

Original Article

Distribution characteristics of vesicular monoamine transporter in brain of mice with Parkinson disease

Xiang-Yang Tian^{1*}, Shu-Yan Li^{2*}, Xing-Zhen Sun³, Lin-Fang Chen¹, Bo Sun¹, Gui-Hua Ni¹, Wei-Dong Zhao¹

¹Departments of Neurology, Huai'an First People's Hospital, Nanjing Medical University, Huai'an 223300, China;

²Department of Ophthalmology, Huai'an First People's Hospital, Nanjing Medical University, Huai'an 223300, China; ³Department of Pediatrics, Huai'an First People's Hospital, Nanjing Medical University, Huai'an 223300, China. *Co-first authors.

Received July 10, 2015; Accepted August 22, 2015; Epub August 1, 2016; Published August 15, 2016

Abstract: Objective: To investigate the distribution of vesicular monoamine transporter (VMAT₂) in PD model of C57BL mice brain and their relationships with Parkinson disease. Methods: PD model of C57BL mice induced by reserpine or MPTP or reserpine plus MPTP was used respectively. The distribution of VMAT₂ and TH in the substantia nigra pars compacta (SNc), the striatum, ventral tegmental area (VTA) and locus coeruleus (LC) were examined by immunohistochemical staining. Results: The number of VMAT₂-positive neurons and the number of TH-positive neurons in SNc were significantly reduced in PD model than the control, but there was no great difference in VTA and LC between PD model and the control. VMAT₂-positive neuron fibers and TH-positive neuron fibers in the striatum region were significantly reduced in PD model. The distribution of VMAT₂ in the control in SNc was less than that in both VTA and LC. Conclusion: The amount of VMAT₂ that has protective effect on the DA neurons in SNc is less than that in both VTA and LC. The decrease of the protecting function of VMAT₂ in SNc may be the important cause of the selective DA neurons loss.

Keywords: Parkinson disease, VMAT₂, immunohistochemistry, tyrosine hydroxylase, C57BL mice

Introduction

Parkinson disease (PD) is a relatively common neurodegenerative disease in middle and elderly people with the characteristic pathological change of progressive degeneration and loss of dopaminergic (DA) neurons in the substantia nigra pars compacta in midbrain, but its pathogenesis has been still unknown yet. It was recently found that vesicular transport abnormality of neurotransmitter DA in nigrostriatal system plays an important role in the pathogenesis of PD, while vesicular monoamine transporter (VMAT₂) plays a key role in the transport of neurotransmitter DA [1]. VMAT₂ can bind 1-methyl-4-phenylpyridinium (MPP⁺), an active metabolite of neurotoxin 1-methyl-4-phenyl-tetrahydropyridine (MPTP), in the synaptic vesicles to protect DA neurons [2, 3]. In this study, distribution of VMAT₂ in mice model was observed to explore the role of VMAT₂ in pathogenesis of PD.

Materials and methods

Animal grouping

A total of 24 healthy male C57/BL brown mice (Supplied by Shanghai Laboratory Animal Center, Chinese Academy of Sciences) at the age of 8 to 10 weeks, weighing (20±3) g, were randomly divided into four groups, including MPTP group, reserpine group, reserpine + MPTP group and control group, with 6 in each group. Mice in the control group were intraperitoneally injected saline at the same volume for 7 consecutive days. The mice acted as usual after injection.

Establishment of MPTP-induced mice model of PD: MPTP (Sigma, USA) was dissolved in saline and intraperitoneally injected at 30 mg·kg⁻¹·24 h⁻¹ for 7 consecutive days. After administration, movement disorders, such as bradykinesia, tremor, piloerection and poor response to external stimuli, at varying degrees were observed in

all mice. With the increasing of MPTP dosage and duration of administration, movement disorders in mice aggravated and duration of symptoms increased. Except bradykinesia, symptoms completely disappeared after 24 hours.

Establishment of reserpine-induced mice model of PD: Reserpine (Shanghai Hongqi Pharmaceutical Factory) was dissolved in saline and intraperitoneally injected at $1 \text{ mg}\cdot\text{kg}^{-1}\cdot 24 \text{ h}^{-1}$ for 7 consecutive days. After administration, movement disorders, such as bradykinesia, tremor, piloerection and poor response to external stimuli, at varying degrees were observed in all mice. With the increasing of reserpine dosage and duration of administration, movement disorders in mice aggravated and duration of symptoms increased. Except bradykinesia, symptoms completely disappeared after 24 hours.

Establishment of reserpine + MPTP-induced mice model of PD: Reserpine was intraperitoneally injected at $1 \text{ mg}\cdot\text{kg}^{-1}\cdot 24 \text{ h}^{-1}$ firstly and 2 h later, MPTP was intraperitoneally injected at $30 \text{ mg}\cdot\text{kg}^{-1}\cdot 24 \text{ h}^{-1}$ for 7 consecutive days. After administration, the movement disorders occurred earlier and also lasted longer.

Sacrifice of mice and specimen collection

Three days after the last injection, mice were anesthetized with intraperitoneal injection of 2% pentobarbital, and heart was exposed through thoracotomy and washed with 50 ml of normal saline through left ventricle. 4% of paraformaldehyde-PB solution (0.1 mol/L, pH 7.4) at 4°C was infused into the heart for maintaining 30 min. Then, the whole brain was taken out rapidly through craniotomy and immersed into 4% paraformaldehyde at 4°C for 24 hours. Based on the mouse brain atlas, corpus striatum, substantia nigra, ventral tegmental area (VTA) and locus coeruleus (LC) were separated in sequence and their coronal sections were made, dehydrated, clearing, embedded in paraffin and sliced with $4 \mu\text{m}$ thickness.

VMAT₂ and TH immunohistochemistry (SP method) [4]

Slices were routinely deparaffinized. After thermal remediation, 3% H_2O_2 was used to block endogenous peroxidase for 10 min and normal

goat serum working solution was used to close for 30 min. Then, VMAT₂ polyclonal antibody (Chemico, USA) and TH monoclonal antibody (Sigma, USA) at a dilution of 1:100 were respectively dropwise added on the slices and stayed overnight in refrigerator at 4°C . The slices were eluted with PBS for 5 min \times 3 times, and secondary antibody (anti-rabbit/mouse IgG) working solution was added on the slices; the slices were incubated in incubator at 37°C for 60 min. The slices were eluted with PBS for 5 min \times 3 times, and horseradish peroxidase conjugated streptavidin working solution was added on the slices; the slices were incubated in incubator at 37°C for 60 min. The slices were eluted with PBS for 5 min \times 3 times, and were stained by using 0.04% 3,3-diaminobenzidine (DAB), stained contrastively by hematoxylin, dehydrated with alcohol, cleared by xylene and fixed with neutral gum. Observation and radiography were conducted under microscope. PBS was used to instead of primary antibody for negative control and other operations were the same as the above. Result was positive if cytoplasm and processes were tan.

Image analysis

LEICA color pathological image analysis system was conducted to analyze the number of VMAT₂ and TH positive cells in brain slices of substantia nigra pars compacta (SNc), VTA and LC during morphological image analysis and data processing. There were 3 brain slices for each part in mice. The counting was conducted under high-power microscope. All positive cells were counted. The total count of positive cell in 3 brain slices in each mouse was calculated to take an average. VMAT₂ and TH positive nerve fibers in the striatum area were determined using mean gray value. Meanwhile, mean gray value of positive VMAT₂ at each group was determined (Note: The greater the mean gray value was, the lower the intensity of immunostaining was and the lower the content of the material was).

Statistical analysis

Experimental data were expressed as mean \pm standard deviation ($\bar{x} \pm s$) and the comparison between groups was analyzed using analysis of variance. SPSS10.0 statistical software was employed and $P < 0.05$ was considered statistically significant.

Table 1. Comparison of the number of VMAT₂ positive cells in each group

Group	SNC	VTA	LC
Reserpine group	48.38±4.52*	130.35±14.67	87.32±9.48
MPTP group	32.64±5.26*.#	122.19±13.57	83.75±8.68
Reserpine + MPTP group	28.83±5.58*.#	118.72±13.45	81.21±8.56
Control group	79.36±8.76	131.24±14.36	86.63±9.14

When compared with control group, **P* < 0.05; when compared with reserpine group, #*P* < 0.05.

Table 2. Comparison of the number of TH positive cells in each group

Group	SNC	VTA	LC
Reserpine group	45.89±6.78*	133.36±13.67	92.37±9.87
MPTP group	30.42±4.41*.#	128.38±14.16	88.25±10.23
Reserpine + MPTP group	27.63±4.76*.#	122.64±13.87	84.31±9.18
Control group	84.45±9.23	136.58±14.29	90.34±9.68

When compared with control group, **P* < 0.05; when compared with reserpine group, #*P* < 0.05.

Table 3. Comparison of gray values for VMAT₂ and TH positive nerve fibers in the striatum area in each group

Group	VMAT ₂	TH
Reserpine group	160.23±5.81*	159.89±5.07*
MPTP group	170.63±4.84*.#	167.39±4.88*.#
Reserpine + MPTP group	172.89±4.78*.#	169.55±4.83*.#
Control group	146.25±4.72	142.13±5.05

When compared with control group, **P* < 0.05; when compared with reserpine group, #*P* < 0.05.

Table 4. Gray value for VMAT₂ in each group

Group	SNC	VTA	LC
Reserpine group	167.38±4.52*	133.35±4.67	122.32±4.48
MPTP group	175.49±5.26*.#	134.19±5.57	123.75±4.68
Reserpine + MPTP group	179.83±6.15*.#	135.72±5.45	124.21±5.26
Control group	155.36±5.76**	132.24±5.36	121.63±5.14

When compared with control group, **P* < 0.05; when compared with reserpine group, #*P* < 0.05; when compared with VTA and LC, ***P* < 0.05.

Results

Results of VMAT₂ immunohistochemical staining

As shown in **Table 1**, the number of VMAT₂ positive cells in SNC in experimental group significantly reduced when compared with that of control group (*P* < 0.05); the number of VMAT₂ positive cells in VTA and LC in experimental

group showed no significant reduction when compared with that of control group (*P* > 0.05); the comparisons among reserpine group, MPTP group and reserpine + MPTP group indicated that VMAT₂ in SNC reduced more significantly (*P* < 0.05).

Results of TH immunohistochemical staining

As shown in **Table 2**, the number of TH positive cells in SNC in experimental group significantly reduced when compared with that of control group (*P* < 0.05); the number of TH positive cells in VTA and LC in experimental group showed no significant reduction when compared with that of control group (*P* > 0.05); the comparisons among reserpine group, MPTP group and reserpine + MPTP group showed that TH in SNC reduced more significantly (*P* < 0.05).

Results of gray values for VMAT₂ and TH positive nerve fibers in the striatum area

As shown in **Table 3**, the difference of gray values for VMAT₂ and TH positive nerve fibers in the striatum area between experimental group and control group was significant (*P* < 0.05); moreover, the differences among reserpine group, MPTP group and reserpine + MPTP group were also significant (*P* < 0.05).

As shown in **Table 4**, the difference of gray value for VMAT₂ in SNC between experimental group and control group was significant (*P* < 0.05); while, the differences of gray value for

Results of gray value for VMAT₂ in SNC, VTA and LC

As shown in **Table 4**, the difference of gray value for VMAT₂ in SNC between experimental group and control group was significant (*P* < 0.05); while, the differences of gray value for

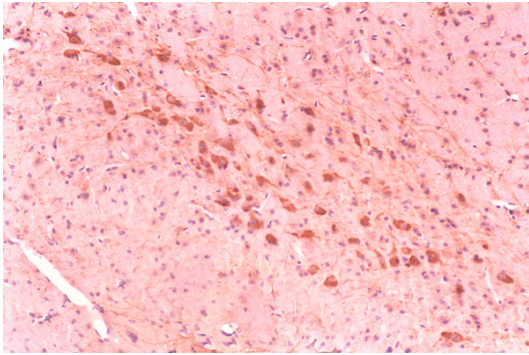


Figure 1. Expression of VMAT₂ in mice SNC in control group (Immunohistochemistry SP method, × 200).

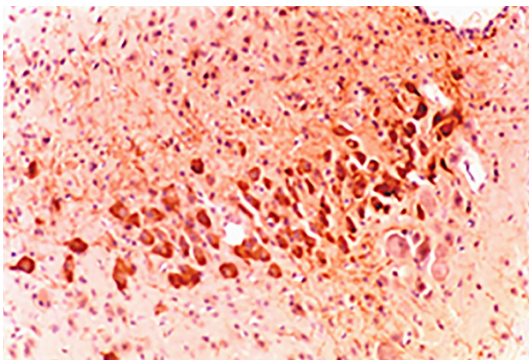


Figure 2. Expression of VMAT₂ in mice LC in control group (Immunohistochemistry SP method, × 200).

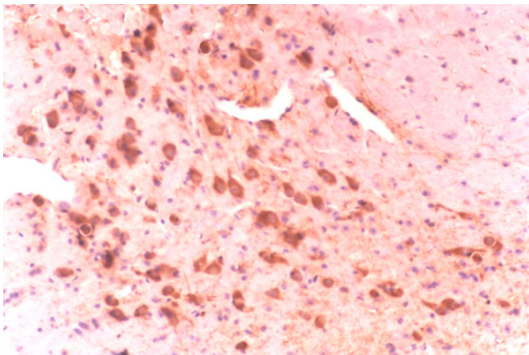


Figure 3. Expression of VMAT₂ in mice VTA in control group (Immunohistochemistry SP method, × 200).

VMAT₂ in VTA and LC between experimental group and control group was not significant ($P > 0.05$); the comparisons between reserpine group, MPTP group and reserpine + MPTP group indicated that VMAT₂ in SNC reduced more significantly ($P < 0.05$); moreover, it was found that the distribution of VMAT₂ in control

group in SNC was less than that in both VTA and LC ($P < 0.05$) (Figures 1-3).

Discussion

There are many theories about the pathogenesis of PD, such as environmental toxins, genetic susceptibility and neurotransmitter theories. But these theories can't provide convincing explanation on PD with selective DA neurons damage and the progressive course of the disease. MPTP is a potent and selective dopaminergic neurotoxin and it has been proved from a large number of studies that MPTP can cause a significant reduction of the content of DA in the striatum area and of the number of dopaminergic neurons in SNC. Neurotoxicity mechanism of MPTP has been demonstrated that MPTP is converted to the active metabolite MPP⁺ under the action of monoamine oxidase B and then MPP⁺ is actively transported to the mitochondria by the selective uptake of DA neurons, leading to degeneration and apoptosis of DA neurons [5-7]. Systemic administration of MPTP to monkeys and C57BL mice induces neurobiochemical and neuropathological changes, which are very similar to those in humans with PD. Therefore, MPTP model is widely accepted as the ideal animal model for the study of primary PD [8]. As an effective anti-hypertensive drug that produces side effects similar to depression, Reserpine is a specific and potent VMAT₂ inhibitor [9] that depletes catecholamines in the central and peripheral sympathetic nerve endings and competes with the monoamine neurotransmitters for the binding on the recognition site on vesicular transporter at a very low concentration, almost an irreversible binding, so as to effectively inhibit the vesicular transport of monoamine neurotransmitters. Administration of large dose of reserpine can cause a series of symptoms similar to paralysis agitans in laboratory animals, which, therefore, can be applied to establish the mice model of PD [10]. Pathological change of PD is a selective DA neurons damage in substantia nigra. Plasmalemma transporters in DA neurons with dopamine transporter (DAT), adrenergic neurons and serotonin neuron on all cell membranes can transport MPP⁺ into cells [11], but only DA neurons in substantia nigra exhibit sensitivity to MPP⁺. The reason is that VMAT₂ has the function of hiding the transport of MPP⁺ and other neurotoxins in cytoplasm

secretory vesicles, so that it can't produce toxic effect on cellular materials outside of vesicles. VMAT₂ plays a neuroprotective effect of different intensities in different nerve cells. The more the number of VMAT₂ is, the stronger the neuroprotective effect of VMAT₂ will be, and vice versa [12, 13]. Sai et al confirmed the anti-toxic effect of VMAT₂ [14]. VMAT₂ mainly locates at dopaminergic, noradrenergic, adrenergic, serotonergic, histaminergic neurons and corresponding neurons terminals in the central nervous system. SNC and VTA are the main focus of DA neurons and LC is one of the main focus parts of noradrenergic neurons, which has many important physiological functions in human. In the substantia nigra-striatum projection system, 95% of VMAT₂ positive fibers are dopaminergic and 5% are serotonergic [15, 16]. Dysfunctional VMAT₂ cannot effectively limit the damage of exogenous and endogenous toxicants on mitochondria, which can lead to the degradation of monoamine neurons [17]. Additionally, reduction of VMAT₂ in both SNC and striatum area in PD patients is observed [18].

In this study, distribution of VMAT₂ in control group in SNC was less than that in both VTA and LC and the result was consistent with that of foreign literature [19]. VMAT₂ positive neuron fibers in SNC and striatum area in experimental group significantly reduced than that in control group. Therefore, SNC was the most vulnerable region due to the weakest protective effect of VMAT₂. On the contrary, neurons, locating at VTA and LC with abundant number of VMAT₂, were hardly damaged. In TH staining, the number of TH positive cells in SNC in experimental group reduced significantly, while in VTA and LC did not significantly reduced. The result of the number of TH positive cells was consistent with the number of VMAT₂ positive cells in experimental group of mice, indicating that VMAT₂, like the number of TH positive cells, can be used as one of the indicators of pathological examination to determine whether PD animal model was established successfully or not. In this study, both behavioral indicator and pathological indicator in animal model of PD induced by reserpine combined with MPTP were significant than those induced by single reserpine or MPTP. It was analyzed that as a specific and potent inhibitor of VMAT₂, reserpine inhibited the protective effect of VMAT₂ and exacerbated the toxicity of intracellular MPP⁺ and other tox-

ins [20, 21]; therefore, this model can be used for PD study. In conclusion, it was shown from this study that the change in distribution of VMAT₂ in mice model of PD demonstrated that VMAT₂ had a protective effect on DA neurons and the distribution of VMAT₂ in SNC was less than that in VTA and LC. Meanwhile, the weak protective effect of VMAT₂ in SNC was an important cause for PD with selective damage in the substantia nigra.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Xing-Zhen Sun, Department of Pediatrics, Huai'an First People's Hospital, Nanjing Medical University, 6 Beijing West Road, Huai'an 223300, Jiangsu, China. Tel: +86-13861590017; E-mail: xingzhensuncn@yeah.net

References

- [1] Lohr KM, Bernstein AI, Stout KA, Dunn AR, Lazo CR, Alter SP, Wang M, Li Y, Fan X, Hess EJ, Yi H, Vecchio LM, Goldstein DS, Guillot TS, Salahpour A, Miller GW. Increased vesicular monoamine transporter enhances dopamine release and opposes Parkinson disease-related neurodegeneration in vivo. *Proc Natl Acad Sci U S A* 2014; 111: 9977-82.
- [2] Muñoz P, Paris I, Sanders LH, Greenamyre JT, Segura-Aguilar J. Overexpression of VMAT-2 and DT-diaphorase protects substantia nigra-derived cells against aminochrome neurotoxicity. *Biochim Biophys Acta* 2012; 1822: 1125-36.
- [3] Gros Y and Schuldiner S. Directed evolution reveals hidden properties of VMAT, a neurotransmitter transporter. *J Biol Chem* 2010; 285: 5076-84.
- [4] Anandhan A, Janakiraman U, Manivasagam T. Theaflavin ameliorates behavioral deficits, biochemical indices and monoamine transporters expression against subacute 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced mouse model of Parkinson's disease. *Neuroscience* 2012; 218: 257-67.
- [5] Ren Z, Yang N, Ji C, Zheng J, Zheng J, Wang T, Liu Y and Zuo P. Neuroprotective effects of 5-(4-hydroxy-3-dimethoxybenzylidene)-thiazolidinone in MPTP induced Parkinsonism model in mice. *Neuropharmacology* 2015; 93: 209-18.
- [6] Yu X, Yao JY, He J and Tian JW. Protection of MPTP-induced neuroinflammation and neurodegeneration by rotigotine-loaded microspheres. *Life Sci* 2015; 124: 136-43.

- [7] Feng G, Zhang Z, Bao Q, Zhang Z, Zhou L, Jiang J and Li S. Protective effect of chinoin in MPTP-induced C57BL/6 mouse model of Parkinson's disease. *Biol Pharm Bull* 2014; 37: 1301-7.
- [8] Takahashi-Niki K, Inafune A, Michitani N, Hatakeyama Y, Suzuki K, Sasaki M, Kitamura Y, Niki T, Iguchi-Arigo SM and Ariga H. DJ-1-dependent protective activity of DJ-1-binding compound no. 23 against neuronal cell death in MPTP-treated mouse model of Parkinson's disease. *J Pharmacol Sci* 2015; 127: 305-10.
- [9] Ashe KM, Chiu WL, Khalifa AM, Nicolas AN, Brown BL, De Martino RR, Alexander CP, Waggener CT, Fischer-Stenger K and Stewart JK. Vesicular monoamine transporter-1 (VMAT-1) mRNA and immunoreactive proteins in mouse brain. *Neuro Endocrinol Lett* 2011; 32: 253-8.
- [10] Leão AH, Sarmiento-Silva AJ, Ribeiro AM and Silva RH. Molecular, Neurochemical, and Behavioral Hallmarks of Reserpine as a Model for Parkinson's Disease: New Perspectives to a Long-Standing Model. *Brain Pathol* 2015; 10: 1111-9.
- [11] Buck KJ, Amara SG. Chimeric dopamine-norepinephrine transporters delineate structural domains influencing selectivity for catecholamine and 1-methyl-4-phenylpyridinium. *Proc Natl Acad Sci U S A* 1994; 91: 12584-8.
- [12] Mukda S, Vimolratana O, Govitrapong P. Melatonin attenuates the amphetamine-induced decrease in vesicular monoamine transporter-2 expression in postnatal rat striatum. *Neurosci Lett* 2011; 488: 154-7.
- [13] Choi SJ, Panhelainen A, Schmitz Y, Larsen KE, Kanter E, Wu M, Sulzer D and Mosharov EV. Changes in Neuronal Dopamine Homeostasis following 1-Methyl-4-phenylpyridinium (MPP+) Exposure. *J Biol Chem* 2015; 290: 6799-809.
- [14] Sai T, Uchida K, Nakayama H. Involvement of monoamine oxidase-B in the acute neurotoxicity of MPTP in embryonic and newborn mice. *Exp Toxicol Pathol* 2013; 65: 365-73.
- [15] Nunes EJ, Randall PA, Hart EE, Freeland C, Yohn SE, Baqi Y, Müller CE, López-Cruz L, Correa M and Salamone JD. Effort-related motivational effects of the VMAT-2 inhibitor tetrabenazine: implications for animal models of the motivational symptoms of depression. *J Neurosci* 2013; 33: 19120-30.
- [16] Anne C, Gasnier B. Vesicular neurotransmitter transporters: mechanistic aspects. *Curr Top Membr* 2014; 73: 149-74.
- [17] Li DW, Yao M, Dong YH, Tang MN, Chen W, Li GR and Sun BQ. Guanosine exerts neuroprotective effects by reversing mitochondrial dysfunction in a cellular model of Parkinson's disease. *Int J Mol Med* 2014; 34: 1358-64.
- [18] Segura-Aguilar J, Paris I, Muñoz P, Ferrari E, Zecca L and Zucca FA. Protective and toxic roles of dopamine in Parkinson's disease. *J Neurochem* 2014; 129: 898-915.
- [19] Smeyne M, Smeyne RJ. Glutathione metabolism and Parkinson's disease. *Free Radic Biol Med* 2013; 62: 13-25.
- [20] Duty S, Jenner P. Animal models of Parkinson's disease: a source of novel treatments and clues to the cause of the disease. *Br J Pharmacol* 2011; 164: 1357-91.
- [21] Hallett PJ, Brotchie JM. Striatal delta opioid receptor binding in experimental models of Parkinson's disease and dyskinesia. *Mov Disord* 2007; 22: 28-40.