Original Article Decreased expression of ARID1A is related to the poor prognosis of glioma patients

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Abstract: Purpose: This study intended to determine the expression of AT rich interactive domain 1A (SWI-like) (*ARID1A*) in serum of glioma patients and explore the relevance between *ARID1A* expression and the prognosis of glioma patients. Methods: The expression of *ARID1A* in serum of glioma patients and healthy controls were measured by high performance liquid chromatography (HPLC). Chi-square test was applied to evaluate the statistical difference between *ARID1A* expression and the clinicopathologic characteristics. Kaplan-Meier analysis combing with log-rank test was used to compare the overall survival of glioma patients with different *ARID1A* expression. Cox regression analysis was performed to evaluate the relationship between *ARID1A* expression and the prognosis of glioma patients. Results: The *ARID1A* expression was significantly lower in serum of glioma patients than in the healthy controls (P < 0.001). Moreover, the *ARID1A* expression was closely related to pathological grading, age and KPS score (P < 0.05), while no relationship was found between *ARID1A* expression and gender, preoperative epilepsy, or tumor range (P > 0.05). Besides, the overall survival time of patients with high *ARID1A* expression was significantly longer than those with low *ARID1A* expression according to Kaplan-Meier analysis (P = 0.005). Cox regression analysis illustrated that *ARID1A* expression was a potential factor for prognosis of glioma patients and it might be an independent biomarker (P = 0.002, HR = 4.992, 95% CI = 1.831-13.611). Conclusion: In a word, our study indicated that down-regulation of *ARID1A* was a promising biomarker for the prognosis of glioma patients.

Keywords: ARID1A, glioma, prognosis

Introduction

Tumor is one of the most severe threats for human beings among which glioma is distinct because of the unique micro-environment [1]. Glioma, which arises from glial cells, is one of the most frequent and most aggressive primary brain tumors in clinic [2, 3]. It accounts for 50%-60% of intracranial tumors and has the five-year survival rate only 20%-30% [4-6]. It has been confirmed that the risk of glioma was influenced by many factors such as hereditary disorders and exposure to high doses of ionizing radiation [7, 8]. Currently, despite the therapies for glioma including surgery, radiotherapy and chemotherapy have progressed a lot, the prognosis of this disease is still poor [9]. Therefore, it has been considered a promising approach to find a novel prognostic biomarker for the therapy of glioma patients.

AT-rich interactive domain 1A (ARID1A) gene, which is also called BAF250a, p270, hOSA1 and SMARCF1, is located at the 1p36.11 region of chromosomes [10]. ARID1A had been proved to participate in the regulation of various cellular processes, such as development, differentiation, proliferation and chromatin remodeling [11-13]. Evidence has confirmed that mutations and deficient of ARID1A were frequently observed in a variety of tumors or cancers, including breast cancer, endometrioid ovarian carcinoma, gastric cancer and Barrett's esophagus carcinoma [14-17]. These observations indicate that ARID1A is a potential candidate tumor suppressor. In addition, previous reports have investigated the prognostic significance of ARID1A mutation in gastric cancer and found that decreased expression of the ARID1A gene was associated with poor prognosis in patients [18], and the same results were also found in



Figure 1. The expression of *ARID1A* in serum of glioma patients and normal controls was assayed by HPLC. *ARID1A* was significantly decreased in glioma patients in contrast to the normal controls (P < 0.001).

clear cell renal cell carcinoma [19]. Although the relationship between expression of *ARID1A* and clinicopathological variables and prognostic significance in some cancers have been reported, the role of *ARID1A* in glioma is still unclear.

The present study attempted to examine the expression of *ARID1A* in glioma patients and evaluate the prognostic value of *ARID1A*.

Materials and methods

Patients and specimens

A total of 83 patients who were pathologically diagnosed with glioma were selected from the Second Hospital of Hebei Medical University. Among the patients, there were 49 males and 34 females with the age range from 10 to 70 years. None of the patients had received radioor chemo- therapy before serum collection. In addition, another 46 healthy human serum specimens (the biochemical indicators and immune indexes were in the normal range) were collected and regarded as controls. The study obtained approval of the Ethic Committee of the Second Hospital of Hebei Medical University. All the participants were asked to sign the informed written consents in advance.

The serum of glioma patients and healthy controls were collected and put into EDTA collection tubes, then stored at -80°C for using. The clinicopathologic characteristics including age, gender, preoperative epilepsy, tumor range, KPS score, and pathological grading were recorded in a database. A 5-years follow-up was conducted and the information was gotten via a telephone or questionnaire. The overall survival period was defined from the day of diagnosis to the day of death. Patients who died from unexpected events or other diseases were excluded from our study.

High performance liquid chromatography analysis

The concentration of ARID1A in serum was determined by high performance liquid chromatography (HPLC). 5 mL 5% perchloric acid was added into 5 mL serum sample for protein precipitation. The residue was separated and centrifugated at 4000 r/min for 10 min. The supernatant was collected and the precipitation was dropped. Then the supernatant was separated and centrifugated at 4000 r/min for 10 min three times. The precipitum of each sample was respectively affiliated and dried at 40°C under nitrogen stream. Then the sediments were reconstituted with 50 µl methyl cyanide and 20 µl of the solution was used for sample injection. HPLC was carried out with C18 column $(12 \times 4 \text{ mm}, 5 \mu\text{m})$ at room temperature and the column temperature was 30°C. The mobile phase consisted of ultrapure water, methyl cyanide and trifluoroacetic acid (500:400:0.5). The flow rate was set at 1.0 mL/min, and the detection wavelength was 282 nm. 0.01 g BAF250a was weighted and diluted in a 50 mL volumetirc flask for standard curve.

Statistical analysis

All the data were carried out by SPSS 18.0 software (SPSS Inc, IL, USA). The difference of *ARID1A* expression between glioma patients and healthy controls was analyzed through Students't test. The statistical significance of *ARID1A* expression and clinicopathologic characteristics of glioma patients was evaluated by Chi-square test. Kaplan-Meier analysis was used to describe the overall survival rate of glioma patients with different *ARID1A* expression. Cox regression analysis was adopted to evaluate the correlation between *ARID1A* expression and the prognosis of glioma patients. *P* < 0.05 was considered to be statistically significant.

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Characteristics	Case(n)	ARID1A Expression		v ²	Pvalue
Undractenstics	003C (II)	Low	High	Ā	i value
Gender			8	0.042	0.838
Male	49	37	12		
Female	34	25	9		
Preoperative epilepsy				0.669	0.413
Yes	38	30	8		
No	45	32	13		
Tumor range				0.478	0.489
Single lobe of brain	46	33	13		
Multiple lobe of brain	37	29	8		
KPS score				4.371	0.037
< 80	44	37	7		
≥80	39	25	14		
Age				5.552	0.018
≤ 50	46	39	7		
> 50	37	23	14		
Pathological grading				4.490	0.034
I, II	35	22	13		
III, IV	48	40	8		

Table 1. Statistical difference in ARID1A expression of
patients with various clinical features

Results

Low expression of ARID1A was found in glioma patients

The *ARID1A* expression was examined in 83 glioma serum specimens and 46 healthy human serum samples by HPLC. The concentration of *ARID1A* in serum of glioma patients was 44.98 ± 11.87 (mean \pm SD), while that in the healthy human serum was 90.75 ± 12.89 (mean \pm SD). A significant decrease of *ARID1A* expression was found in the glioma samples compared to the controls, indicating that *ARID1A* might be a tumor suppressor in glioma patients (**Figure 1**, *P* < 0.001).

Relevance of ARID1A expression and the clinicopathologic characteristics of glioma patients

In order to elucidate the clinical significance of *ARID1A* in glioma, we estimated the association between *ARID1A* expression and the clinicopathologic characteristics. The specimens were divided into two groups manually with the median expression of *ARID1A* in glioma patients: the patients with an *ARID1A* expression

sion of no less than 51.01 µg/ml were attributed to the high *ARID1A* expression group while the others were belonged to the low *ARID1A* expression group. The result revealed that *ARID1A* expression level was correlated with age (P = 0.018), KPS score (P = 0.037), and pathological grading (P = 0.034), but shared no significant relationship with gender, preoperative epilepsy and tumor range (P > 0.05), as shown in **Table 1**. This might indicate that the *ARID1A* expression was related to the development of glioma.

Correlation between ARID1A expression and the overall survival of glioma patients

During the follow-up, 61.3% (38 out of 62) cases died in the low ARID1A expression group, and 23.8% (5 out of 21) patients died in the high ARID1A expression group. Kaplan-Meier analysis manifested that patients with low ARID1A expression had shorter overall survival time than those with high ARID1A expression confirmed that ARID1A acted as an independent prognostic factor for glioma patients (Table 2, P = 0.002, HR = 4.992, 95% CI = 1.831-13.611).

Discussion

Gliomas represent a series of low and high grade brain tumors that belong to the central nervous system [20]. These tumors are classified as grade I-IV by the World Health Organization (WHO) according to the histology and morphological characteristics [21]. In addition, it has been confirmed that sophisticated gene interactions and molecular modulations were involved in the development of glioma. Glioma also has a highly invasive rate which makes the complete resection difficult and lead to a poor prognosis [22]. Therefore, we aimed to find a candidate biomolecular for prognosis of glioma patients and expected to provide a new therapy for this disease.

ARID1A encode a large nuclear protein which can interact with other proteins and form a switch/sucrose nonfermentable (SWI/SNF)



Figure 2. Kaplan-Meier analysis was made to evaluate the overall survival rate of glioma patients. Low *ARID1A* expression appeared to be correlated with unfavorable overall survival rate of glioma patients (P = 0.005). The *P* value was determined by log-rank test.

 Table 2. Multivariate analysis for the prognostic factors in the patients with glioma

Characteristics	P value	HR	95% CI
Condor	0.424	0.724	0 227 1 500
Genuer	0.424	0.724	0.327-1.399
Age	0.636	1.184	0.588-2.387
Pathological grading	0.921	1.044	0.446-2.446
ARID1A expression	0.002	4.992	1.831-13.611

chromatin remodeling complex [11, 23]. And it has been repeatedly reported to be mutated in various cancers as chromatin remodeling complexes. Loss or decreased expression of *ARID1A* is often found in a variety of cancers [18, 19], which has been proved to significantly relate to cancer recurrence and tumor aggravation. For instance, Ozawa Y et al. found *ARID1A* was decreased in esophageal squamous cell carcinoma and was a positive factor to the infiltrative growth of this tumor [24]. Itamochi H et al. investigated the effects of *ARID1A* and confirmed its down-regulation contributed to a poor prognosis in the stage I/II clear cell carcinoma of the ovary [25]. In the study of Cho H et al. the decrease of ARID1A participated in the tumor progression and predicted the prognosis of cervical cancer [26]. Besides, ARID1A also a prognostic factor in gastric cancer, breast cancer, and colorectal cancer because of its decreased or deleted expression in these cancers [18, 27, 28]. According to previous studies, ARID1A has often emerged as a novel tumor suppressor. However, its role in glioma had never been reported.

In the present study, we detected the expression of *ARID1A* in serum of glioma patients and found it was significantly decreased in the patients. Our finding was in agreement with a series of researches that *ARID1A* expression was frequently decreased or lost in a variety of cancers and indicated *ARID1A* might be a tumor suppressor in glioma. Then in order to estimate whether *ARID1A* was

involved in the development of glioma, the relationship between its expression and clinicopathologic characteristics of glioma patients was analyzed. The result demonstrated that *ARID1A* expression was associated with the KPS score, age and pathological grading. So we inferred that *ARID1A* might participate in the progression of glioma development.

As a variety of studies all confirmed that the *ARID1A* expression was linked with the prognosis of cancers, we speculated that it was also associated with the prognosis of glioma. As shown in this study, Kaplan-Meier analysis verified a significant correlation between the decrease of *ARID1A* expression and overall survival of glioma patients. The results revealed that the overall survival time of patients with high *ARID1A* expression lived longer than those with low *ARID1A* expression. Next, Cox regression analysis further confirmed that the association between the expression level of *ARID1A* and the prognosis of glioma. The outcome proved that *ARID1A* expression could impact

the prognosis and might be an independent prognostic indicator for glioma patients. However, the mechanism of this effect of *ARID1A* on glioma has not been fully understood. Some researchers had considered nonsense or deletion mutation of *ARID1A*, the alterations of PI3K-AKT pathway, and the p53 expression were all relative factors for the function mechanism of *ARID1A* in several cancers [29-32]. Therefore, we conjectured that loss or decreased expression of *ARID1A* in glioma was correlated with PI3K-AKT or p53 pathways, and this deduction needs to be further investigated in the future studies.

In conclusion, the current study confirmed that the *ARID1A* expression was down-regulated in glioma and explained the clinical significance of *ARID1A*. What's more, abnormal expression of *ARID1A* was proved to be an independent prognostic marker and correlated with unfavorable prognosis in glioma patients.

Disclosure of conflict of interest

None.

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References

- [1] Ruan S, Yuan M, Zhang L, Hu G, Chen J, Cun X, Zhang Q, Yang Y, He Q and Gao H. Tumor microenvironment sensitive doxorubicin delivery and release to glioma using angiopep-2 decorated gold nanoparticles. Biomaterials 2015; 37: 425-435.
- [2] Sun G, Shi L, Li M, Jiang N, Fu L and Guo J. Lefty inhibits glioma growth by suppressing Nodal-activated Smad and ERK1/2 pathways. J Neurol Sci 2014; 347: 137-142.
- [3] Huasong G, Zongmei D, Jianfeng H, Xiaojun Q, Jun G, Sun G, Donglin W and Jianhong Z. Serine protease inhibitor (SERPIN) B1 suppresses cell migration and invasion in glioma cells. Brain Res 2015; 1600: 59-69.
- [4] Ohgaki H and Kleihues P. Epidemiology and etiology of gliomas. Acta Neuropathol 2005; 109: 93-108.
- [5] Davis FG, Freels S, Grutsch J, Barlas S and Brem S. Survival rates in patients with primary malignant brain tumors stratified by patient age and tumor histological type: an analysis based on Surveillance, Epidemiology, and End

Results (SEER) data, 1973-1991. J Neurosurg 1998; 88: 1-10.

- [6] Jansen M, de Witt Hamer PC, Witmer AN, Troost D and van Noorden CJ. Current perspectives on antiangiogenesis strategies in the treatment of malignant gliomas. Brain Res Brain Res Rev 2004; 45: 143-163.
- [7] Reuss D and von Deimling A. Hereditary tumor syndromes and gliomas. Recent Results Cancer Res 2009; 171: 83-102.
- [8] Hocking B. Occupational exposure to ionizing and non-ionizing radiation and risk of glioma. Occup Med (Lond) 2008; 58: 148-149; author reply 149.
- [9] Butowski NA, Sneed PK and Chang SM. Diagnosis and treatment of recurrent highgrade astrocytoma. J Clin Oncol 2006; 24: 1273-1280.
- [10] Zhang X, Sun Q, Shan M, Niu M, Liu T, Xia B, Liang X, Wei W, Sun S, Zhang Y, Liu XS, Song Q, Yang Y, Ma Y, Liu Y, Yang L, Ren Y, Zhang G and Pang D. Promoter hypermethylation of ARID1A gene is responsible for its low mRNA expression in many invasive breast cancers. PLoS One 2013; 8: e53931.
- [11] Ho L and Crabtree GR. Chromatin remodelling during development. Nature 2010; 463: 474-484.
- [12] Wilson BG and Roberts CW. SWI/SNF nucleosome remodellers and cancer. Nat Rev Cancer 2011; 11: 481-492.
- [13] Weissman B and Knudsen KE. Hijacking the chromatin remodeling machinery: impact of SWI/SNF perturbations in cancer. Cancer Res 2009; 69: 8223-8230.
- [14] Decristofaro MF, Betz BL, Rorie CJ, Reisman DN, Wang W and Weissman BE. Characterization of SWI/SNF protein expression in human breast cancer cell lines and other malignancies. J Cell Physiol 2001; 186: 136-145.
- [15] Jones S, Wang TL, Shih le M, Mao TL, Nakayama K, Roden R, Glas R, Slamon D, Diaz LA Jr, Vogelstein B, Kinzler KW, Velculescu VE and Papadopoulos N. Frequent mutations of chromatin remodeling gene ARID1A in ovarian clear cell carcinoma. Science 2010; 330: 228-231.
- [16] Wang K, Kan J, Yuen ST, Shi ST, Chu KM, Law S, Chan TL, Kan Z, Chan AS, Tsui WY, Lee SP, Ho SL, Chan AK, Cheng GH, Roberts PC, Rejto PA, Gibson NW, Pocalyko DJ, Mao M, Xu J and Leung SY. Exome sequencing identifies frequent mutation of ARID1A in molecular subtypes of gastric cancer. Nat Genet 2011; 43: 1219-1223.
- [17] Streppel MM, Lata S, DelaBastide M, Montgomery EA, Wang JS, Canto MI, Macgregor-Das AM, Pai S, Morsink FH, Offerhaus GJ, Antoniou E, Maitra A and

McCombie WR. Next-generation sequencing of endoscopic biopsies identifies ARID1A as a tumor-suppressor gene in Barrett's esophagus. Oncogene 2014; 33: 347-357.

- [18] Wang DD, Chen YB, Pan K, Wang W, Chen SP, Chen JG, Zhao JJ, Lv L, Pan QZ, Li YQ, Wang QJ, Huang LX, Ke ML, He J and Xia JC. Decreased expression of the ARID1A gene is associated with poor prognosis in primary gastric cancer. PLoS One 2012; 7: e40364.
- [19] Park JH, Lee C, Suh JH, Chae JY, Kim HW and Moon KC. Decreased ARID1A expression correlates with poor prognosis of clear cell renal cell carcinoma. Hum Pathol 2015; 46: 454-460.
- [20] Hua C, Zhao G, Li Y and Bie L. Minichromosome Maintenance (MCM) Family as potential diagnostic and prognostic tumor markers for human gliomas. BMC Cancer 2014; 14: 526.
- [21] Nikas JB. A mathematical model for short-term vs. long-term survival in patients with glioma. Am J Cancer Res 2014; 4: 862-873.
- [22] Bellail AC, Hunter SB, Brat DJ, Tan C and Van Meir EG. Microregional extracellular matrix heterogeneity in brain modulates glioma cell invasion. Int J Biochem Cell Biol 2004; 36: 1046-1069.
- [23] Wang X, Nagl NG, Wilsker D, Van Scoy M, Pacchione S, Yaciuk P, Dallas PB and Moran E. Two related ARID family proteins are alternative subunits of human SWI/SNF complexes. Biochem J 2004; 383: 319-325.
- [24] Ozawa Y, Nakamura Y, Fujishima F, Felizola SJ, Takeda K, Okamoto H, Ito K, Ishida H, Konno T, Kamei T, Ohuchi N and Sasano H. Decreased Expression of ARID1A Contributes to Infiltrative Growth of Esophageal Squamous Cell Carcinoma. Tohoku J Exp Med 2015; 235: 185-191.
- [25] Itamochi H, Oumi N, Oishi T, Shoji T, Fujiwara H, Sugiyama T, Suzuki M, Kigawa J and Harada T. Loss of ARID1A expression is associated with poor prognosis in patients with stage I/II clear cell carcinoma of the ovary. Int J Clin Oncol 2015; [Epub ahead of print].

- [26] Cho H, Kim JS, Chung H, Perry C, Lee H and Kim JH. Loss of ARID1A/BAF250a expression is linked to tumor progression and adverse prognosis in cervical cancer. Hum Pathol 2013; 44: 1365-1374.
- [27] Zhao J, Liu C and Zhao Z. ARID1A: a potential prognostic factor for breast cancer. Tumour Biol 2014; 35: 4813-4819.
- [28] Wei XL, Wang DS, Xi SY, Wu WJ, Chen DL, Zeng ZL, Wang RY, Huang YX, Jin Y, Wang F, Qiu MZ, Luo HY, Zhang DS and Xu RH. Clinicopathologic and prognostic relevance of ARID1A protein loss in colorectal cancer. World J Gastroenterol 2014; 20: 18404-18412.
- [29] Maeda D, Mao TL, Fukayama M, Nakagawa S, Yano T, Taketani Y and Shih le M. Clinicopathological significance of loss of ARID1A immunoreactivity in ovarian clear cell carcinoma. Int J Mol Sci 2010; 11: 5120-5128.
- [30] Bosse T, ter Haar NT, Seeber LM, v Diest PJ, Hes FJ, Vasen HF, Nout RA, Creutzberg CL, Morreau H and Smit VT. Loss of ARID1A expression and its relationship with PI3K-Akt pathway alterations, TP53 and microsatellite instability in endometrial cancer. Mod Pathol 2013; 26: 1525-1535.
- [31] Samartzis EP, Noske A, Dedes KJ, Fink D and Imesch P. ARID1A mutations and PI3K/AKT pathway alterations in endometriosis and endometriosis-associated ovarian carcinomas. Int J Mol Sci 2013; 14: 18824-18849.
- [32] Allo G, Bernardini MQ, Wu RC, Shih le M, Kalloger S, Pollett A, Gilks CB and Clarke BA. ARID1A loss correlates with mismatch repair deficiency and intact p53 expression in highgrade endometrial carcinomas. Mod Pathol 2014; 27: 255-261.