21.7%)	54 (18.9%)	0.65 (0.40-1.06)		1.19 (0.76-1.85)		0.58 (0.34-0.98)			
Overdominant	G/G-A/A	95 (36.5%)	122 (57.5%)	144 (50.4%)	1.00	0.0024**	1.00	0.11	1.00	<0.0001**		
	A/G	165 (63.5%)	90 (42.5%)	142 (49.6%)	1.72 (1.25-2.5)		0.75 (0.52-1.08)		2.27 (1.54-3.45)			
Allele												
	Α	235 (45.2%)	182 (42.9%)	250 (43.7%)	1.00	0.63	1.00	0.85	1.00	0.51		
	G	285 (54.8%)	242 (57.1%)	322 (56.3%)	0.94 (0.74-1.19)		1.03 (0.80-1.33)		0.91 (0.70-1.18)			

\*P<0.05, \*\*P<0.01, \*Adjusted by age and menstrual status.

trols were all in accordance with the Hardy-Weinberg equilibrium, the P value were 0.249, 0.904 and 0.657 respectively. The genotype and allele frequencies of the three polymorphisms were analyzed in **Tables 3-5**. It was indicated the allele T of rs760805 (P=0.002, OR=0.68, 95% Cl=0.53-0.87) was observed to be associated with the risk of CC. But there was no significant association between CC risk and alleles of rs2236852 (P=0.63, OR=0.94, 95% Cl=0.74-1.19) or rs7528484 (P=0.49, OR=1.11, 95% Cl=0.85-1.47). In addition, there was no

statistically significant association between CIN risk and alleles of rs760805 (0.27, OR= 0.86, 95% CI=0.67-1.11), rs2236852 (P=0.85, OR=1.03, 95% CI=0.80-1.33) and rs7528484 (P=0.61, OR=1.09, 95% CI=0.81-1.45). And we neither found strong LD in both patients and controls about the three SNPs.

# Comparison between CC and controls

We compared the SNPs between CC and controls and found that a significantly reduced risk

**Table 5.** Genotype distributions of rs7528484 in cases/controls and the association with CC/CIN risk estimates

					CC vs Contr	ol	CIN vs Cont	rol	CC vs CIN		
rs7528484	Geno- CC cases CIN cases Controls  type N (%) N (%) N (%)  Controls  Logistic  regression F  OR (95% CI) <sup>a</sup>		Р	Logistic regression OR (95% CI)	Р	Logistic regression OR (95% CI) <sup>a</sup>	Р				
Genetic model											
Codominant	T/T	151 (58.1%)	113 (53.3%)	153 (53.5%)	1.00	0.34	1.00	0.16	1.00	0.06	
	C/T	91 (35%)	93 (43.9%)	115 (40.2%)	0.76 (0.52-1.10)		1.10 (0.76-1.59)		0.74 (0.49-1.10)		
	C/C	18 (6.9%)	6 (2.8%)	18 (6.3%)	1.06 (0.51-2.21)		0.45 (0.17-1.18)		2.13 (0.79-5.89)	)	
Dominant	T/T	151 (58.1%)	113 (53.3%)	153 (53.5%)	1.00	0.18	1.00	0.97	1.00	0.33	
	C/T-C/C	109 (41.9%)	99 (46.7%)	133 (46.5%)	0.78 (0.55-1.11)		1.01 (0.70-1.45)		0.83 (0.56-1.22)		
Recessive	T/T-C/T	242 (93.1%)	206 (97.2%)	268 (93.7%)	1.00	0.89	1.00	0.066	1.00	0.066	
	C/C	18 (6.9%)	6 (2.8%)	18 (6.3%)	1.05 (0.52-2.13)		0.43 (0.17-1.11)		2.44 (0.90-6.67)		
Overdominant	T/T-C/C	169 (65%)	119 (56.1%)	171 (59.8%)	1.00	0.15	1.00	0.41	1.00	0.072	
	C/T	91 (35%)	93 (43.9%)	115 (40.2%)	0.76 (0.53-1.10)		1.16 (0.81-1.67)		0.69 (0.47-1.03)		
Allele											
	С	127 (24.4%)	105 (24.8%)	151 (26.4%)	1.00	0.49	1.00	0.61	1.00	0.94	
	Т	393 (75.6%)	319 (75.2%)	421 (73.6%)	1.11 (0.85-1.47)		1.09 (0.81-1.45)		1.02 (0.76-1.37)		

<sup>&</sup>lt;sup>a</sup>Adjusted by age and menstrual status.

of CC was associated with the TT homozygous carriers of rs760805 in a codominant model, compared with AA genotype (P=0.01, OR=0.46, 95% CI=0.27-0.76). Then, compared with AA and AT homozygous carriers, TT genotype carriers also have a reduced CC risk in a recessive model (P=0.01, OR=0.55, 95% CI=0.35-0.88). In addition, there was significantly reduced CC susceptibility associated with allele T carriers (P=0.016, OR=0.63, 95% CI=0.44-0.92) in a dominant model (Table 3). For rs2236852, we found that, compared with GG homozygous, AG genotype may significantly increase the risk of CC in a codominant model (P=0.0098, OR=1.69, 95% CI=1.12-2.56). And compared with homozygous of AA and GG in an overdominant model, AG genotype carriers also increased the risk of CC (P=0.0024, OR=1.72, 95% CI=1.25-2.5) (**Table 4**). But for rs7528484, there was no statistically significant difference between CC and controls (Table 5). Further, we performed stratification analysis of genotype distribution with CC patients for different age, clinical stage, pathological type, tumor differentiation, lymph node status, parametrial invasion, vessel invasion and HPV type. As shown in **Table 6.** only AT genotype of rs760805 in an overdominant model was identified significant association between clinical stage I and stage II-IV of CC, compared with the homozygote AA/ TT genotypes (P=0.038, OR=0.6, 95% CI=0.36-0.97). While no significant association was detected between other clinical features of rs760805, rs2236852 and rs7528484 (data were not shown completely).

# Comparison between CIN and controls

We also compared the three SNPs between CIN and controls, but there was no significant difference of the risk of CIN (Tables 3-5). And we conducted stratified analyses with different age, HPV type and pathological type. Part of analysis results were summarized in Table 6. We found TT genotype carriers of rs760805 had a significant association between high risk type (HR) HPV infection and negative ones in codominant model (0.015, OR=9.09, 95%) CI=1.18-100) and recessive model (0.0042, OR=8.33, 95% CI=1.12-50.0). Moreover, CC genotype carrier for rs7528484 of CIN patients were significantly different between HR-HPV and negative ones in a codominant model (0.031, OR=0.18, 95% CI=0.03-0.96) and recessive model (0.027, OR=0.14, 95% CI=0.027-0.72). However, no significant association was detected between other clinical features and the three polymorphisms.

# Comparison between CC and CIN

Moreover, we compared CIN and CC for the three variants (**Tables 3-5**) and found that AG genotype carriers of rs2236852 also increased the risk of CC in the codominant model (P<0.0001, OR=2.27, 95% CI=1.45-3.57), overdominant model (P<0.0001, OR=2.27, 95%

# RUNX3 polymorphisms in cervical cancer and CIN

Table 6. Analysis of CC/CIN patients characteristics and polymorphisms of RUNX3

		rs760805									rs7528484																
Clinical	G	en	otype		Genetic model					Ge	enoty	ре		Genetic model													
features	AA	A AT TT	AA AT	AT TT	AA AT TT	AA AT TT	AA AT TT	AT TT	AT TT	AT TT	AT TT	AT TT	Dominant (AA vs AT/TT	)	Recessiv (AA/AT vs		Overdomina (AA/TT vs A		СС	СТ	TT	Dominant (TT vs CC/CT	.)	Recessive (TT/CT vs Co		Overdomina (TT/CC vs C	
				OR (95% CI)	Р	OR (95% CI)	Р	OR (95% CI)	Р				OR (95% CI)	Р	OR (95% CI)	Р	OR (95% CI)	Р									
CC Age																											
<45 (n=133	) 41%	4	5% 14%	1.24 (0.75-2.05)	0.4	1.12 (0.56-2.25)	0.74	1.16 (0.71-1.89)	0.55	5%	36%	59%	1.05 (0.64-1.72)	0.85	1.71 (0.64-4.55)	0.28	0.91 (0.54-1.51)	0.71									
≥45 (n=127	) 36%	49	9% 15%							9%	34%	57%															
Clinical stage																											
I (n=125)	34%	5	4% 12%	0.70 (0.42-1.15)	0.16	1.43 (0.70-2.89)	0.32	0.60 (0.36-0.97)	0.038*	6%	34%	59%	1.09 (0.67-1.79)	0.72	1.17 (0.45-3.07)	0.75	1.05 (0.63-1.75)	0.85									
II-IV (n=135	43%	4:	1% 16%							7%	36%	57%															
CIN Age																											
<40	32%	4:	3% 25%	0.79 (0.0.45-1.41)	0.42	0.69 (0.56-1.35)	0.27	1.07 (0.60-1.79)	0.9	3%	48%	49%	0.73 (0.42-1.25)	0.25	1.00 (0.19-4.76)	0.93	0.73 (0.42-1.25)	0.26									
≥40	38%	4	4% 18%							3%	40%	57%															
HPV type																											
HRHPV	33%	4:	3% 24%	1.72 (0.78-3.85)	0.18	8.33 (1.12-50.0)	0.0042**	0.74 (0.33-1.64)	0.46	2%	46%	52%	1.43 (0.64-3.23)	0.38	0.14 (0.027-0.72)	0.027*	2.17 (0.91-5.26)	0.071									
Negative	46%	50	0% 4%							11%	29%	61%															

\*P<0.05, \*\*P<0.01.

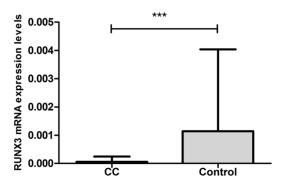


Figure 1. RUNX3 mRNA expression level was significantly decreased in CC (P<0.0001).

CI=1.54-3.45) and dominant model (P=0.0049, OR=1.85, 95% CI=1.2-2.86), but AA homozygous carriers might reduce the risk of CC in a recessive model (P=0.04, OR=0.58, 95% CI=0.34-0.98).

### mRNA expression level of RUNX3

At last, we analyzed the expression levels of *RUNX3* mRNA in CC with controls. *RUNX3* mRNA expression was significantly decreased in CC (P<0.0001) (**Figure 1**). However, no significant relationship was found between RUNX3 mRNA expression and polymorphisms of rs-760805, rs2236852 and rs7528484 in CC (P=0.4005, 0.67450 and 0.1437 respectively) or controls (P=0.8798, 0.7500 and 0.2990 respectively).

# Discussion

It is well known that CC and CIN are closely related to the HR-HPV infection [3]. But most infected women will eliminate virus and only a small part of infected women will progress to CIN or even CC [2-4]. In the previous studies, the immunological mechanism and genetic factors of hosts may play an important role in cervical lesions [30, 31]. In this study, we continued to explore the potential effect of gene for CIN and CC and selected three SNPs (2 tag SNPs from proximal promoter and 1 dbSNP from distal promoter) in the RUNX3 gene. According to the results, we conclude that RUNX3 is probably a potential gene for CC susceptibility. The polymorphisms of RUNX3 may be associated with HPV infection and CIN progression in Western China.

RUNX3 gene is a member of RUNX family and located in chromosome 1p36, which is deemed

to be a tumor suppress gene for gastric cancer and bladder cancer etc [13, 20, 21]. In previous study, Qing et al. found that gastric epithelial cells from RUNX3-/- nude mice with p53-/- background were tumorigenic, while those from RUNX3<sup>+/+</sup>p53<sup>-/-</sup> mice were not [16]. But controversially, some studies have demonstrated it over-expressed in carcinomas of head and neck, BCC and ovarian [23, 32, 33]. Nevadunsky et al. found RUNX3 had a role in cell proliferation and viability in ovarian cancer because of the overexpression of immunohistochemistry and qRT-PCR. In addition, the overexpressed RUNX3 in SKOV3 ovarian cancer cells resulted in increased cell viability while silencing RUNX3 expression by siRNA transfection resulted in a decrease in proliferation demonstrate the potential oncogenic role [32]. To date, scientists cannot fully elucidate the mechanism of RUNX3 with cancer and the genetic variations in RUNX3 that may affect signaling pathway for the development of cancer [17]. Researchers recognized RUNX3 is a downstream target gene of TGFB signaling which is a tumor suppress pathway. It regulates the expression of Bim and p21, negatively regulates VEGF, and thereby affects apoptosis, cell growth arrest and angiogenesis, respectively [14, 17]. In previous studies, only a few studies investigated on RUNX3 polymorphisms with cancer. The studies of bladder cancer and gastric cancer of an eastern Chinese population found that the genetic variants of proximal promoter (rs11249206 at intron 1, rs760805 at intron 3 and rs2236852 at intron4) in RUNX3 may modulate the risk of both cancer which may be identified with function in gene transcription and protein expression [27, 28]. In addition, in a Korean population study, the change of SNP rs7528484 located in the RUNX3 distal promoter may contribute differentially to intestinal-type gastric cancer and the proximal and distal promoters may have opposite regulatory actions [26]. But the specific affection of RUNX3 polymorphisms to cancer is still not clear and need further functional studies.

In this study, we found that TT genotype of rs760805 polymorphism was associated with a significantly reduced risk of CC in Western China, which was contrary to the previous studies of bladder and gastric cancer [27, 28]. Further, we found that rs2236852 AG genotype was related to a significantly increased risk of CC, the same as the discovery of Wu et al. [27]

but not as Suárez-Villanueva et al. [34]. In addition, we also investigated that rs2236852 was associated with CIN and CC and might predict CIN progression. After stratification analysis, we found rs760805 of RUNX3 was associated with not only the susceptibility to CC but also the clinical stage of CC and HR-HPV infection of CIN. And rs7528484 was also associated with the risk of HR-HPV infected in CIN patients. All these evidences suggested that the rs760805 polymorphism may be a contributor to pathogenesis of CC and CIN while the specific functions should be demonstrated. Further, we found the expression of RUNX3 mRNA decreased significantly in CC group. It might suggest RUNX3 inhibited the tumor growth [35, 36]. However, no relationship was found between the three SNPs and the low expression. It may be some other factors to regulate RUNX3 transcription rather than the polymorphisms. And it might also be the limitation of sample selections in our study. These findings also indicate that RUNX3 may play complex roles for different tumors, such as functions of immunity and inflammation, and indirectly influence tumor development [25]. In the future, the polymorphisms of RUNX3 might become useful diagnostic biomarkers for CC and CIN. We also look forward novel RUNX3 related genetic therapeutic could be considered for advanced CC. The results also encourage us to explore how does RUNX3 gene affect tumor and whether it could be used for therapy.

In conclusion, we demonstrated that *RUNX3* SNPs may play important roles with the susceptibility to CC and CIN. However, due to the restriction of samples, further studies are needed to explore the association between *RUNX3* and CC/CIN. Most importantly, further researches about functional evaluations of *RUNX3* are also needed to confirm our findings.

# Acknowledgements

This work was supported by the National Natural Science Foundation of China (No. 81572573, No. 81172440); and the Science Foundation for The Excellent Youth Scholars of Sichuan University (No. 2011SCU04A16).

#### Disclosure of conflict of interest

None.

Address correspondence to: Dr. Ming-Rong Xi, Department of Obstetrics & Gynecology, West China Second University Hospital, Sichuan University, Chengdu 610041, Sichuan, P. R. China. Tel: +86 28 85501633; Fax +86 28 85501633; E-mail: xmrjzz@ 126.com

#### References

- [1] Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J and Jemal A. Global cancer statistics, 2012. CA Cancer J Clin 2015; 65: 87-108.
- [2] McCredie MR, Sharples KJ, Paul C, Baranyai J, Medley G, Jones RW and Skegg DC. Natural history of cervical neoplasia and risk of invasive cancer in women with cervical intraepithelial neoplasia 3: a retrospective cohort study. Lancet Oncol 2008; 9: 425-434.
- [3] Berkhof J, de Bruijne MC, Zielinski GD and Meijer CJ. Natural history and screening model for high-risk human papillomavirus infection, neoplasia and cervical cancer in the Netherlands. Int J Cancer 2005; 115: 268-275.
- [4] Rodriguez AC, Burk R, Herrero R, Hildesheim A, Bratti C, Sherman ME, Solomon D, Guillen D, Alfaro M, Viscidi R, Morales J, Hutchinson M, Wacholder S and Schiffman M. The natural history of human papillomavirus infection and cervical intraepithelial neoplasia among young women in the Guanacaste cohort shortly after initiation of sexual life. Sex Transm Dis 2007; 34: 494-502.
- [5] Gonzalez-Herrera L, Rodriguez-Morales P, Gonza Lez-Losa Mdel R, Perez-Mendoza G, Canul-Canche J, Rosado-Lopez I and Cetina TC. MTHFR/p53 polymorphisms as genetic factors for cervical intraepithelial neoplasia and cervical cancer in HPV-infected Mexican women. Int J Biol Markers 2014; 29: e142-149.
- [6] Kuglik P, Kasikova K, Smetana J, Vallova V, Lastuvkova A, Moukova L, Cvanova M and Brozova L. Molecular cytogenetic analyses of hTERC (3q26) and MYC (8q24) genes amplifications in correlation with oncogenic human papillomavirus infection in Czech patients with cervical intraepithelial neoplasia and cervical carcinomas. Neoplasma 2015; 62: 130-139.
- [7] Grimm C, Watrowski R, Baumuhlner K, Natter C, Tong D, Wolf A, Zeillinger R, Leodolter S, Reinthaller A and Hefler L. Genetic variations of interleukin-1 and -6 genes and risk of cervical intraepithelial neoplasia. Gynecol Oncol 2011; 121: 537-541.
- [8] Natter C, Polterauer S, Rahhal-Schupp J, Cacsire Castillo-Tong D, Pils S, Speiser P, Zeillinger R, Heinze G and Grimm C. Association of TAP gene polymorphisms and risk of cervical

- intraepithelial neoplasia. Dis Markers 2013; 35: 79-84.
- [9] Kiran B, Karkucak M, Ozan H, Yakut T, Ozerkan K, Sag S and Ture M. GST (GSTM1, GSTT1, and GSTP1) polymorphisms in the genetic susceptibility of Turkish patients to cervical cancer. J Gynecol Oncol 2010; 21: 169-173.
- [10] Zidi S, Gazouani E, Stayoussef M, Mezlini A, Ahmed SK, Yacoubi-Loueslati B and Almawi WY. IL-10 gene promoter and intron polymorphisms as genetic biomarkers of cervical cancer susceptibility among Tunisians. Cytokine 2015; 76: 343-347.
- [11] Pu Y, Zhang Z, Zhou B, Chen P, Zhang K, Song Y, Gao Q, Wang K, Quan Y, Xi M and Zhang L. Association of an insertion/deletion polymorphism in IL1A 3'-UTR with risk for cervical carcinoma in Chinese Han Women. Hum Immunol 2014; 75: 740-744.
- [12] Quan Y, Zhou B, Wang Y, Duan R, Wang K, Gao Q, Shi S, Song Y, Zhang L and Xi M. Association between IL17 polymorphisms and risk of cervical cancer in Chinese women. Clin Dev Immunol 2012; 2012: 258293.
- [13] Lund AH and van Lohuizen M. RUNX: a trilogy of cancer genes. Cancer Cell 2002; 1: 213-215.
- [14] Ito Y and Miyazono K. RUNX transcription factors as key targets of TGF-beta superfamily signaling. Curr Opin Genet Dev 2003; 13: 43-47.
- [15] Hwang KT, Han W, Bae JY, Hwang SE, Shin HJ, Lee JE, Kim SW, Min HJ and Noh DY. Downregulation of the RUNX3 gene by promoter hypermethylation and hemizygous deletion in breast cancer. J Korean Med Sci 2007; 22 Suppl: S24-31.
- [16] Li QL, Ito K, Sakakura C, Fukamachi H, Inoue K, Chi XZ, Lee KY, Nomura S, Lee CW, Han SB, Kim HM, Kim WJ, Yamamoto H, Yamashita N, Yano T, Ikeda T, Itohara S, Inazawa J, Abe T, Hagiwara A, Yamagishi H, Ooe A, Kaneda A, Sugimura T, Ushijima T, Bae SC and Ito Y. Causal relationship between the loss of RUNX3 expression and gastric cancer. Cell 2002; 109: 113-124.
- [17] Subramaniam MM, Chan JY, Yeoh KG, Quek T, Ito K and Salto-Tellez M. Molecular pathology of RUNX3 in human carcinogenesis. Biochim Biophys Acta 2009; 1796: 315-331.
- [18] Xiao WH and Liu WW. Hemizygous deletion and hypermethylation of RUNX3 gene in hepatocellular carcinoma. World J Gastroenterol 2004; 10: 376-380.
- [19] Gao F, Huang C, Lin M, Wang Z, Shen J, Zhang H, Jiang L and Chen Q. Frequent inactivation of RUNX3 by promoter hypermethylation and protein mislocalization in oral squamous cell carcinomas. J Cancer Res Clin Oncol 2009; 135: 739-747.

- [20] Ito K, Liu Q, Salto-Tellez M, Yano T, Tada K, Ida H, Huang C, Shah N, Inoue M, Rajnakova A, Hiong KC, Peh BK, Han HC, Ito T, Teh M, Yeoh KG and Ito Y. RUNX3, a novel tumor suppressor, is frequently inactivated in gastric cancer by protein mislocalization. Cancer Res 2005; 65: 7743-7750.
- [21] Kim WJ, Kim EJ, Jeong P, Quan C, Kim J, Li QL, Yang JO, Ito Y and Bae SC. RUNX3 inactivation by point mutations and aberrant DNA methylation in bladder tumors. Cancer Res 2005; 65: 9347-9354.
- [22] Lau QC, Raja E, Salto-Tellez M, Liu Q, Ito K, Inoue M, Putti TC, Loh M, Ko TK, Huang C, Bhalla KN, Zhu T, Ito Y and Sukumar S. RUNX3 is frequently inactivated by dual mechanisms of protein mislocalization and promoter hypermethylation in breast cancer. Cancer Res 2006; 66: 6512-6520.
- [23] Kudo Y, Tsunematsu T and Takata T. Oncogenic role of RUNX3 in head and neck cancer. J Cell Biochem 2011; 112: 387-393.
- [24] Tsunematsu T, Kudo Y, Iizuka S, Ogawa I, Fujita T, Kurihara H, Abiko Y and Takata T. RUNX3 has an oncogenic role in head and neck cancer. PLoS One 2009; 4: e5892.
- [25] Lotem J, Levanon D, Negreanu V, Bauer O, Hantisteanu S, Dicken J and Groner Y. Runx3 at the interface of immunity, inflammation and cancer. Biochim Biophys Acta 2015; 1855: 131-143.
- [26] Lim B, Ju H, Kim M and Kang C. Increased genetic susceptibility to intestinal-type gastric cancer is associated with increased activity of the RUNX3 distal promoter. Cancer 2011; 117: 5161-5171.
- [27] Wu D, Tian Y, Gong W, Zhu H, Zhang Z, Wang M, Wang S, Tan M and Wu H. Genetic variants in the Runt-related transcription factor 3 gene contribute to gastric cancer risk in a Chinese population. Cancer Sci 2009; 100: 1688-1694.
- [28] Zhang Z, Wang S, Wang M, Tong N and Fu G. Genetic variants in RUNX3 and risk of bladder cancer: a haplotype-based analysis. Carcinogenesis 2008; 29: 1973-1978.
- [29] Sole X, Guino E, Valls J, Iniesta R and Moreno V. SNPStats: a web tool for the analysis of association studies. Bioinformatics 2006; 22: 1928-1929.
- [30] Li C, Ma C, Zhang W and Wang J. The immune function differences and high-risk human papillomavirus infection in the progress of cervical cancer. Eur J Gynaecol Oncol 2014; 35: 557-561.
- [31] Punt S, Houwing-Duistermaat JJ, Schulkens IA, Thijssen VL, Osse EM, de Kroon CD, Griffioen AW, Fleuren GJ, Gorter A and Jordanova ES. Correlations between immune response and

# RUNX3 polymorphisms in cervical cancer and CIN

- vascularization qRT-PCR gene expression clusters in squamous cervical cancer. Mol Cancer 2015; 14: 71.
- [32] Nevadunsky NS, Barbieri JS, Kwong J, Merritt MA, Welch WR, Berkowitz RS and Mok SC. RUNX3 protein is overexpressed in human epithelial ovarian cancer. Gynecol Oncol 2009; 112: 325-330.
- [33] Salto-Tellez M, Peh BK, Ito K, Tan SH, Chong PY, Han HC, Tada K, Ong WY, Soong R, Voon DC and Ito Y. RUNX3 protein is overexpressed in human basal cell carcinomas. Oncogene 2006; 25: 7646-7649.
- [34] Suarez-Villanueva S, Ayala-Madrigal ML, Peregrina-Sandoval J, Macias-Gomez N, Ramirez-Ramirez R, Muniz-Mendoza R, Moreno-Ortiz JM, Centeno-Flores M, Maciel-Gutierrez V, Cabrales and E MG-A. RUNX3 gene polymorphisms and haplotypes in Mexican patients with colorectal cancer. Genet Mol Res 2015; 14: 15505-15510.
- [35] Bae SC and Choi JK. Tumor suppressor activity of RUNX3. Oncogene 2004; 23: 4336-4340.
- [36] Menheniott TR, Judd LM and Giraud AS. RUNX3 methylation and anti-tumor immunity. Oncoscience 2015; 2: 789-790.