Original Article

MicroRNA-212-5p down-regulation suppresses colorectal cancer migration and invasion by up-regulating SMAD4

Mingmin Huang1*, Tengfei He2*, Chungen Xing2

¹VIP Ward 303, Nanjing Drum Tower Hospital, The Affiliated Hospital of Nanjing University Medical School, Nanjing, China; ²Department of General Surgery, The Second Affiliated Hospital of Soochow University, Suzhou, China. *Equal contributors.

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Abstract: The expression and exact roles of miR-212-5p in CRC and the underlying molecular mechanism is still unclear. This study we aimed to investigate the expression and the role of miR-212-5p in colorectal cancer and further explore the underlying molecular mechanism. We first detected the expression level of miR-212-5p in colorectal cancer tissues and cells and we found that miR-212-5p was up-regulated in colorectal cancer. We performed TargetScan and Miranda databases to predict the putative targets of miR-212-5p, and the prediction was verified by dual-luciferase reporter assay. To investigate the role of miR-212-5p in colorectal cancer, a stable miR-212-5p-low-expression cell line was established by using miR-212-5p inhibitor. MTT, cell migration and invasion assay were used to investigate the proliferation, cell migration and invasion ability of colorectal cancer HCT116 cells. Moreover, gene mRNA and protein expression were detected by qRT-PCR and western blotting. We found that miR-212-5p directly targets Smad4, and Smad4 was significantly increased after miR-212-5p down-regulation. After down-regulating miR-212-5p, the proliferation of HCT116 cells was significantly decreased, and the migration and invasion ability of HCT116 cells were markedly declined. In addition, the findings suggested that compared with the control, miR212-5p low-expression significantly decreased the protein expression level of N-cadherin in HCT116 cells, while the E-cadherin expression level remarkably increased. In conclusion, microRNA-212-5p down-regulation suppresses colorectal cancer migration and invasion by up-regulating SMAD4.

Keywords: MiR-212-5p, Smad4, target, proliferation, migration, invasion

Introduction

Colorectal cancer (CRC), which accounting for about 10% of the cancer mortalities every year, is one of the most common cancer in both men and women and a major cause of cancer-related deaths in the world [1-3]. About 1.4 million cases colorectal cancer are newly diagnosed and nearly 50% of the patients died from this disease every year [4]. Currently, the mainstay of anti-colorectal cancer treatment is surgery, chemotherapy and radiotherapy [5]. Although a lot of improvement has been made for surgical and medical treatments of colorectal cancer, there are still limitations of these treatments [6, 7]. Because of the recurrence and metastasis of tumor, the long-term survival and prognosis of patients with colorectal cancer is still very poor [8, 9]. Therefore, it is significantly and urgently to investigate and identify new and effective therapy targets for the treatment of colorectal cancer.

MicroRNAs (miRNAs) are small (19-25 nucleotides in length) non coding RNAs, widely exists in eukaryotic cells, completely or partially base bind to the 3' untranslated regions (UTR) of their multiple target mRNAs, leading to the degradation or translation inhibition of their target mRNAs, and post-transcriptionally regulate gene expression [10, 11]. Studies have revealed that miRNAs involved in the development of various cancers, including the colorectal cancer [12-15], and a large amount of evidence indicated that the abnormal expression of miRNAs is associated with the occurrence and develop-

Table 1. Primer sequence for PCR

Sequence (5'-3')

Smad4-Forward: 5'GACAGTGTCTGTGAATCC3' Smad4-Reverse: 5'TACTTGGCGGGTGTTGGATG3' MiR-212-5p-Forward: 5'CGCTAACAGTCTCCAGTC3' MiR-212-5p-Reverse: 5'GTGCAGGGTCCGAGGT3' U6-Forward: 5'GCTTCGGCAGCACATATACTAAAAT3' U6-Reverse: 5'CGCTTCACGAATTTGCGTGTCAT3' β-actin-Forward: 5'GGAGTCCTGTGGCATCCACG3' β-actin-Reverse: 5'CTAGAAGCATTTGCGGTGGA3'

ment of colorectal cancer by regulating the expression of their target gene including oncogenes and tumor inhibitor genes [14-18].

To the best of our knowledge, the expression and exact roles of miR-212-5p in CRC and the underlying molecular mechanism is still unclear. Therefore, in the present study, we will investigate the expression and the role of miR-212-5p in colorectal cancer and further explore the underlying molecular mechanism.

Materials and methods

Tissue samples

The study was approved by the Human Ethics Committees Review Board at Nanjing Drum Tower Hospital, The Affiliated Hospital of Nanjing University Medical School, Nanjing, China and informed consent was obtained from all patients. 20 tumor tissues from patients diagnosed with colorectal cancer at Nanjing Drum Tower Hospital between 2014 and 2015 and 20 paired adjacent non-tumoural tissues were collected and used for miRNA-212-5p expression detection. Native tumor and paired non-tumoural colon tissues were obtained after surgical resection and immediately stored in -80°C.

Cell culture

Human colorectal cancer cell line HCT116, human normal colon epithelial cell line FHC and human embryonic kidney cell line 293T were purchased from American type culture collection (ATCC, USA). HCT116 cells were grown in RPMI-1640 medium and 293T cells were grown in high glucose DMEM (Gibco, USA) medium supplemented with 10% fetal bovine serum (FBS) (Gibco, USA), 1% streptomycin-penicillin solution, and cells were incubated in a humidi-

fied atmosphere with 5% CO₂ at 37°C. Cells were passaged every 2-3 days. FHC cells were grown in DMEM/F12 medium (Gibco, USA) containing 10% FBS and 1% streptomycin-penicillin solution at 37°C with 5% CO₂.

Dual luciferase reporter assay

To determine whether miR-212-5p directly targets the 3'UTRs of Smad4, the vectors named SMAD4-3'UTR-WT and SMAD4-3'UTR-MUT with wild-type and mutated 3'UTR of SMAD4 mRNA were established as previously described [19]. 293T cells were seeded in a 24-well plate and then co-transfected with SMAD4-3'UTR-WT or SMAD4-3'UTR-MUT and miR-212-5p or its negative control (hsamiR-NC) vector using Lipofectamine 2000 transfection reagent in accordance with the manufacturer's instructions. 48 h after transfection, the Dual-Luciferase Reporter Assay Kit (Promega, USA) was performed to measure the luciferase activity according to the manufacturer's instructions.

Cell transfection

The day before transfection, human colorectal cancer HCT116 cells were seeded in 6-well plates at a density of 3×10^5 per well. When cells reached 50%-60% confluence, miR-212-5p inhibitor and its negative control were transfected into HCT116 cells with 50 μ l Lipofectamine 2000 transfection reagent (Invitrogen, USA) following the manufacturer's instructions. Cells without treatments act as the control. After incubating for 4 h, fresh culture medium was replaced. The transfected cells were used for following experiments analysis after incubating for another 24 h.

Cell proliferation assay

Cell proliferation ability of HCT116 cells was determined by performing MTT assay. 24 h after transfection, Log-phase human colorectal cancer HCT116 cells were collected using 0.25% trypsin, then the cells were seeded in a 96-well plate with 1.5×10^4 cells per well. 20 μL MTT (5 g/L) was added into each well and then incubating for 4 h. After the culture medium was discarded, 200 μL DMSO was added into each well. The OD value at 490 nm was measured using a spectrophotometer. Experiments repeated in triplicate.

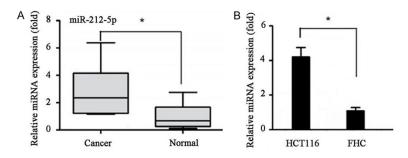


Figure 1. MiR-212-5p is up-regulated in CRC. QRT-PCR was performed to determine the miR-212-5p expression. A: Relative expression of miR-212-5p in tissues of CRC patients; B: Relative expression of miR-212-5p in CRC cells. Normal: the adjacent non-tumoural tissues of the CRC patients. Cancer: tissues of CRC patients; FHC: human normal colon epithelial cell line; HCT116: human colorectal cancer cell line. *P<0.05, experiments were performed in triplicate.

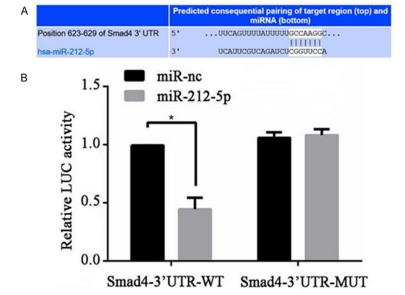


Figure 2. Smad4 is a direct target of miR-212-5p. A: Interaction between miR-212-5p and 3'UTR of Smad4 was predicted using TargetScan; B: Luciferase activity of a reporter containing a wild-type Smad4 3'UTR or a mutant Smad4 3'UTR are presented. "Smad4-3'UTR-MUT" indicates the Smad4 3'UTR with a mutation in the miR-212-5p binding site. UTR, untranslated region. *P<0.05 vs. control. All data are presented as the mean \pm SD of three independent experiments.

Cell migration and invasion assay

For cell migration ability detection, cell scratch test was performed. 24 h after transfection, HCT116 cells were seeded in a 6-well plate (5×10^5 per well) and incubated in a normal cell culture condition with 5% CO $_2$ at 37°C. When cells were in 80% confluence, wounds were formed by the 200-µl pin. After washing with PBS, cells were cultured in media without serum. Pictures were captured using an inverted microscope (Olympus, Japan).

For cell invasion ability detection, Transwell assay was performed. 24 h after transfection, cells were trypsinized and transferred to the upper chamber with matrigel-coated membrane matrix in serumfree medium. At the end of the tests, the non-invading cells on the upper membrane were struck off, and cells on the lower surface were fixed with 4% paraformaldehyde and stained with hematine. Cells were counted under microscope. Each experiment was independently performed three times.

Western blot

We performed Western blot analysis to detect the expression of proteins in CRC cell lines. Transfected HCT116 cells were lysed using RIPA buffer (CST, USA). Proteins were resolved by 10% SDS-PAGE and then transferred to a PVDF membrane. After blotted overnight with a primary antibody against Smad4, N-cadherin, E-cadherin (CST, USA; dilution ratio, 1:1000) or β-actin (CST, USA; dilution ratio, 1:2000), the blots were then incubated with a secondary antibody (CST, USA; dilution ratio, 1:5000) at room temperature for 2 h. Protein bands were visualized using Substrates (Millipore, ECL USA) and imaged and then analyzed.

ORT-PCR

Total RNAs were extracted from tissues and cells using Trizol reagent (Qiagen, USA) according to the manufacturer's protocol. β-actin or U6 was used as the internal control. The first-strand cDNA was generated by using the PrimeScript RT reagent Kit (TaKaRa, Japan) following the manufacturer's instructions. SYBR Premix Ex Taq (TaKaRa) was performed for real-time PCR analysis. The relative gene expression was calculated by using the Comparative

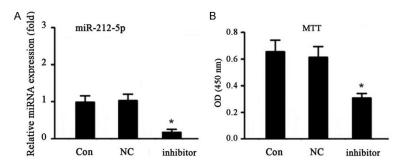


Figure 3. MiR-212-5p down-regulation inhibits the proliferation in HCT116 cells. A: Relative expression of miR-212-5p in HCT116 cells transfected with miR-212-5p inhibitor or NC, respectively; B: Cell proliferation was analyzed by MTT assay in the HCT116 cells. Con, control; NC, negative control. $^*P<0.05$ vs. Con. All results represent the means \pm SD of three independent experiments.

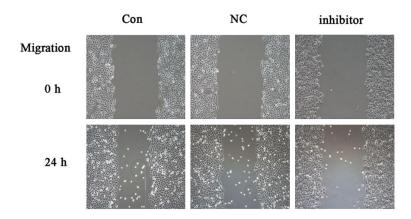


Figure 4. MiR-212-5p down-regulation inhibits the migration of HCT116 cells. The migration ability of the HCT116 cells was detected using cell scratch test at 24 h after the transfection of miR-212-5p inhibitor or NC, respectively. Con, control; NC, negative control. Experiments were performed in triplicate.

Ct method. Each experiment independently repeated for three times. The PCR primers for miR-212-5p and Smad4 were obtained from GenScript company (China) and shown in **Table 1**.

Statistical analysis

Data are presented as the mean \pm SD. We performed SPSS 17.0 statistical software (SPSS, Chicago, IL, United States) for all statistical analyses. T-test was used to evaluate the difference between groups. A value of P<0.05 was considered statistically significant.

Results

MiR-212-5p is up-regulated in human colorectal cancer

To investigate the role of miR-212-5p in the development of colorectal cancer, we first

detected miR-212-5p expression by qRT-PCR in the tumor tissues and adjacent nontumoural tissues of the CRC patients. At the same time, the expression level of miR-212-5p in the human normal colon epithelial cell line FHC and the human colorectal cancer cell line HCT116 was measured. The findings suggested that compared with the control, miR-212-5p significantly increased in human colorectal cancer tissues and the HCT-116 cells, indicating that miR-212-5p was up-regulated in CRC (Figure 1).

MiR-212-5p directly targets Smad4

In order to explore the mechanism of miR-212-5p function in CRC, TargetScan and Miranda databases was used to predict the target gene of miR-NA-212-5p, and Dual luciferase reporter Assay was performed to reveal our prediction. The miR-212-5p-Smad4-WT or miR-212-5p-Smad4-MUT reporter were co-transfected into 293T cells with miR-212-5p, and the results indicated that the luciferase

activity was significantly declined in miR-212-5p and miR-212-5p-Smad4-WT co-transfected 293T cells, but co-transfection of miR-212-5p and miR-212-5p-Smad4-MUT did not (**Figure 2**). The data revealed that miR-212-5p directly targets Smad4. In addition, the mRNA and protein expression of Smad4 were detected using qRT-PCR and western blotting. As shown in **Figure 6**, compared with the control, miR-212-5p low-expression markedly increased the expression of Smad4 at both mRNA and protein level. This once again proved Smad4 is a target gene of miR-212-5p.

MiR-212-5p low-expression inhibits the proliferation of HCT116 cells

To explore the effect of miR-212-5p on CRC cell proliferation, miR-212-5p was down-regulated using miR-212-5p inhibitor in HCT116 cells. We performed QRT-PCR to determine the expres-

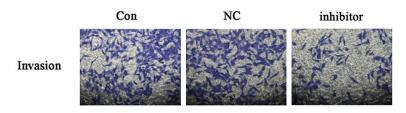


Figure 5. MiR-212-5p down-regulation inhibits the invasion of HCT116 cells. The invasion ability of the HCT116 cells was detected using transwell assay at 24 h after the transfection of miR-212-5p inhibitor or NC, respectively. Con, control; NC, negative control. Experiments were performed in triplicate.

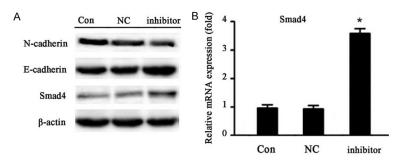


Figure 6. Expression level of Smad4, N-cadherin and E-cadherin in HCT116 cells. HCT116 cells were transfected with miR-212-5p inhibitor or NC, respectively. 24 h after the transfection, mRNA and protein expression of Smad4 were measured by QRT-PCR and western blotting. Protein expression level of N-cadherin and E-cadherin were measured by western blot analysis. A: Protein expression levels of Smad4, N-cadherin and E-cadherin in HCT116 cells; B: Relative mRNA expression levels of Smad4 in HCT116 cells. Con, control; NC, negative control. *P<0.05 vs. Con. All results represent the means ± SD of three independent experiments.

sion level of miR-212-5p and the findings suggested that miR-212-5p was effectively down-regulated when HCT116 cells were transfected with miR-212-5p inhibitor (**Figure 3A**).

Subsequently, the proliferation ability of HCT-116 cells was measured by CCK8 assay. Our results suggested that down-regulation of miR-212-5p significantly decreased the proliferation ability of HCT116 cells (Figure 3B). The data indicated that miR-212-5p down-regulation suppressed the proliferation of human colorectal cancer cells.

MiR-212-5p low-expression inhibits the migration and invasion of HCT116 cells

The impact of miR-212-5p on migration and invasion of HCT116 cells were evaluated in vitro, and cell scratch test and the transwell assay were performed. The results of the cell scratch test suggested that compared with the control, miR-212-5p low-expression significant-

ly declined the migration ability of HCT116 cells (Figure 4). We also found that HCT-116 cells transfected with miR-212-5p inhibitor invaded slower than those transfected with miR-NC. (Figure 5). The data indicated that miR-212-5p low-expression could suppress CRC cell migration and invasion.

MiR-212-5p low-expression affects the expression of N-cadherin and E-cadherin in HCT116 cells

Protein expression of the mesenchymal marker (N-cadherin) and the epithelial marker (E-cadherin) were also determined by western blotting in our study. As shown in **Figure 6**, compared with the control, the protein expression level of N-cadherin was significantly decreased when miR212-5p was low-expressed in HCT116 cells, and the E-cadherin expression level remarkably increased.

Discussion

In the present study, our results showed that miR-212-5p low-expression suppressed the proliferation, migration and invasion of colorectal cancer cells. MiR-212-5p directly targets Smad4 and miR-212-5p low-expression promotes its expression. MiR-212-5p low-expression thus indirectly inhibits the expression of N-cadherin and the E-cadherin expression level remarkably increased. First, we verified that miR-212-5p highly expressed in human colorectal cancer tissues and cells compared with the adjacent non-tumoural tissues and the human normal colon epithelial cell line FHC. We next revealed that miR-212-5p low-expression inhibited the proliferation, migration and invasion of colorectal cancer cells in vitro. Furthermore, we found a novel functional link between miR-212-5p and Smad4 in human colorectal cancer progress. These results indicated that miR-212-5p down-regulation plays essential roles in tumor suppression, thus, miR-212-5p may serve as a therapeutic target in the treatment of colorectal cancer.

A large amount of data indicate that miRNAs play essential roles in the diagnosis, therapy and prognosis of cancer, and they are associated with tumorigenesis and tumor metastasis [20-22]. Thus, miRNAs may be potential therapeutic targets for the treatment of cancer. A variety of studies have indicated that many miRNAs play important roles in the regulation of colorectal cancer cell proliferation, invasion and migration [19, 23-26]. However, to the best of our knowledge, the expression and the exact roles of miR-212-5p in human colorectal cancer have not been investigated. In our present study, the data strongly revealed that miR-212-5p expression was significantly up-regulated in both the tissues of CRC patients and human colorectal cancer HCT116 cells, and that miR-212-5p low-expression inhibited the proliferation, migration and invasion of HCT116 cells.

As a kind of tumor suppression genes, the genetic mutation, missing or negative expression of Smad4 can result in the deactivation, and play an important role in the development of tumor [27]. Studies have found that Smad4 play critical roles in inhibiting the metastasis of colorectal cancer cell [28]. In the process of Epithelial-mesenchymal transition (EMT), epithelial cells lose their polarity and cell-cell adhesion, and mesenchymal cells acquire migratory and invasive properties [29, 30]. Thus, EMT is essential for the initiation of cancer metastasis. In the present study, we found that miR-212-5p low-expression decreased the protein expression of mesenchymal marker N-cadherin but the protein expression of epithelial marker E-cadherin was increased, indicating that the inhibition of migration and invasion caused by miR-212-5p down-regulation was associated with its inhibitory effect on EMT. Multiple mechanisms are involved in regulating the progress of EMT and metastasis, including TGF-β1/SMAD pathways. Our results suggested that Smad4 is a target gene of miR-212-5p, and it can be proved by miR-212-5p down-regulation.

In conclusion, we proved for the first time that miR-212-5p was up-regulated in CRC, and its down-regulation can inhibit the proliferation, migration and invasion of colorectal cancer cells via directly targeting Smad4. MiR-212-5p

may play as a therapeutic target for CRC treatment.

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Disclosure of conflict of interest

None.

Address correspondence to: Chungen Xing and Tengfei He, Department of General Surgery, The Second Affiliated Hospital of Soochow University, 1055 Sanxiang Road, Suzhou 215000, China. E-mail: xcg295968@163.com (CGX); Tel: +86-512-67784109; Fax: +86-512-67784109; E-mail: hetf1789@163.com (TFH)

References

- [1] Al-Sheikh YA, Genie HK, Softa KI, Al-Jobran AA, Al-Obeed O, Mohamed MA, Abdulla M, Aboul-Soud MA. Expression profiling of selected microRNA signatures in plasma and tissues of Saudi colorectal cancer patients by qPCR. Oncol Lett 2016; 11: 1406-1412.
- [2] Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. CA Cancer J Clin 2011: 61: 69-90.
- [3] Markowitz SD, Bertagnolli MM. Molecular basis of colorectal cancer. N Engl J Med 2009; 361: 2449-2460.
- [4] Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. Cancer Incidence and Mortality Worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer 2015; 136: E359-386.
- [5] Shen A, Lin W, Chen Y, Liu L, Chen H, Zhuang Q, Lin J, Sferra TJ, Peng J. Pien Tze Huang inhibits metastasis of human colorectal carcinoma cells via modulation of TGF-β1/ZEB/miR-200 signaling network. Int J Oncol 2015; 46: 685-690.
- [6] O'Connell JB, Maggard MA, Ko CY. Colon cancer survival rates with the new American Joint Committee on Cancer sixth edition staging. J Natl Cancer Inst 2004; 96: 1420-1425.
- [7] Siegel R, Naishadham D, Jemal A. Cancer statistics, 2016. CA Cancer J Clin 2016; 66: 7-30.
- [8] Koyanagi K, Bilchik AJ, Saha S, Turner RR, Wiese D, McCarter M, Shen P, Deacon L,

- Elashoff D, Hoon DS. Prognostic relevance of occult nodal micrometastases and circulating tumor cells in colorectal cancer in a prospective multicenter trial. Clin Cancer Res 2008; 14: 7391-7396.
- [9] Bilchik AJ, Hoon DS, Saha S, Turner RR, Wiese D, DiNome M, Koyanagi K, McCarter M, Shen P, Iddings D, Chen SL, Gonzalez M, Elashoff D, Morton DL. Prognostic impact of micrometastases in colon cancer: interim results of a prospective multicenter trial. Ann Surg 2007; 246: 568-575.
- [10] Lee RC, Feinbaum RL, Ambros V. The C. elegans heterochronic gene lin-4 encodes small RNAs with antisense complementarity to lin-14. Cell 1993; 75: 843-854.
- [11] Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. Cell 2004; 116: 281-297.
- [12] Hu L, Ai J, Long H, Liu W, Wang X, Zuo Y, Li Y, Wu Q, Deng Y. Intergrative microRNA and gene profiling data analysis reveals novel biomarkers and mechanisms for lung cancer. Oncotarget 2016; 7: 8441-8454.
- [13] sai MM, Wang CS, Tsai CY, Huang HW, Chi HC, Lin YH, Lu PH, Lin KH. Potential diagnostic, prognostic and therapeutic targets of microR-NAs in human gastric cancer. Int J Mol Sci 2016; 17: 945.
- [14] Kaukoniemi KM, Rauhala HE, Scaravilli M, Latonen L, Annala M, Vessella RL, Nykter M, Tammela TL, Visakorpi T. Epigenetically altered miR-193b targets cyclin D1 in prostate cancer. Cancer Med 2015; 4: 1417-1425.
- [15] Calin GA, Croce CM. MicroRNA signatures in human cancers. Nat Rev Cancer 2006; 6: 857-866
- [16] Wang H, Wu J, Meng X, Ying X, Zuo Y, Liu R, Pan Z, Kang T, Huang W. MicroRNA-342 inhibits colorectal cancer cell proliferation and invasion by directly targeting DNA methyltransferase 1. Carcinogenesis 2011; 32: 1033-1042.
- [17] Ito M, Mitsuhashi K, Igarashi H, Nosho K, Naito T, Yoshii S, Takahashi H, Fujita M, Sukawa Y, Yamamoto E, Takahashi T, Adachi Y, Nojima M, Sasaki Y, Tokino T, Baba Y, Maruyama R, Suzuki H, Imai K, Yamamoto H, Shinomura Y. MicroRNA-31 expression in relation to BRAF mutation, CpG island methylation and colorectal continuum in serrated lesions. Int J Cancer 2014; 135: 2507-2515.
- [18] Zhou J, Zhang M, Huang Y, Feng L, Chen H, Hu Y, Chen H, Zhang K, Zheng L, Zheng S. MicroRNA-320b promotes colorectal cancer proliferation and invasion by competing with its homologous microRNA-320a. Cancer Lett 2015; 325: 669-675.
- [19] Zhang GJ, Zhou H, Xiao HX, Li Y, Zhou T. Upregulation of miR-224 promotes cancer cell proliferation and invasion and predicts relapse of colorectal cancer. Cancer Cell Int 2013; 13: 104.

- [20] Cho WC. MicroRNAs: Potential biomarkers for cancer diagnosis, prognosis and targets for therapy. Int J Biochem Cell Biol 2010; 42: 1273-1281.
- [21] Garzon R, Marcucci G. Potential of microRNAs for cancer diagnostics, prognostication and therapy. Curr Opin Oncol 2012; 24: 655-659.
- [22] Hayes J, Peruzzi PP, Lawler S. MicroRNAs in cancer: biomarkers, functions and therapy. Trends Mol Med 2014; 20: 460-469.
- [23] Ma Q, Wang X, Li Z, Li B, Ma F, Peng L, Zhang Y, Xu A, Jiang B. microRNA-16 represses colorectal cancer cell growth in vitro by regulating the p53/survivin signaling pathway. Oncol Rep 2014; 29: 1652-1658.
- [24] Zhang GJ, Xiao HX, Tian HP, Liu ZL, Xia SS, Zhou T. Upregulation of microRNA-155 promotes the migration and invasion of colorectal cancer cells through the regulation of claudin-1 expression. Int J Mol Med 2013; 31: 1375-1380.
- [25] Nosho K, Igarashi H, Nojima M, Ito M, Maruyama R, Yoshii S, Naito T, Sukawa Y, Mikami M, Sumioka W, Yamamoto E, Kurokawa S, Adachi Y, Takahashi H, Okuda H, Kusumi T, Hosokawa M, Fujita M, Hasegawa T, Okita K, Hirata K, Suzuki H, Yamamoto H, Shinomura Y. Association of microRNA-31 with BRAF mutation, colorectal cancer survival and serrated pathway. Carcinogenesis 2014; 35: 776-783.
- [26] Wang G, Cao X, Lai S, Luo X, Feng Y, Wu J, Ning Q, Xia X, Wang J, Gong J, Hu J. Altered p53 regulation of miR-148b and p55PIK contributes to tumor progression in colorectal cancer. Oncogene 2015; 34: 912-921.
- [27] Papageorgis P, Cheng K, Ozturk S, Gong Y, Lambert AW, Abdolmaleky HM, Zhou JR, Thiagalingam S. Smad4 inactivation promotes malignancy and drug resistance of colon cancer. Cancer Res 2011; 71: 998-1008.
- [28] Zhang B, Halder SK, Kashikar ND, Cho YJ, Datta A, Gorden DL, Datta PK. Antimetastatic role of Smad4 signaling in colorectal cancer. Gastroenterology 2010; 138: 969-980, e961e963
- [29] Pecina-Slaus N, Cicvara-Pecina T, Kafka A. Epithelial-tomesenchymal transition: possible role in meningiomas. Front Biosci (Elite Ed) 2012; 4: 889-896.
- [30] Hur K, Toiyama Y, Takahashi M, Balaguer F, Nagasaka T, Koike J, Hemmi H, Koi M, Boland CR, Goel A. MicroRNA-200c modulates epithelial-to-mesenchymal transition (EMT) in human colorectal cancer metastasis. Gut 2013; 62: 1315-1326.